

ISSN (E): 2320-3862
ISSN (P): 2394-0530
NAAS Rating: 3.53
JMPS 2018; 6(6): 79-83
© 2018 JMPS
Received: 11-09-2018
Accepted: 15-10-2018

Vijender Singh
School of Pharmacy, Sharda University, 32, 34 Knowledge Park-III, Greater Noida, Uttar Pradesh, India

Charu Pahuja
Ram-Eesh Institute of Vocational and Technical Education, 3, Knowledge Park I, Greater Noida, Uttar Pradesh, India.

Mohammed Ali
Phytochemistry Research Laboratory, School of Pharmaceutical Sciences and Research, Jamia Hamdard, New Delhi, India

Shahnaz Sultana
a) Phytochemistry Research Laboratory, School of Pharmaceutical Sciences and Research, Jamia Hamdard, New Delhi, India
b) College of Pharmacy, Jazan University, Jazan, Saudi Arabia

Analysis and bioactivities of essential oil of the flower buds of *Syzygium aromaticum* (L.) Merr. et LM Perry

Vijender Singh, Charu Pahuja, Mohammed Ali and Shahnaz Sultana

Abstract

Syzygium aromaticum L. (Myrtaceae), known as clove, is an aromatic dried flower bud grown in India and other tropical countries. It is aromatic, analgesic, antiseptic, spasmodic and stimulant and used to relieve cough, indigestion, toothache, arthritis, rheumatism and to manufacture perfumes, soaps and toothpastes. This research is principally focused on evaluating the effect of the flower bud on some acneogenic pathogens and anti-inflammatory and anti-oxidant activities. The GC-MS analysis of the clove bud essential oil showed the presence of eugenol (55.6%) as a major volatile constituents followed by eugenol acetate (12.5%), chavicol (6.5%), methyl acetate (5.9%), β -caryophyllene (5.2%) and α -caryophyllene (4.9%). The essential oil (1 %, v/v) exhibited significant antibacterial activity against *Propionibacterium acne* (17.8 mm) and *Staphylococcus epidermidis* (16.8 mm) using Clindamycin as a standard. The essential oil showed an excellent scavenging activity against DPPH radical relative to Vitamin C (standard) at $P<0.05$. Dose dependent anti-inflammatory response was observed with increasing concentration of the clove bud oil. However, the essential oil (1%, v/v) exhibited significant effect comparable results to that of Diclofenac taken as reference standard. These observations justify the use of clove for the treatment of acne and pimples due to its marked antimicrobial, anti-oxidant and anti-inflammatory activities.

Keywords: *Syzygium aromaticum*, flower buds, essential oil, anti-acne effect, anti-oxidant, anti-inflammatory activities

Introduction

Syzygium aromaticum (L.) Merr. et L.M. Perry, syn. *Caryophyllus aromaticus* L., *Eugenia caryophyllata* Thunb, *Jambosa caryophyllus* (Thunb.) Nied, *Myrtus caryophyllus* Spreng. (family Myrtaceae), known as clove and lavang, is grown in Indonesia, India, Malaysia, Sri Lanka, Madagascar, Tanzania and Brazil. It is an evergreen tree, up to 8–12 m tall, with large leaves and crimson flowers grouped in terminal clusters; flower buds have a pale hue initially, gradually turn green, then to a bright red when ready for harvest (Fig. 1). Cloves are used as an anodyne anthelmintic, antiseptic, aromatic, carminative, stimulant; used to treat arthritis, asthma, bronchitis, bruises, burns, cholera, colds, colic, coughs, diarrhoea, digestive disorders, earaches, gum diseases, headaches, hypertension, impotence, inflammation of the pharynx, intestinal worms, nausea, rheumatism, toothache, ulcers, vomiting and wounds^[1, 2]. Clove oil is used as an anodyne, and in aromatherapy to produce bath salt, soaps, and perfumes.



