



ISSN 2320-3862
JMPS 2014; 2(4): 15-18
© 2014 JMPS
Received: 02-06-2014
Accepted: 21-06-2014

Mohamed Ayad Berfad
M.Sc of Aquatic Biology – Marine
Biotechnology, Lecturer at the Faculty
of Arts and Sciences - Almergib
University

Tarig M. S. Alnour
Assistant professor,
Faculty of Medical Technology,
Mesellata, AlMergib University

Phytochemical analysis and Antibacterial activity of the 5 different extract from the seagrasses *Posidonia oceanica*

Mohamed Ayad Berfad, Tarig M. S. Alnour

ABSTRACT

This study was conducted in the faculty of Art and sciences and aimed to determine the chemical composition of the seagrass *Posidonia oceanica* using Ethyl acetate, Chloroform, Ethanol, Cyclohexan and Acetone and to determine the antibacterial activity of the crude extracts against: *Staphylococcus aureus*, *Salmonella typhi*, *Pseudomonas aeruginosa*, *Proteus mirabilis*, *Klebsiella pneumoniae* and *Escherichia coli*. The seagrass was collected, identified, washed, dried, grind and exposed to extraction by soxhlet apparatus, and the crude extracts was subjected to chemical reaction and agar disk diffusion method to detect the presence of the chemical compounds and antimicrobial susceptibility tests. *Posidonia oceanica* possess several chemical compounds including: alkaloids, flavanoides, phenols, tannins, saponins, phlobatannins, sterols, proteins, reducing sugars, polysaccharides, and resins which differ in concentration according to the solvent used and has potent antimicrobial activity against *Pseudomonas aeruginosa* and *Staphylococcus aureus* with variable activity against other used bacterial strains.

Keywords: Seagrass, *Posidonia oceanica*, Phytochemical analysis, Ethyl acetate, Chloroform, Ethanol, Cyclohexan and Acetone.

1. Introduction

Posidonia oceanica is seagrass able to grow in the clean water with a depth reaching -45 m, and is common plant in the White Mediterranean Sea ^[1]. *Posidonia oceanica* belongs to the family Posidoniaceae and its genus *Posidonia* has nine species; in which *Posidonia oceanica* is completely restricted to the White Mediterranean Sea ^[2]. The grasses used as indicator of good seawater quality as it's highly sensitive to marine pollutions ^[3]. Several researches stated the chemical composition of the species *Posidonia oceanica* and the role of this compound especially the phenolic compounds as biotic and abiotic resistant agents ^[3-5]. The ability of this compound to kills microbes allow us to look about the chemical activity of different extracts against several bacterial agents.

This study aimed to determine the chemical composition of the seagrass *Posidonia oceanica* using several chemical solvents including (Ethyl acetate, Chloroform, Ethanol, Cyclohexan and Acetone) and to determine the antibacterial activity of the extracts against bacteria including: *Staphylococcus aureus*, *Salmonella typhi*, *Pseudomonas aeruginosa*, *Proteus mirabilis*, *Klebsiella pneumoniae* and *Escherichia coli*.

2. Materials and Methods

2.1 Samples

Posidonia oceanica grasses were collected by scuba diving from 5 meters depth in September 2012, from the White Mediterranean Sea coast at Alkhums City – Libya. The grass was identified and classified in the Faculty of Arts and Sciences – AlMergib University – Libya. The seagrass was washed by tap water (to remove the salts) and then by distilled water and kept away from Sunrays with mixing from time to time for up to 48 hours. Finally, the grasses were dried by oven at 60 °C for 48 hours. The grasses were grinded with electric grinder till having very fine granules and kept at room temperature in dark bottle until used ^[6,7].

The bacterial strains were obtained from the Biotechnology Research Center in Tripoli.

2.2 Method of extraction

Crude extraction of the grasses was done using the soxhlet method by dissolving 10 g of the

Correspondence:
Mohamed Ayad Berfad
M.Sc of Aquatic Biology – Marine
Biotechnology, Lecturer at the
Faculty of Arts and Sciences -
Almergib University

powder in 200 ml sterile distilled water and passed through the soxhlet apparatus for 8 hours.

