

ISSN (E): 2320-3862 ISSN (P): 2394-0530 NAAS Rating: 3.53 JMPS 2018; 6(6): 217-221 © 2018 JMPS Received: 01-09-2018 Accepted: 03-10-2018

#### C Porchselvi

Department of Botany, Ayya Nadar Janaki Ammal College (Autonomous), Sivakasi, Tamil Nadu, India

#### S Muthulakshmi

Department of Botany, Sri Parasakthi College for Women (Autonomous), Courtallam, Tamil Nadu, India

# Pharmacognostic and phytochemical evaluation of Antiaris toxicaria (Pers). Lesch

## C Porchselvi and S Muthulakshmi

#### Abstract

**Background:** *Antiaris toxicaria* (Pers). Lesch. (Moraceae) historically has been used as arrow poison and also for a variety of medicinal purposes. Despite the popular medicinal utilization, still no conclusive study has been reported so far regarding the pharmacognostical standardization.

**Aim:** Thus, the present study was focused to scientifically establish a standard monograph of *Antiaris toxicaria* (Pers). Lesch on the basis of pharmacognostical and phytochemical parameters.

**Materials and methods:** The detailed macroscopic and qualitative as well as quantitative microscopic characters and phytochemical characters of *Antiaris toxicaria* (Pers). Lesch were analysed.

**Results:** The morphological characters of *Antiaris toxicaria* were established. The transverse section of leaf shows adaxial epidermis has mucillagenous hairs, the palisade is single layered, the cells are wide and columnar. Calcium oxalate crystals are abundant in the ground cells of the petiole. Phytochemical parameters of *Antiaris toxicaria* (Pers). Lesch showed loss on drying 7.01% and total ash 4.97%.

**Conclusion:** The present study provided useful information about its correct identity and evaluation. It helps to diagnose drugs from the plant.

Keywords: Pharmacognostic, phytochemical evaluation, Antiaris toxicaria (Pers). Lesch

# Introduction

Antiaris toxicaria (Pers). Lesch belonging to the family Moraceae, commonly known as Marauri. Antiaris toxicaria is a tall tree recorded from western ghats at altitudes of 300m height. Leaves oval, oblong, obtuse or rather acute unequally cordate when young toothletted and hairy on both sides, peduncle simple. Fruit fleshy one seeded drupe. Antiaris toxicaria is distributed in Western Ghats of Tamil Nadu and Andaman Nicobar Islands. Antiaris toxicaria is reported to have antioxidants, antibacterial antifungal and anti-inflammatory activity. The plant is highly poisonous. Female flowers are one of the most virulent of known poisons some persons are exposed to danger when they only approach the trees. Four persons sent a man up into a tree he became very ill, his body swelled for several days, he suffered severely by vertigo nausea and vomiting others nevertheless experience on inconvenience from the exhalations of the tree. The bark, leaf and seeds of Antiaris toxicaria is useful for the treatment of febrifuge and dysentery. The milky sap contains the poisonous glycosides antiarin and antiosidin which exerts a rapid and aggressive action on the heart. Antiaris toxicaria contains glycosides, saponins, various phenolic compounds, volatile oil, tannin, fatty acids, triterpenes, protein starch, glucose and minerals. As far as phytochemistry and pharmacognosy of *Antiaris* toxicaria is concerned large number of scientific data is available but a pharmacognostical standardization study is still lacking. Hence, the present study was focused to investigate pharmacognostical and phytochemical properties of Antiaris toxicaria.

## **Materials and Methods Plant Collection**

Antiaris toxicaria was collected in the flowering stage from the way to Chirtaruvi in Courtallam, Western Ghats, Tamil Nadu, India. During June 2012. According to Beddome, it is the largest tree of the evergreen forests of Western Ghats.

## **Chemicals and Instruments**

All reagents and chemicals used for pharmacognostic screening were analytical grade, compound light microscope was used for the study. The photography was done by using microscope camera. Camera lucida was used for determination of quantitative microscopical

Correspondence C Porchselvi Department of Botany, Ayya Nadar Janaki Ammal College (Autonomous), Sivakasi, Tamil Nadu, India characters. Fluorescent analysis of the leaf powder in different solvents were carried out according to the methods of Chase and Pratt. Physicochemical characters were determined by standard methods.