Fig 1: Clove buds and clove plant

Correspondence
Mohammed Ali
Phytochemistry Research Laboratory, School of Pharmaceutical Sciences and Research, Jamia Hamdard, New Delhi, India

The main constituents of the clove essential oil are phenyl propanoids such as eugenol, carvacrol, thymol, cinnamaldehyde, eugenol acetate, β -caryophyllene, α -humulene, β -pinene, limonene, farnesol, benzaldehyde, 2-heptanone and ethyl hexanoate^[1-10]. Clove contained phenolic acids viz., gallic, caffeic, ferulic, ellagic and salicylic acids, tannins, kaempferol, quercetin, its glycosides and sesquiterpenoids^[1, 2, 11, 12]. The buds yielded limonin, ferulic aldehyde, eugenol, tamarixetin 3-O- β -D-glucoside, ombuin 3-O- β -D-glucoside and quercetin^[4, 13].

The clove extracts and the isolated flavonoids showed strong antioxidant and phytotoxic and activities^[14 – 18]. An ethanol extract of clove exhibited remarkable hepatoprotective activity against paracetamol-induced liver injury in female rats^[13]. The clove extracts elicited antibacterial, anti-Herpes simplex, anti-hepatitis C viruses, anticarcinogenic, antifungal and antiviral activities^[2, 11, 19-25]. The clove essential oil possessed antiinflammatory, antimicrobial, antinociceptive, cytotoxic, insect repellent, insecticidal, fumigant, antioxidant and anaesthetic properties^[1, 2, 10, 26 – 35]. Clove oil reverses learning and memory deficits in scopolamine treated mice^[36]. The purpose of this work is to study chemical composition of the bud essential oil of *S. aromaticum* available in Delhi and to evaluate antimicrobial, anti-oxidant and anti-inflammatory activities of the clove extracts.

Experimental

Plant material

Dried unripe flower buds of *Syzygium aromaticum* were purchased from a local market, Khari Baoli, Delhi and authenticated by Dr. H. B Singh, Taxonomist, Division of Herbology, AIMIL Pharmaceuticals (I) Ltd, New Delhi. A voucher specimen is preserved in the herbarium of School of Pharmacy, Sharda University, Greater Noida, U.P.

Isolation

The clove buds (100 g) were steam distilled according to the method recommended in British Pharmacopoeia, 2009 (6). The dark green oil obtained was dried over anhydrous sodium sulphate and stored at 4°C in the dark. The yield was 1.1 % v/w based on fresh weight of sample.

GC analysis

The gas chromatographic analysis of the essential oil was carried out on a GC-2010 (Shimadzu) equipped with a flame ionization detector (FID) and QP-2000 fused silica capillary column (60 m × 0.25 mm × 0.25 μ m). The injector and detector (FID) temperatures were maintained at 250 and 270 °C, respectively. The carrier gas used was nitrogen at a flow rate of 1.21 ml/min with column pressure of 155.1 kPa. The sample (0.2 μ l) was injected into the column with a split ratio of 80:1. Component separation was achieved following a linear temperature programmed from 60 to 230 °C at a rate of 30C/min and then held at 230 °C for 9 min, with a total run time of 55.14 min. Percentage of the constituents were calculated by electronic integration of FID peak areas.

GC- MS analysis

The GC-MS analysis was carried out on a GC-MS-QP 2010 Plus (Shimadzu) fitted with a column AB-Innowax (60 m × 0.25 mm i.d., film thickness 0.25 μ m). The carrier gas was nitrogen at a flow rate 1.21 ml/min. The oven column temperature was initially kept at 60 °C for 10 min and increased up to 230 °C at a rate of 4 °C/min, then held at 230 °C for 10 min and increased up to 260 °C at a rate of 1 °C/min

and then held at 260 °C for 10 min. The split flow was 101 ml/min. The split ratio was 1:80. The injector temperature was 240 °C and detector temperature was 280 °C. Injection volume was 0.3 μ l. The ionization energy (voltage) was 70 eV and mass scan range (m/z) was 40-850 amu. The percentage composition of the oil was calculated automatically from the FID peak area without any correction.

Identification of compounds

The individual compounds were identified by comparing their Kovat's indices (KI) of the peaks on Innowax fused silica capillary column with literature values, matching against the standard library spectra, built up using pure substances and components of known essential oils. Further identification was carried out by comparison of fragmentation pattern of the mass spectra obtained by GC-MS analysis with those stored in the spectrometer database of NBS 54 K L, WILEY 8 libraries and published literature^[37, 38]. Relative amounts of identical components were based on peak areas obtained without FID response factor correction.

