



ISSN (E): 2320-3862
ISSN (P): 2394-0530
NAAS Rating: 3.53
JMPS 2019; 7(4): 123-126
© 2019 JMPS
Received: 21-05-2019
Accepted: 23-06-2019

Saifuzzaman Sumon
Department of Pharmacy,
BRAC University, Dhaka,
Bangladesh

Md. Rezaul Hoque
Department of Pharmacy,
BRAC University, Dhaka,
Bangladesh

Sajib Paul
Department of Pharmacy,
BRAC University, Dhaka,
Bangladesh

Pharmacological activity investigation of *Callicarpa americana*

Saifuzzaman Sumon, Md. Rezaul Hoque and Sajib Paul

Abstract

Callicarpa americana plant extract used to assess its different biological activity. Extract was made by soaking the dried plant powder in methanol. After comparing with the standard we found that Methanol extract of the sample gave the activity against all the experimented microbes of ZI (zone of inhibition) against *E.coli* and *B.subtillis*. After performing the antioxidant, thrombolytic, antidiarrheal, hypoglycemic and cytotoxic activity assay of methanol extract of sample plant we saw that it has a good biological activity that can be used as a potential traditional medicine.

Keywords: *Callicarpa americana*, antioxidant, antimicrobial, antidiarrheal, hypoglycemic, thrombolytic activity

Introduction

Callicarpa americana also known as American beauty berry is a plant from lamiaceae family. Species from lamiaceae family are mostly shrubs and herbs containing 6900-7200 species^[4] that are largely used^[3] in the traditional medicine of Lebanon to treat several diseases as it shows antimicrobial, antibacterial, antifungal, antioxidant and some other pharmacological properties.

Some examples like *Callicarpa bodinieri* leaf is used^[1, 2, 5] to treat haematemesis, blennorrhoea, and haematuria. In addition, *Callicarpa dolichophylla* used in the treatment of diarrhea, fever, syphilis, and gastrorrhagia. Similarly, *Callicarpa caudata* and *Callicarpa macrophylla* is used as cure for stomach troubles. After that, *Callicarpa erioclona* and *Callicarpa loboapiculata* leaf is used as a wash to treat skin conditions such as pruritis.

The genus shows a lot of important pharmacological uses in ancient and recent time. So, several tests like phytochemical screening was done to find whether the plant have Antioxidant, Anti-microbial, cytotoxicity, thrombolytic activities, or any pharmacological active ingredients or not.

Methods and Materials

Collection of Plant Materials

The leaf part of *Callicarpa americana* plant was collected in May, 2017 from Chittagong hill tract. After collection, the National Herbarium Bangladesh (NHB), Mirpur, and Dhaka authenticated the plant material and provided a plant identification number, which was 47695.

Preparation of The Extract

At first, the leaves part was washed with fresh water to remove the unwanted dust particles and plant scrap. After that, the cleaned leaves were dried under the sun for a day. Then the leaves were again dried for 1 hour at 30-40°C in hot air oven. By using a high capacity grinding machine, the dry and crusty leaves were ground. After that, at a normal ambient temperature (22-25°C) around 900 g of ground powder was soaked in 2.5 L of methanol for a period of 2 days with occasional stirring. With the help of cotton filter (pore size: 110mm) filtration was done and rotary evaporator was used at 100 rpm at 30°C to evaporate the maximum amount of solvent. For vaporizing the solvent completely from the extract, the leaf extract was kept under laminar airflow cabinet. Moreover, it was used to avoid any possibility of microbial growth in the extract while drying. Finally, 22.4 g of plant leaf extract was obtained and kept in dry and cool place and proper labeling was done. After that, this extract was used to conduct antioxidant, brine shrimp lethality assay,

Correspondence
Saifuzzaman Sumon
Department of Pharmacy,
BRAC University, Dhaka,
Bangladesh

thrombolytic, antidiabetic, antimicrobial and hypoglycemic studies.

Chemicals

The chemicals were gallic acid [Sigma-Aldrich, USA], sodium chloride [Sigma-Aldrich, USA], Folin-Ciocalteu reagent [Sigma-Aldrich, USA], vincristine sulphate [Sigma-Aldrich, USA], 2,2-Diphenyl-1-Picrylhydrazyl (DPPH) [Sigma-Aldrich, USA], sodium carbonate [Merck, India] and ascorbic acid (ASA) [Merck, India], dimethyl sulfoxide (DMSO) [Fisher Scientific, UK], Castor oil (WELL's Health Care, Spain), 0.9% sodium chloride solution (normal saline) (Orion Infusions Ltd., Bangladesh), charcoal meal (10% activated charcoal in 5% gum acacia), and loperamide (Square Pharmaceuticals Ltd., Bangladesh) were used for antidiarrheal activity test, and dimethyl sulfoxide (DMSO) (Sigma-Aldrich, USA) and sodium chloride (Sigma) were used for cytotoxic activity test. All the chemicals used in this study were of analytical grade.