## Morphology and Microscopy

The morphological characters such as shape, size, colour, odour, taste, surface and fractures were determined. Microscopic features of *Antiaris toxicaria* were evaluated by preparing thin hand section. The sections were cleared with alcohol and stained. Histo chemical reactions were applied with various chemicals and photomicroscopy was performed with camera. The leaf constants were measured using Camera lucida.

#### **Powder Microscopy**

The powder photomicroscopy of shade dried *Antiaris* toxicaria was carried for identification of the powder characteristics by using camera.

#### Results

## **Morphological Characteristics**

Morphological studies shows (Fig. 1) that the leaf was simple, leathery, bifarious, penninerved, glossy on the upper surface, short- stalked, oval with an asymmetrical, heart shaped base and softly spiny at the top. Leaf margin is irregular and wavy. The tree has a straight cylindrical trunk. The flowers are arranged in catkins at the tips of the branches. Flowers monoecious, more crowded on the surface of an axillary pedunculate receptacle, surrounded by confluent imbricating bracts, 3-4 spathulate imbricate sepals, having a large number of stamens and longer stalks than the female flowers. Female flowers minute solitary in an involucres of many confluent bracts without a perianth, ovary adnate to involucres, style arms recurved. The fruit is an elongate berry covered with fleshy scales and containing a hard pit, seed exalbuminous, testa hard, embryo globose, cotyledons equal, radicle small (Fig. 1).

## Qualitative microscopically characteristics Transverse section of leaf

The leaf has prominent midrib projecting both adaxially and abaxially and their lamina. The lateral veins are not prominent. The lamina is dorsiventral smooth and even. The adaxial epidermis has mucillagenous hairs (Fig 2). The cells are rectangular to squarish and vary in size. Cuticle is fairly thick. The lamina is  $100\mu m$  thick. The palisade cells are  $40\mu m$  in height. The cells are wide and columnar. The spongy mesophyll consist of 2 layers of lobed and spherical cells. The lateral vein has circular collateral vascular strand with parenchymatous bundle sheath.

## Midrib

The midrib is  $1\mu m$  thick and  $800\mu m$  in breadth. It has an adaxial thick and blunt hump and abaxial broad hemispherical body. The outline is wavy bearing thick walled sparsely distributed trichomes. The vascular bundles occur in two strands; there is a wide abaxial bowl shaped main strand and another adaxial thick and flat accessory strand (Fig. 3). The xylem elements occur in regular radial files and phloem in

thin abaxial arc. Calcium oxalate crystals (druses or sphero crystals) are abundant in ground cells of the petiole (Fig. 3). The mesophyll crystals are arranged in the form of petals (Fig. 3).

#### **Stomata**

Stomata occur only on the abaxial side and are anomocytic type (Fig. 3). The epidermal cells are much lobed. The anticlinal walls are highly wavy or thin walled and smooth (Fig. 2 and 3). The epidermal cells from which the trichome originates are circular, thick walled and lignified (Fig. 3).

#### Veination

The lateral veins are fairly thick with distinct Veinlets. The vein islets are not distinct. The vein terminations are long and thin; they are either simple or branched once or twice (Fig. 3). Crystals mostly secreted in the form of clusters. Structures resembling cystoliths also occur in the tips or suspended from the walls of the hairs in species of *Antiaris* (Metcalf and Chalk, 1972). Silicified pegs attached to the outer walls of the epidermal cells.

## **Preliminary Phytochemical Screening**

The preliminary phytochemical screening with the various qualitative chemical tests and fluorescence analysis were carried out. The results were shown in Table 1, 2 and 3.