Anti-microbial activity

Alcoholic extraction of flower buds

The dried unripe flower buds of clove (100 g) were exhaustively extracted with ethyl alcohol (95%) in Soxhlet apparatus for 15 hrs. The extract was dried under reduced pressure to obtain a dark brown semisolid residue (9.5 g).

Preparation of standard drugs solution

Clindamycin was used as standard solutions for comparison of anti-bacterial studies. Both the standard drugs were taken in DMSO. The concentration of standard drug solutions was 0.10 mg. / ml.

Anti-microbial activity

The antibacterial activities of essential oil and alcoholic extract of clove buds were performed against *Propionibacterium acne* and *Staphylococcus epidermidis* in the Department of Microbiology, School of Pharmacy, Greater Noida.

Media

Nutrient agar media was composed of beef extract (1.0 g), yeast extract (2.0 g), peptone (5.0 g), sodium chloride (5.0 g), agar (15.0 g) and distilled water (1.0 L). Sabouraud dextrose agar media was composed of dextrose (40.0 g), mycological peptone (10.0 g), agar (15.0 g) and distilled water (1.0 L).

Preparation of media

Nutrient agar medium (28 g) was accurately weighed, suspended in 1000 ml of distilled water in a conical flask and heated to boiling to dissolve the medium completely. The conical flask containing the nutrient agar medium was plugged with a non-absorbent cotton plug and covered properly with an aluminum foil. It was sterilized by autoclaving at 15-lbs/in² pressure (121 °C) for 15 min. Sabouraud dextrose agar medium (65 g) was accurately weighed, suspended in 1000 ml of distilled water in a conical flask and heated to boiling to dissolve the medium completely. The conical flask containing the sabouraud dextrose agar medium was plugged with a non-absorbent cotton plug and covered properly with an aluminum foil. It was sterilized by autoclaving at 15-lbs/in² pressure (121 °C) for 15 min.

Preparation of organisms or inoculums The test organisms

were maintained on freshly prepared medium slants. The slants were incubated at 37°C for 24 h. The organisms from the medium slants were washed using 3 ml of saline solution and incubated for 24 h at 37 ± 20C. The developed organisms from the nutrient media were washed using 50 ml of distilled water. A dilution factor was determined which gave 25% light transmission at 530 nm. The amount of suspension to be added to each 100 ml nutrient broth was determined by use of test plates or test broth. The test organisms were stored under refrigeration.

The identification of microbial strains was based on morphological, cultural and biochemical tests. The microbes were procured from Institute of Microbial Technology, Chandigarh. The in-vitro antimicrobial activities of essential oil and dried alcoholic extract of the dried clove flower buds were studied by the cup plate method (14- 17) against various microorganisms mentioned in the Table-3. Clindamycin was used as standard and the activities of the essential oil and alcoholic extract were compared with corresponding concentration of standard drugs. The plates were incubated at 37 ± 2 °C for antibacterial activity, after 48 hrs of incubation. The Petri dishes were taken out from the incubator and the antimicrobial activity of essential oil and alcoholic extract of dried unripe flower buds of clove were compared by measuring the diameter of the zone of inhibition. (Table-4).

DPPH Radical scavenging activity

Preparation of aqueous extract

The air dried and coarsely powdered material (100g) was exhaustively extracted with distilled water in a reflux condenser for 4 -5 hrs. The extracts obtained were dried under reduced pressure to obtain a red brown colored residue (8.3g).

Preparation of essential oil concentrations

The essential oil (0.1% v/v, 0.5 % v/v, 1% v/v) and dried alcoholic extract (5.0% w/v) were dissolved in aqueous alcoholic solution (1: 1).

Preparation of DPPH solution: Solution of DPPH (0.1 mm) in methanol was prepared by dissolving 1.9 mg of DPPH in methanol and volume was made up to 100 ml with methanol. The solution was kept in darkness for 30 minutes to complete the reaction.