The crude extract, then subjected to several solvent including (Acetate, Chloroform, Ethanol, Cyclohexan and ethyl Acetate). Different solvent extracts were reacted with several chemical compounds (Table: 1) to determine the presence of alkaloids, flavanoides, phenols, tannins, saponins, phlobatannins, sterols, proteins, reducing sugars, polysaccharides, and resins. The presence of each compound was reported and the density was scored as 3+, 2+, 1+, ± or 0 according to the intensity of the colour.

Antimicrobial activity of the 5 solvent extracts were done using agar desk diffusion method; in brief 5 mm desks were impregnated in the solvent extract and allowed to dry in desiccant. The desks were placed on Muller and Hinton agar after spreading a swab impregnated in Muller and Hinton broth with 10⁸CFU/ml bacterial strains (for each stain single Petri-dish was used). The plate was examined after 24 hours and 48 hours for the presence of inhibition zone and the diameter of the zone was recorded.

Table 1: Different reactions used to determine the presence of some chemical composition of *Posidonia oceanica*

Detected chemicals	Reacted materials/ method	End point of the reaction
Tannins	1% Lead acetate	Appearance of gelatinous precipitate
	1% Ferric chloride	Appearance of bluish – green colour
Phlobotannins	Boiling + 1% HCL	Appearance of Red precipitate
Resins	95% Ethanol + 4% HCL	Turbidity
Saponins	Mercuric chloride	Appearance of white precipitate
Reducing substances (monosaccharide's)	Benedict's solution	Development of yellow – red colour
Flavonoides	Addition of diluted ammonia	Development of yellow colour
Phenolic compound	1% Ferric chloride + Folin's reagent	Appearance of bluish – green colour
Sterols	Chloroform + H ₂ SO ₄	Development of green, then red colour
Alkaloids	Mayer's	Appearance of white precipitate
Protein	Boiling + H ₂ SO ₄	Appearance of condensed white precipitate
Polysaccharides	Iodine	Blue colour

3. Results

Several chemical compounds were detected in the *Posidonia oceanica* using five different extraction solutions. The chemical composition of the *Posidonia oceanica* showed high concentration (i.e.: 3+) of phlobatannins obtained by using both acetone and ethanol extraction methods, abundant reducing substances by ethanol and ethyl acetate extraction methods, and sterols using acetone, chloroform and ethanol extracting agents (Table: 2). Alkaloids composition of the grasses was not detected by all extracting agents except the ethanol which showed weak reaction indicating few composition in the grass (Table: 2). Weak saponins

concentration was detected by acetone extraction method only (Table: 2).

Excellent antibacterial activity of the *Posidonia oceanica* was noted in the cyclohexane extract. The extract showed good inhibition against all tested organisms except *P. mirabilis*. Interestingly, the highly resistant *P. aeruginosa* had the biggest zone of inhibition (figure: 1). Ethyl acetate extract showed no antibacterial activity, while *Proteus mirabilis* on the other hands showed high resistant mechanisms (figure: 1). The most sensitive strain was *Staphylococcus aureus* which showed sensitivity zone to all but not the ethyl acetate extract (figure: 1).

Table 2: Phytochemical constituent of the *Posidonia oceanica* using different extracts

Chemical compound	Chemical extracts				
	Acetone	Chloroform	Cyclohexan	Ethanol	Ethyl acetate
Alkaloids	0	0	0	1+	0
Flavonoides	1+	2+	2+	1+	±
Phenols	0	0	0	1+	0
Phlobatannins	3+	2+	1+	3+	2+
Polysaccharides	1+	1+	1+	1+	1+
Proteins	1+	1+	1+	0	1+
Reducing sugars	1+	1+	1+	3+	3+
Resins	1+	2+	0	0	1+
Saponins	1+	0	0	0	0
Sterols	3+	3+	1+	3+	2+
Tannins	1+	1+	±	2+	1+

0= No reaction, ± = unclear results, += weak positive, 2+ = good reaction, 3+= strong reactions