**Table 1:** Determination of physical constants:

Sl. No	Parameters	Results (%w/v)
1.	Total ash	4.91
2.	Water soluble Ash	1.16
3.	Acid insoluble Ash	0.33
4.	Loss on drying at 105 <sup>0</sup> C	6.01
5.	Water soluble extractives	15.10
6.	Alcohol soluble extractives	9.9
7.	Extractive value (Successive extraction)	
	Hexane	1.2
	Petroleum ether	2.6
	Chloroform	2.9
	Ethyl alcohol	5.1

**Table 2:** Preliminary phytochemical screening of ethanolic extract of *Antiaris toxicaria* (Pers). Lesch.

Components	Results	
Steroid	Positive	
Triterpenoid	Positive	
Flavonoid	Positive	
Furan	Negative	
Sugar	Positive	
Coumarin	Positive	
Quinone	Positive	
Alkaloid	Positive	
Tannin	Positive	
Phenol	Positive	
Acid	Negative	
Saponin	Positive	

**Table 3:** Fluorescence analysis of the leaf of *Antiaris toxicaria* (Pers). Lesch.

Sl. No	Treatment	Under visible light	Under UV 265 nm	Under UV 365 nm
1.	Powder	Dark Green	Dark Green	Dark Brown
2.	Powder +Petroleum ether	Dark Green	Brown	Black
3.	Powder + Hexane	Dark Green	Brown	Blackish Brown
4.	Powder + Chloroform	Brown	Black	Black
5.	Powder + Ethylacetate	Pale green	Dark Green	Black
6.	Powder + Ethyl alcohol	Brown	Dark green	Brown
7.	Powder + Distilled water	Light green	Pale green	Dark green
8.	Powder + 1N NaOH	Reddish Brown	Dark green	Blackish Brown
9.	Powder + 1N HCL	Light brown	Pale green	Dark brown



Fig 1: Antiaris toxicaria (Pers.) Lesch. Leaf

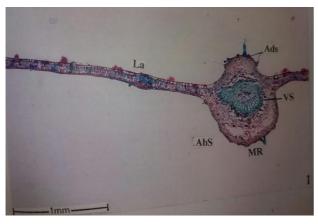


Fig 2: T.S. of leaf

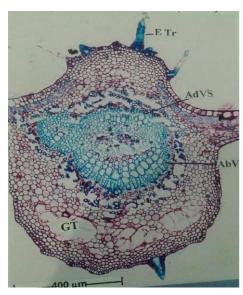


Fig 3: T.S. of Midrib

#### **Discussion**

Standardization is an important tool in identifying crude drug correctly. For establishing the correct identity of source materials, microscopic method is one of the simplest and best methods. Therefore, the results of the present study, may serve as a basis for identification, collection and standardization of the plant.

Microscopical study the leaf showed the presence of mucillagenous hairs on the adaxial epidermis. The spongy mesophyll consist of 2 layers of lobed and spherical cells. He lateral vein has circular collateral vascular strand with parenchymatous bundle sheath. The vascular bundles occur in two strands, there is a wide abaxial bowl shaped main strand and another adaxial thick and flat accessory strand. The xylem elements occur in regular radial files and phloem in thin abaxial arc. Calcium oxalate crystals are abundant in ground cells of the petiole. The vein islets are not distinct. The vein terminations are long and thin. These characters constitute the reliable features for botanical identity of the plant.

Physicochemical parameters of *Antiaris toxicaria* (Pers.) Lesch showed loss on drying

6.01% and total ash 4.91%, the weight of the ash left behind after the combustion is of important parameter for the standardization of drug. Every part of the plant provides a particular amount of ash. The weight of total ash therefore gives information whether it is adultered with any other organic or inorganic materials. *Antiaris toxicaria* contains the acid insoluble ash 0.33% and water soluble ash 1.16%. The acid insoluble ash gives an idea about the earthy matter and other impurities which might be present along with drug.

Extractive values of the plant with different solvents give a preliminary picture of the percentage of the compounds extracted. In *Antiaris toxicaria* extractive value was found with ethanol (4.2%) minimum (1.02%) with hexane.

This result shows the solvent ethanol is preferable to other solvent for the yield of more of the compounds.

The extract of *Antiaris toxicaria* is exposed to UV light, it exhibits fluorescent effects, that provides evidence for the presence of fluorescent compounds.