Determination of antioxidant activity

1.0 ml of DPPH solution was added to 1.0 ml of different extracts and allowed to stand at room temperature for 30 min, and then absorbance was measured at 517 nm in a spectrophotometer. Similarly 1.0 ml of the extract in distilled water was added to 0.6 ml of hydrogen peroxide solution and the absorbance was measured at 230 nm in a spectrophotometer. The percentage inhibition was measured by following formula (18-20):

$$\% \text{ inhibition} = (\text{Ac}-\text{At}) \times 100/\text{Ac}$$

Ac is the absorbance of control, at is the absorbance of test sample

Anti-inflammatory activity

The reaction mixture (5 ml) was consisted of 0.2 ml of egg albumin (from fresh hen's egg), 2.8 ml of phosphate buffered saline (PBS, pH 6.4) and 2 ml of varying concentrations of alcoholic extract of the clove bud to obtain concentrations of 100, 200, 400, 800 and 1000 µg/ml. Similar volume of double-distilled water served as a control. Then the mixtures were incubated at (37±2) °C in a Biochemical oxygen demand (BOD) incubator for 15 min and then heated at 70°C for 5 min. After cooling, their absorbance was measured at 660 nm. Diclofenac sodium (100 µg/ml) was used as a reference drug. The percentage inhibition of protein denaturation was calculated by using the following formula:

$$\text{Percentage inhibition} = (\text{Abs control} - \text{Abs test sample}) * 100 / \text{Abs control}$$

Whereas Abs = Absorbance.

Result and Discussion

The essential oil components of the flower buds of *S. aromaticum* are listed in Table-1. The components are arranged in order to GC elution on QP-2000 column. The oil was characterized by a large amount of eugenol (55.6%) and followed by eugenol acetate (12.5%), chavicol (6.5%), methyl salicylate (5.9%), β-aryophyllene (5.2%), α- caryophyllene (4.9%), p-allyl phenol (2.8 %), δ - cadinene (2.5 %), caryophyllene oxide (1.4%) and linalool (1.1 %). α-Pinene and p-cymene occurred in trace amounts.

Antimicrobial activities of the dried alcoholic extract and different concentrations of the essential oil are summarized in Table-2. The maximum antibacterial activity was observed with 1% v/v of the essential oil against *Propionibacterium acne* (17.8 mm) followed by *S. epidermidis*. (16.8 mm).

The free radical scavenging activity of the essential oil was determined at all concentrations from 10 to 200 µg /ml and was significant with alcoholic extract (10 %) essential oil (0.5 %) of the flower buds (Table 3). The results demonstrated that the extracts of the *S. aromaticum* flower buds and isolated flavonoids have effective activity as hydrogen donors and as primary antioxidants by reacting with lipid radicals. Anti-inflammatory activities are summarized in Table - 4. Dose dependent anti-inflammatory response was observed with increasing concentration of the clove bud oil. However, 1% v/v of the essential oil exhibited significant effect comparable results to that of Diclofenac taken as reference standard.

Table 1: Essential oil constituents of the dried unripe flower buds of *S. aromaticum*

S. No.	Essential oil components	Kovat's indices	Area %
1	α - Pinene	925	0.9
2	p - Cymene	1026	0.7
3	Linalool	1102	1.1
4	Methyl salicylate	1190	5.9
5	p - Allyl phenol	1251	2.8
6	Chavicol	1255	6.5
7	Eugenol	1351	55.6
8	B- Caryophyllene	1444	5.2
9	α - Caryophyllene	1454	4.9
10	δ - cadinene	1483	2.5

11	Eugenol acetate	1506	12.5
12	Caryophyllene oxide	1572	1.4

Table 2: Anti-microbial activity of the essential oil, alcoholic extract and aqueous extract of the flowers buds of *S. aromaticum*

S. No.	Test Organism	Zone of Inhibition in mm ^a			Dried Alcoholic Extract 5.0 %w/v	Standard Clindamycin (0.1 mg/ml)		
		Conc. of Volatile Oil						
		0.1 %v/v	0.5 %v/v	1.0 %v/v				
1	<i>Propionibacterium acne</i>	16.4	17.1	17.8	17.6	18.6		
2	<i>Staphylococcus epidermidis</i>	16.0	16.4	16.8	16.4	17.8		

^a an average of triplicate

Clindamycin - Against acneogenic microbes

Table 3: DPPH Radical Scavenging Activity of the essential oil and aqueous and alcoholic extralcts of flower buds of *S. aromaticum*