Anti-oxidant activity

Total phenolic content (TPC)

The phenols were oxidized by Folin-Ciocalteu in ionic phenolic solution. When the solution became yellow to dark blue, it is understood that the oxidation has been completed. After that, this color changed mixture measured in 760 nm in UV spectrophotometer. Finally, the value of the absorbance plotted in gallic acid calibration curve and data was evaluated as gallic acid equivalents (GAE).

Total Flavonoid Content

Aluminum chloride was used to determine the total amount of flavonoids. Firstly, 0.5 ml of plant extract was made the final volume of 1 ml for reaction medium (MeOH/H₂O/CH₃COOH=14:5:1) which was then mixed with Aluminum chloride reagent (4 ml, 133 mg of AlCl₃ × 6 H₂O and 400 mg of CH₃COONa dissolved in 100 ml H₂O). After 5 minute, the absorbance was measured at 430 nm. Based on the calibration curve, total flavonoid content was calculated and it was expressed as gram equivalents.

DPPH free Radical Scavenging Assay

The antioxidant activity of *Callicarpa americana* was determined by performing DPPH free radical scavenging assay. To run this assay, different concentrations of plant extracts were mixed with 2, 2-diphenyl-1-picrylhydrazyl (DPHH) solution. In methanol or aqueous solution, free radicals were generated due to delocalization of the free electrons and a deep purple colored solution is produced. Then absorbance of different concentration solutions was measured at 517 nm in UV spectrophotometer. The decreasing value of DPHH at 517 nm is directly proportional to the radical scavenging activity.

Percentage of inhibition of DPHH free radical (1%) was calculated by using the following equation:

$$(1\%) = (\text{Absorbance of blank} - \text{Absorbance of sample}) / \text{Absorbance of blank} \times 100$$

50% of inhibition of the extract concentration was calculated from the graph and the percentage of inhibition was plotted against extract concentration.

Cytotoxic Activity

Brine Shrimp Lethality Assay

In this assay, *Artemia salina* shrimp was used. Its offspring was hatched in replicated seawater to cultivate nauplii. Here,

calculated amount of dimethyl-sulfoxide (DMSO) was added with sample and desired concentration of sample was prepared by dilution. The counted nauplii were placed in vials that contained approximately 5 mL simulated seawater with visual inspection. With the help of micropipette, various concentrations of samples were added to tubes. Here, vincristine sulfate was used as standard. The sample containing tubes were then placed in a dry place for 24 hours at room temperature. At the last, after 24 hours, the survived nauplii were counted. Percentage (%) of mortality was calculated by using the following equation:

$$\text{Percentage of mortality} = (\text{Number of nauplii taken} - \text{Number of nauplii alive}) / \text{Number of nauplii taken} \times 100$$

50% of lethal concentration of extract concentration was calculated from the graph plotted percentage of mortality against concentration.

Thrombolytic Activity

The normal blood flow to the cells and tissues can be hampered due to thrombus as it blocks the blood vessel which can lead to lack of blood and oxygen. There are some thrombolytic medications like utokinase, clopifogrel, and streptokinase remove this thrombus and cells and tissues are remained in normal conditions. For this assay, fresh human blood was collected. Then, they were taken in three different pre-weighed sterile microbes and incubated for 45 minutes at 37°C. The upper fluid was entirely dispensed from all micro-tube lines when the clot was appeared. As a standard streptokinase was used and as a negative control distilled, water was used. 100 microliter of plant extract was taken in each tube and incubated for 90 minutes at 37°C. Next, liquid that was released from the clot was removed and the tubes were weighted again to observe the weight difference when the clot disruption occurred.

Percentage of clot lysis was calculated by following equation: (%) of clot lysis = (released clot weighted) / (clot weight after clot disruption) × 100

Antimicrobial Assay

Disc Diffusion Assay Method

In recent years, different studies are developing as antimicrobial agents to fight antibiotics resistance from different sources and highest concentration has given to screen and evaluate the antimicrobial activity. By using disc diffusion assay method, antimicrobial activity of *Callicarpa americana* was evaluated. *E. coli* bacteria (gram negative) and *Bacillus Subtilis* bacteria (gram positive) were used in this study. Mular Hinton agar (MHA) was used as media in this assay. Firstly, every petri dish was autoclaved for sterilization and 20 ml of MHA was poured in every petri dish. After that, the plates were kept for a time being to be settled. With the help of cotton swab, the nutrient broth of bacterial strains was incubated in MHA. Small disc of filter paper was made by using paper punch machine and then different concentrations of plant extract (200 mg/mL and 400 mg/mL) were used to swallow that filter paper. When the discs become dry, they were transferred to the petri dishes and kept in incubator for 24 hours at 37°C. After 24 hours the zone of inhibition were calculated and for keeping the contamination limited, whole experiment was done under laminar flow.