Qualitative tests carried out in the leaf of *Antiaris toxicaria* confirmed the presence of various pharmacologically important plant constituents like triterpenoid, phenol, flavonoid, coumarin, quinine, glycosides, sugars, alksloids, steroids and saponins. Furan and acids are absent in the extract. For instance, the presence of tannins may be responsible for ability of *Antiaris toxicaria* to cure diseases like diabetes, diarrhea, sore throat, skin ulcer and dysentery. The presence of flavonoids in *Antiaris toxicaria* may be responsible for its uses to cure cancer, inflammations and allergies. The presence of alkaloids may be useful to cure heart diseases.

## Conclusion

The present study provided useful information about its

correct identity and evaluation. It helps to differentiate from the closely related species of *Antiaris*. This is also useful for the future identification of the plant, and serves as a standard monograph for identification and evaluation of plant.

#### Acknowledgement

The authors are grateful to Botanists Dr. V. Chelladurai, Rtd. Senior Research Officer, Tirunelveli and Dr. P. Jayaraman, Director, Plant Anatomy Research Centre, Chennai, Tamil Nadu for their sustained interest and offering valuable suggestions during the course of this research work.

## References

- 1. Ahn BT, Kang SS. Phytochemical studies on *Euphorbia pekinensis*. Medicinal and Aromatic Plants Abstracts. 1998; 20(1):312-314.
- 2. Ahn BT, Lee KS. Phytochemical studies on *Euphorbia ebracteolata*. Medicinal and Aromatic Plants Abstracts. 1998; 20(1):53-55.
- 3. Akinyumi KO, Oluwa OK, Omomigbehin EO. Antimicrobial activity of crude extracts of three medicinal plants used in South West Nigerian folk medicine on some food borne bacterial pathogens. Africans Journal of Traditional complementary and alternative medicines. 2006; 3(4):13-22.
- Asha VV, Sheeba MS, Suresh V, Wills PJ. Hepatoprotective activity of *Phyllanthus maderaspatensis* against experimentally induced liver Injuiry in rats. Journal of Ethnopharmacol. 2006; 104(1-2):79-86.
- Asolkar LV, Kakkar KK, Chakre OJ. II supplement to Glossary of Indian Medicinal Plants with Active Principles, Part-I (A-K), C.S.I.R., New Delhi, 1992.
- Audu JA, Kala SL, Uroma VV. Antimicrobial activity of quinolinium fluoro chromate on some plants. Journal of Economic and Taxonomic Botany. 1995; 24(3):261-263.
- 7. Bagal Kottkar G, Sagijneedu SR, Saad MS, Stanslas J. Phytochemical from Phyllanthus niruri, Linn and their Pharmacological properties a review. Journal of Pharmacol. 2006; 58(12):1559-70.
- 8. Bailey LH. The standard cycopedia of Horticulture vol (1). The MacMillan Company, New York. Chicago, 1982, 118.
- 9. Bhattacharjee R, Sil PC. The Protein fraction of Phyllanthus niruri plays a protective role against acetaminophen induced hepatic disorder via its antioxidant properties. Phytother Res. 2006; 20(7):597-601
- 10. Chatterjee M, Sil PC. Hepatoprotective effect of aqueous extract of Phyllanthus niruri on nimesulide induced oxidative *in vivo*. Indian Journal of biochemistry and Biophysics. 2006; 43(5):299-305.
- 11. Chauhan CK, Nahivadekar SA. Indian Journal of Pharmacology. 1992; 24(2):107-110.
- 12. Chelldurai V. Personal Communication Researcher, Botany CCRAS, Palayamkottai, 2006.
- 13. Chopra RN, Nayer SL, Chopra IC. Glossary of Indian Medicinal Plants. C.S.I.R. New Delhi, 1956, 330.
- 14. Datta SC. Systematic Botany 4<sup>th</sup> edition. Wiley Eastern Ltd. Madras, 1989, 318-319.
- 15. Datta SC Systematic Botany 4<sup>th</sup> edition Reprint, New age International (P) Ltd Publishers. Delhi, 1999, 321.
- 16. Dhanda Pani, Sathya R, Moorthy J, Kalanithi C. Antimicrobial activity of *Aristolochia indica*. L., National symposium on currents trends in Medicinal Plants and