S. No.	Concentration (µg/ml)	Ascorbic acid	% of essential oil of <i>S. aromaticum</i> flower buds (v/v)			Aqueous extract of clove buds	Alcoholic extract of clove buds
			0.1%	0.5%	1.0%		
1	10	46.35±3.31	33.05±5.44	34.78±3.90	34.90±4.48	32.05±1.04	34.06±1.90
2	50	71.66±6.11	34.85±5.36	34.99±7.06	34.98±4.89	32.35±3.55	34.54±2.89
3	100	79.10±5.80	34.90±6.69	35.10±5.19	35.89±4.34	32.90±6.56	34.87±2.07
4	200	87.44±4.74	34.67±5.05	35.67±4.76	35.99±2.05	33.67±3.89	35.67±6.04
5	250	94.44±4.13	35.17±6.31	35.90±7.29	30.01±4.32	33.17±7.85	35.17±5.09
6	EC 50	14.9 µg	158.50 µg	164.90 µg	170.80 µg	25 µg	15 µg

Values are expressed as mean ± S.D., n = 4

Table 4: *In vitro* Anti-inflammatory activity of essential oil *S. aromaticum* flower buds

S. No.	Conc. (µg/ml)	% of inhibition (10%w/w aq. ext)	% of inhibition (10% w/w alc. ext)	% of inhibition (0.1% v/v of v. oil)	% of inhibition (0.5% v/v of v. oil)	% of inhibition (1.0% v/v of v. oil)
1	100	1.51±0.87	1.89±0.45	3.11±1.89	4.02±0.93	5.43±0.86
2	200	1.78±0.74	1.99±0.79	3.66±0.90	4.67±0.96	5.68±0.58
3	300	1.90±0.67	2.01±0.59	3.78±1.43	5.40±1.42	6.47±1.91
4	400	2.09±0.72	2.78±0.90	3.92±1.45	6.62±1.67	6.72±1.75
5	500	2.87±0.89	2.96±0.69	4.40±0.70	6.80±1.78	6.90±1.65
6	Diclofenac	7.04±0.98	7.04±0.98	7.04±0.98	7.04±0.98	7.04±0.98

Conclusion

The essential oil of the flower buds of clove (*Syzygium aromaticum*) was constituted mainly of eugenol (55.6%), eugenol acetate (12.5%), chavicol (6.5%), methyl salicylate (5.9%), β-caryophyllene (5.2%) and α- caryophyllene (4.9%). The alcoholic extract of clove buds showed significant antibacterial activity against *Propionibacterium acne* and *Staphylococcus epidermidis*. The clove essential oil exhibited strong free radical scavenging activity in comparison to standard ascorbic acid. The clove bud oil showed dose dependent anti-inflammatory response against reference standard Diclofenac sodium. These observations justify the use of clove for the treatment of acne and pimples due to its marked antimicrobial, anti-oxidant and anti-inflammatory activities.

Acknowledgment

The authors are thankful to the Head, Advanced Instrumentation Research Facility, Jawaharlal Nehru University, New Delhi for recording GC-MS analytical data of the essential oil.

References

- Chaieb K, Hajlaoui H, Zmantar T, Kahla-Nakbi AB, Rouabha M, Mahdouani K, Bakhouf A. The chemical composition and biological activity of clove essential oil, *Eugenia caryophyllata* (*Syzygium aromaticum* L. Myrtaceae): a short review. *Phytther Res.* 2007; 21(6):501-506.
- Cortés-Rojas DF, Fernandes de Souza CR, Oliveira WP. Clove (*Syzygium aromaticum*): a precious spice. *Asian Pac J Trop Biomed.* 2014; 4(2):90-96.
- Jirovetz L, Buchbauer G, Stoilova I, Stoyanova A, Krastanov A, Schmidt E. Chemical composition and antioxidant properties of clove leaf essential oil. *J Agric Food Chem.* 2006; 54(17):6303-6307.
- Nassar MI, Gaara AH, El-Ghorab AH, Farrag ARH, Shen H, Huq E, Mabry TJ. Chemical constituents of clove (*Syzygium aromaticum*, Fam. Myrtaceae) and their antioxidant activity. *Rev. Latinoamer. Quím.* 2007; 35(3):47 – 57.
- Oliveira RA, Reis TV, Sacramento CK, Duarte LP, Oliveira FF. Volatile chemical constituents of rich spices in eugenol. *Rev Bras Farmacognosia.* 2009; 19(3):771-775.
- Huang X-W, Feng Y-C, Huang Y, Li H-L. Chemical composition, antioxidant and the possible use as skincare ingredient of clove oil (*Syzygium aromaticum* (L.) Merr. & Perry) and citronella oil (*Cymbopogon goeringii*) from China, *J Essen Oil Res.* DOI: 10.1080/10412905.2013.775082. 2013; 25(4):315-323.
- Safrudin I, Maimulyanti A, Prihadi AR. Effect of crushing of clove bud (*Syzygium aromaticum*) and distillation rate on main constituents of the essential oil. *American J Essential Oils and Natural Products.* 2015; 2(3): 12-15.
- Sohilait HJ. Chemical composition of the essential oils in *Eugenia caryophylata*, Thunb from Amboina Island. *Science J Chem.* 2015; 3(6):95-99.
- Mohammed KAK, Abdulkadhim HM, Noori SI. Chemical composition and anti-bacterial effects of clove (*Syzygium aromaticum*) flowers. *Int. J. Curr. Microbiol.*