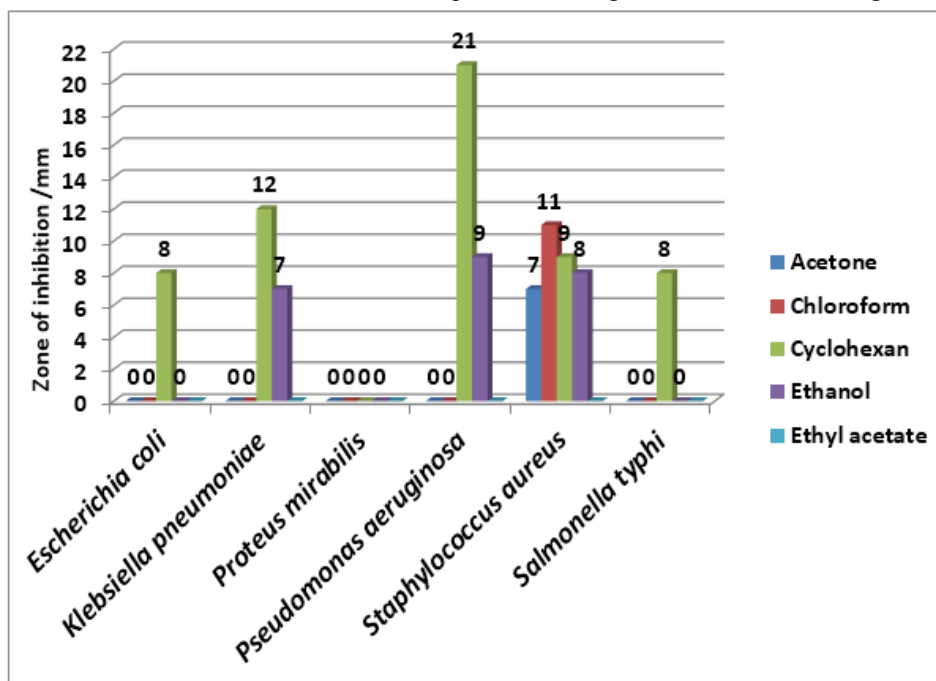


Fig 1: Antimicrobial susceptibility of the *Posidonia oceanica* different extracts

4. Discussion

Posidonia oceanica is an important White Mediterranean seagrass, as these grasses attenuate the wave streams, protect the hinterland from wave attack, and stabilize the seabed^[8]. They also use as indication of marine contamination as they are highly sensitive to marine pollutions^[3].

Several chemical compounds including (alkaloids, flavanoides, phenols, tannins, saponins, phlobatannins, sterols, proteins, monosaccharides, polysaccharides, and resins) have been detected in the seagrass *Posidonia oceanica*. Some of these have also been detected in similar seagrasses or other plants such as *Piper umbellatum* and *Piper pellucid*^[9], *Syringodium isoetifolium*^[10, 11], *Raphanus sativus niger*^[12], *Cymodocea serrulata*^[11, 13], *Halodule pinifolia*^[11], and *Halophila ovalis*^[11].

Acetone considered one of the best extracting agents able to extract 9/11 chemical compounds, while cyclohexan failed to extract 4/11 chemical compounds, although the uses of multiple extracting agents is better for good extraction of different chemical components. Several researcher describe different fatty acids, phenolic compounds and tannins content of the seagrass *Posidonia oceanica*^[3, 5, 14, 15], although our study detected small concentration of alkaloids, phenols and saponins, this might be due to differences in the extraction and/or detection methods.

Posidonia oceanica seagrass extracts showed good antimicrobial activity against several bacterial strains. This finding contradicts with the finding of Orhan I. *et al*^[16] and Ballesteros E. *et al*^[17] who showed no activity of *Posidonia oceanica* extracts against bacterial strains. This difference might be due to differences in the extraction methods or in the tested bacterial strains. Best antimicrobial activity was obtained by cyclohexan followed by ethanol extracts, while poor activity was noted from ethyl acetate. This difference may be contributed to ability of different chemical extraction method to extract chemical compounds with different concentration and may contributes to its antimicrobial activity against different bacterial strains by masking them or inhibiting their activity.

5. Conclusion

This study concluded that the seagrass *Posidonia oceanica* possess several chemical compounds including: alkaloids, flavanoides, phenols, tannins, saponins, phlobatannins, sterols, proteins, reducing sugars, polysaccharides, and resins which differ in concentration according to the solvent used and has potent antimicrobial activity against *Pseudomonas aeruginosa* and *Staphylococcus aureus* with variable activity against other used bacterial strains.

6. Conflict of interest

We, the authors of this manuscript certify that there was no conflict of interest to any private or Governmental institutes.