- Herbal Technology, 2006, 154.
- 17. Dickey RA, Hussain. Phytochemical studies on Phyllanthus niruri. Journal of Natural Products. 1995; 58(10):1515-1520.
- 18. Dietrich Brandis KCIE. Indian Trees. International Book Distributors. 9/3, Rajpur Road, Dehradun, 1978, 563.
- 19. Divya Goyal, Seema Bhaduria. Characterization of four Emblica officinalis grown in nurseries and plantation at Agra. 2006; 5(3):443-445.
- 20. Drury H. Hand Book of the Indian flora. Periodical expert Book agency, D-42, Vivek Vihar, Delhi. 1869; 3:151-152.
- 21. Gamble JS. A Manual of Indian Timbers. Bishen Singh Mahendra Pal Singh, 23-A. New Connaught place. Dehradun, 1972, 611.
- 22. Gamble JS. Flora of the Presidency of Madras published under the authority of the secretary of state for Indian council, London. 1921; 3:1310.
- 23. Gill LS, Akinwunmi C. Nigerian Medicine practice and Beliefs of the onelo people. Journal of Ethnopharmacology. 1986; 18:257-266.
- 24. Ezhilarasi R, Amerijyothy S. Pharmacognostic and Antimicrobial studies on Xanthium strumarium. Linn. (Asteraceae). National symposium on current trends in Medicinal Plants and Herbal Technology, 2006, 132.
- 25. Gitanjali G, Muthulakshmi S, Revathy N. Journal of Theoretical and Experimental Biology, Elias Academic Publishers, India. 2006; 3(1):43-46.
- 26. Harami M, Adamu alibilar A, Chelea Mathew N. Antimicrobial and Phytochemical studies of some selected medicinal plants in Bauchi (Nigeria). Medicinal and Aromatic Plants Abstracts. 2000; 24(1):216.
- 27. Hari Krishnan S, Henry Joseph K. New Millenium Seminar Proceedings, 2000, 97-98.
- 28. Haslam L, Liley TH, Yachi M, Magnolatha R. Traditional Herbal Medicine. The role of polyhedrons. Plants Medica. 1989; 55:1-8.
- 29. Henry AN, Kumari GR, Chitra V. Flora of Tamil Nadu, India. Botanical Survey of India, Southern Circle, Coimbatore. India. 1987; 3:236-239.
- 30. Henry Joseph L, Inigo VI. New Millenium Seminar on Medicinal Plants Proceeding, 2000, 232-233.
- 31. Hooker JD. System of Botany. Descriptive and Analytical. International Books and Periodical supply service. 24B/5. Deshbandhu Gupta Road, Karol Bagh. New Delhi, 1986, 692.
- 32. Huang. Phytochemical studies on Phyllanthus niruri. Planta Medica. 1952; 58(5):473-474.
- 33. Hutchinson J. Evolution and Phylogeny of flowering Plants. Academic Press. London. New York. P: 277.
- 34. Hutchinson, J. 1973. The Families of Flowering Plants III<sup>rd</sup> edition. Clarendon Press. Oxford, 1969, 330.
- 35. Jawahar C, Raveendran. Some lesser known Edible Plants among the tribals of Kerala. India. Ethnobiology in Human Welfare, Deep Publications, Delhi, 1996, 52-53.
- 36. Jones SBJ, Luchsinger AE. Plant Systematics. (2<sup>nd</sup> ed.). Mc. Graw-Hill Book. Co., New York, 1987.
- 37. Kassuya CA, Silvestre A, Menezes-de-Liman JX, Marotta DH, Rehder VL, Calixto JB. Anti inflammatory and antiallodynic actions of lignan niranthin isolated from Phyllanthus amarus. Evidence for interaction with platelet activating factor receptor. European Journal of Pharmacology. 2006; 548(1-3):182-188.
- 38. Khatoon S, Raj V, Rauval AK, Mehrotra S. Comparative Pharmacognostic studies on three Phyllanthus species,