- App. Sci. 2016; 5(2):483-489.
10. Hamad A, Mahardika MGP, Yuliani I, Hartanti D. Chemical constituents and antimicrobial activities of essential oils of *Syzygium polyanthum* and *Syzygium aromaticum*. *Rasayan J Chem.* 2017; 10(2):564-569.
 11. Zheng GQ, Kenny PM, Lam KT. Sesquiterpenes from clove (*Eugenia caryophyllata*) as potential anticarcinogenic agents. *Journal of Natural Products.* 1992; 55:999–1003.
 12. Cai L, Wu CD. Compounds from *Syzygium aromaticum* possessing growth inhibitory activity against oral pathogens. *Journal of Natural Products.* 1996; 59:987-990.
 13. Kong QL, Song YZ, Zhang LL, Chen LY, Li QF. Natural antifungal compounds from *Syzygium aromaticum* (L.) Merr. et Perry. *Acta Agriculturae Shanghai.* 2004; 20(3):68–72.
 14. Gülcin İ, Şatlı İG, Beydemira Ş, Elmastaş M, Kürevioglu Öl. Comparison of antioxidant activity of clove (*Eugenia caryophyllata* Thunb) buds and lavender (*Lavandula stoechas* L.). *Food Chem.* 2004; 8(3):393-400.
 15. Abdel-Wahhab MA, Aly SE. Antioxidant property of *Nigella sativa* (black cumin) and *Syzygium aromaticum* (clove) in rats during aflatoxicosis. *J Appl Toxicol.* 2005; 25(3):218-223.
 16. Bamdad F, Kadivar M, Keramat J. Evaluation of phenolic content and antioxidant activity of Iranian caraway in comparison with clove and BHT using model systems and vegetable oil. *Int J Food Sci Technol.* 2006; 41(1):20-27.
 17. Gülcin I, Elmastaş M, Aboul-Enein HY. Antioxidant activity of clove oil-A powerful antioxidant source. *Arab J Chem.* 2012; 5(4):489-499.
 18. De Oliveira MS, Da Costa WA, Pereira DS, Botelho RJS, De Alencar Menezes TO, de Aquiar Andrade EH, et al. Chemical composition and phytotoxic activity of clove (*Syzygium aromaticum*) essential oil obtained with supercritical CO₂. *The Journal of Supercritical Fluids.* 2016; 118:185-193.
 19. Chami F, Chami N, Bennis S, Trouillas J, Remmal A. Evaluation of carvacrol and eugenol as prophylaxis and treatment of vaginal candidiasis in an immunosuppressed rat model. *J Antimicrob Chemother.* 2004; 54(5):909-914.
 20. Ali SM, Khan AA, Ahmed I, Musaddiq M, Ahmed KS, Polasa H, et al. Antimicrobial activities of eugenol and cinnamaldehyde against the human gastric pathogen *Helicobacter pylori*. *Ann Clin Microbiol Antimicrob.* 2005; 4:20.
 21. Pérez-Conesa D, Mc Landsborough L, Weiss J. Inhibition and inactivation of *Listeria monocytogenes* and *Escherichia coli* O157:H7 colony biofilms by micellar-encapsulated eugenol and carvacrol. *J Food Prot.* 2006; 69(12):2947-2954.
 22. Ali S, Prasad R, Mahmood A, Routray I, Shinkafi TS, Sahin K, Kucuk O. Eugenol-rich fraction of *Syzygium aromaticum* (Clove) reverses biochemical and histopathological changes in liver cirrhosis and inhibits hepatic cell proliferation. *Journal of Cancer Prevention.* 2014; 19 (4):288-300.
 23. Park MJ, Gwak KS, Yang I, Choi WS, Jo HJ, Chang JW, et al. Antifungal activities of the essential oils in *Syzygium aromaticum* (L.) Merr. Et Perry and *Leptospermum petersonii* Bailey and their constituents against various dermatophytes. *J Microbiol.* 2007; 45(5):460-465.