7. Acknowledgements

The authors are grateful to the Faculty of Arts and Science AlMergib University – Libya for laboratory facilities and the Biotechnology Research Center in Tripoli for supplying the bacterial strains.

8. References

- Romero J, Perez M, Alcoverro T, Ángel M, Mateo MA, Sanchez-Lizaso JL. Production ecology of *Posidonia oceanica* (L.) Delile meadows in Nueva Tabarca Marine Reserve: Growth, biomass and nutrient stocks along a bathymetric gradient. *Oecologia aquatic* 1998; 11:111-121.
- Larkum AWD. Taxonomy and biogeraphy of Seagrasses. in: *Seagrasses: Biology, Ecology and Conservation*. Edited by C. den Hartog, and John Kuo. Springer (Netherlands), 2006, 1-23.
- Cozza R, Chiappetta A, Petrarulo M, Salimonti A, Rende F, Bitonti MB, Innocenti AM. Cytophysiological features of *Posidonia oceanica* as putative markers of environmental conditions. *Chemistry and Ecology* 2004; 20(3):215-223.
- Haznedaroğlu MZ, Akarsu F. Anatomical Features of *Posidonia Oceanica* (L.) Delile Growing in Turkey. *Hacettepe University Journal of the Faculty of Pharmacy* 2009; 29(1):37-43.

5. Kersaoui O, Marzouki MN, Maugard T, Limam F. *In vitro* evaluation of antioxidant activities of free and bound phenolic compounds from *Posidonia oceanica* (L.) Delile Leaves. African Journal of Biotechnology 2011; 10(16):3176-3185.
6. Mohammed A, Al-Dulami M, Saour K. Physicochemical and partial purification of crude alkaloid compounds in berries, leaves and roots of (*Solanum nigrum*) plants. Scie Iraqi J 2009; 50(3):303-314.
7. Zineb S, Mohamed L, FAID Mohamed F, Khadija FZ. Inhibition of growth and mycotoxins formation in moulds by marine algae *Cystoseira tamariscifolia*. African Journal of Biotechnology 2004; 3(1):71-75.
8. Koftis T, Prinos P. Estimation of wave attenuation over *Posidonia Oceanica*. In Proceedings of the 5th international short conference on applied coastal research: 6th – 9th June, 2011 – RWTH Aachen University, Germany.
9. Mensah JK, Ihenyen JO, Okhiure MO. Nutritional, phytochemical and antimicrobial properties of two wild aromatic vegetables from Edo State. J Natural Products Plant Resour 2013; 3(1):8-14.
10. Mani AE, Aiyamperumal V, Patterson J. Phytochemicals of the Seagrass *Syringodium isoetifolium* and its Antibacterial and Insecticidal Activities. European Journal of Biological Sciences 2012; 4(3):63-67.
11. Qi SH, Zhang S, Qian PY, Wang BG. Antifeedant, antibacterial, and antilarval compounds from the South China Sea seagrass *Enhalus acoroides*. Botanica Marina 2008; 51:269-277.
12. Janjua S, Shahid M, Fakhir-i-Abbas. Phytochemical analysis and *in vitro* antibacterial activity of root peel extract of *Raphanus sativus* L. var niger. Advancement in Medicinal Plant Research 2013; 1(1):1-7.
13. Ravikumar S, Syed Ali M, Anandh P, Ajmalkhan M, Dhinakaraj M. Antibacterial activity of *Cymodocea serrulata* root extract against chosen poultry pathogens. Indian J Sci Technol 2011; 4(2):98-100.
14. Haznedaroglu MZ, Zeybek U. HPLC detection of chicoric acid in leaves of *Posidonia oceanica*. Pharmaceutical Biology 2007; 45(10):745-748.
15. Castellano G, Tena J, Torrens F. Classification of phenolic compounds by chemical structural indicators and its relation to antioxidant properties of *Posidonia oceanica* (L.) Delile. MATCH Commun. Math Comput Chem 2012; 67:231-250.
16. Orhan I, Wisespongpan P, Atici T, Sener B. Toxicity properties of some marine and fresh-water Algae as their chemical defense. Ankara Ecz Fak Derg 2003; 32(1):19-29.
17. Ballesteros E, Martin D, Uriz J. Biological activity of extracts from some Mediterranean Macrophytes. Botanica Marina 1992; 35:481-485.