- Journal of Sep. Sci. 2005; 28(6):581-585.
- 39. Kishore G, Bhat AT, Andrade SG, Karadesai BM, Hema Shittar M, Patil CS. Antimicribial susceptibility of Salmonella typhii. The Indian Journal of Medicinal Research. 1998; 101:247.
- Leite DF, Kassuya CA, Mazzuco TL, Silvestre A, Demelo LV, Rehder VL, et al. The cytotoxic effect and the multi drug resistance reversing action of lignans from Phyllanthus amarus. Plants Medica. 2006; 72(15):1353-1355.
- 41. Lieut CH. Hand Book of the Indian flora. Periodical Expert Book Agency. D-42. Vivekvihar. Delhi. 1869; 3:152.
- 42. Maridoss M, Victor B, Benniamin A, Mannarmannan M, John De Britto. Anti inflammatory activity of Phyllanthus singampattiana leaf extract, 2005.
- 43. Mathew KM. The flora of Tamil Nadu Karnatic. Gamopetalae and Monochlamydeae. The Rapinat Herbarium, St. Johns College. Tiruchirappalli, India. 1983; 3:1462-1470.
- 44. Metcalfe CR, Chalk L. Anatomy of the Dicotyledons vol:II Clarendon Press. Oxford. 1950; 1219, 1212, 1213, 1217
- 45. Mulchandanai NB, Hassaranai SA. Isolation of alkaloids in Phyllanthus species. Indian Journal of Chemistry. 1991; 298(9):801-803.
- 46. Muthulakshmi S, Umadevi S. New Millenium Seminar on Medicinal Plants Proceedings, 2000, 172-175.
- 47. O' Brein TP, Feder N, MC Cull, ME. Polychromatic Staining of Plant cell wall by toluidine blue. Journal of Protoplasma. 1964; 59:364-373.
- 48. Olivera DF, Pereira AC, Figueiredo HC, Carvalho DA, Silva G, Nunes AS, *et al.* Antibacterial activity of plants extracts from Brazilian South Eastern Region. Filoterapia. 2007; 78(2):142-145.
- 49. Pandey BP. Taxonomy of Angiosperms. S. Chand and company, Ltd. Ram Nagar. New Delhi, 1982, 218.
- 50. Parameswari CS, Tulasi Latha A. Antimicrobial activity of Ricinus communis. Medicinal and Aromatic Plants Abstracts. 2002; 24(4):537.
- 51. Pelzar MI, Chan ECS, Kreieg NR. Microbiology 5<sup>th</sup> edition. Tata McGraw Hill Publishing Company Ltd. New Delhi, 1993, 918.
- 52. Radhakrishnan KG, Pandurangan AG, Pushpagandan V. Ethnobotany of wild edible plants. Ethnobiology in Human Welfare. Deep Publications. Delhi, 1996, 48-51.
- 53. Rajamani S, Wasiullakhan R, Betty D. Antibacterial activity of four essential Oils. Journal of Ecotoxicology and Environmental Monitoring. 1997; 7(3):211.
- 54. Ramesh N, Vishwanathan MB, Selvi VT, Lakshmanaperumalsamy P. Antimicrobial and Phytochemical studies on the leaves of Phyllathus singampattiana. Medicinal Chemistry Research. 2004; 13(6-7):348-360.
- 55. Rosa Kutty DJ, Ignacimuthu MS. Nem Millenium Seminar on Medicinal Plants Proceedings, 2000, 219.
- Sass JE. Elements of Botanical Micro-Technique. McGraw Hill Book Co. New York and London, 1940, 222.
- 57. Shanmugasundaram KR, Seethapathy PG, Shanmugasundaram ERS. Therapeutic uses of some medicinal plants. Medicinal and Aromatic Plants Abstracts. 1984; 6(1):67.
- 58. Shanthi A, Lowis Jesudass L. New Millenium Seminar on Medicinal Plants Proceedings. 2000; 3(5):20.