 24. Fu Y, Zu Y, Chen L, Shi X, Wang Z, Sun S, et al. Antimicrobial activity of clove and rosemary essential oils alone and in combination. *Phytother Res.* 2007; 21(10):989-994.
 25. Rana IS, Rana AS, Rajak RC. Evaluation of antifungal activity in essential oil of the *Syzygium aromaticum* (L.) by extraction, purification and analysis of its main component eugenol. *Braz J Microbiol.* 2011; 42(4):1269-1277.
 26. Park IK, Shin SC. Fumigant activity of plant essential oils and components from garlic (*Allium sativum*) and clove bud (*Eugenia caryophyllata*) oils against the Japanese termite (*Reticulitermes speratus* Kolbe). *J Agric Food Chem.* 2005; 53(11):4388-4392.
 27. Daniel AN, Sartoretto SM, Schimidt G, Caparroz-Assef SM, Bersani-Amado CA, Cuman RK. Anti-inflammatory and antinociceptive activities of eugenol essential oil in experimental animal models. *Rev Bras Farmacogn.* 2009; 19(1B):212-217.
 28. Kafle L, Shih CJ. Toxicity and repellency of compounds from clove (*Syzygium aromaticum*) to red imported fire ants *Solenopsis invicta* (Hymenoptera: Formicidae). *J Econ Entomol.* 2013; 106(1):131-135.
 29. Afify AE, El-Beltagi HS, Aly AA, El-Ansary AE. Antioxidant enzyme activities and lipid peroxidation as biomarker for potato tuber stored by two essential oils from caraway and clove and its main component carvone and eugenol. *Asian Pac J Trop Biomed.* 2012; 2(2):772-780.
 30. Chaieb K, Zmantar T, Ksouri R, Hajlaoui H, Mahdouani K, Abdelly C, Bakhouf A. Antioxidant properties of essential oil of *Eugenia caryophyllata* and its antifungal activity against a large number of clinical Candida species. *Mycoses.* 2007; 50:403- 406.
 31. Hekimoğlu MA, Ergun M. Evaluation of clove oil as anaesthetic agent in fresh water angelfish, *Pterophyllum scalare*. *Pak J Zool.* 2012; 44(5):1297-1300.
 32. Huang Y, Ho SH, Lee HC, Yap YL. Insecticidal properties of eugenol, isoeugenol and methyl eugenol and their effects on nutrition of *Sitophilus zeamais* Motsch. (Coleoptera: Curculionidae) and *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *J Stored Prod. Res.* 2002; 38:403 –412.
 33. Atanasova-Pancevska N, Bogdanov J, Kungulovski D. *In Vitro* Antimicrobial activity and chemical composition of two essential oils and eugenol from flower buds of *Eugenia caryophyllata*. *Open Biological Sciences Journal.* 2017, 3:16-25.
 34. Alshaikh N, Perveen K. Anti-candidal activity and chemical composition of essential oil of clove (*Syzygium aromaticum*). *Journal of Essential Oil Bearing Plants.* 2017; 20(4):951-958.
 35. Lee S, Najiah M, Wendy W, Nadirah M. Chemical Composition and antimicrobial activity of the essential oil of *Syzygium aromaticum* flower bud (Clove) against fish systemic bacteria isolated from aquaculture sites. *Frontiers of Agriculture in China.* 2009; 3(3):332-336.
 36. Halder S, Mehta AK, Kar R, Mustafa M, Mediratta PK, Sharma KK. Clove oil reverses learning and memory deficits in scopolamine treated mice. *Planta Med.* 2011; 77(8):830-834.
 37. Adams RP. Identification of Essential Oil Component by Gas Chromatography/ Mass Spectrometry, (4th ed.). IL: Allured Publishing Co., Carol Stream, 2007.
 38. Ali M. Techniques in terpenoid identification. Birla Publications, Delhi, 2001.