Result and Discussion

Antioxidant Activity

Total Phenolic Content (TPC)

In total phenolic content test, Gallic acid was used ad standard

and methanol extract of leaves which was used as a sample. The absorbance of the sample plotted in Gallic acid calibration curve. The absorbance of the plant extract was found 0.575 and TPC value was 78.71 GAE/g against that absorbance which indicates that the plant has antioxidant activity.

Total Flavonoid Content

The content of total flavonoid of the plant extracts was measured spectrophotometrically by using the aluminium chloride colorimetric assay. The flavonoid content of the extracts was expressed as mg quercetin equivalent per gram of the extract and it is 282.90 QE/g against the absorbance of 0.291 that indicates the present of flavonoid content.

Cytotoxic Activity

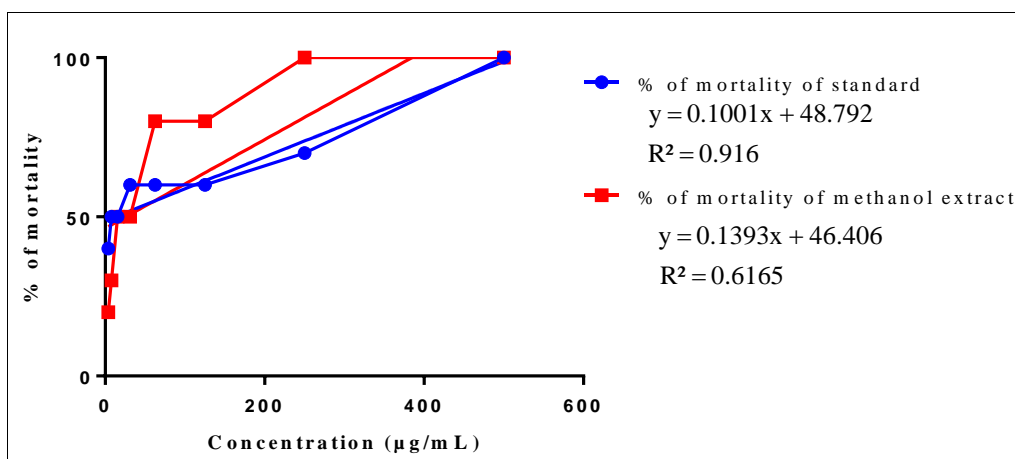


Fig 1: Cytotoxic Activity of Callicarpa Americana

This brine shrimp lethality assay was used to assess the cytotoxic property of methanol extract of plant material. Here, different concentrations standard and sample were plotted that provided different percentages of mortality. Percentage of mortality was found to increase along with the increasing

DPPH free Radical Scavenging Assay

It is known that DPPH free radical scavenging activity is increasing along with increasing concentration of the methanol extract. As the reference standard, ascorbic acid was used in this experiment for which IC₅₀ value was 65.688 µg/ml. on the other hand, the IC₅₀ value of the methanol extract of the sample plant was 311.82 µg/ml. this result indicates the presence of antioxidant activity which is less significant.

Table 1: Evaluation of DPPH free radical scavenging activity of methanol extract of *Callicarpa americana*.

	R ² value	IC ₅₀
Standard	0.6277	65.688
Sample (methanol extract)	0.4975	311.82

concentrations of standard and methanol extract. This study indicates the methanol extract of plant material has cytotoxic activity.

Thrombolytic Activity

Table 2: Evaluation and results of the thrombolytic activity.

Name of the sample	W1	W2	W3	W4	W5	% of clot lysis
Plant extract	0.742	1.558	1.079	0.347	0.372	75.46
Standard	0.795	1.519	1.272	0.578	0.05	15.90
Blank	0.795	1.478	1.163	0.468	0.115	23

Here, W1 = Micro-tube weight, W2 = Clot with micro-tube weight, W3 = Clot with micro-tube weight after clot disruption, W4 = Clot weight after clot disruption, W5 = Released clot weight. Plasminogen enzyme is usually activated by thrombolytic agents and it also removes fibrin bonds in blood, as a result, the clot becomes soluble and blood flow is restored. Here, methanol extract showed much lower level of thrombolytic activity than standard. Standard

gave 15.90% clot lysis, distilled water was used as a negative control, which provided 23% clot lysis and methanol extract of plant leaves showed 75.46% clot lysis. After comparing the clots lysis value of plant extract with the positive control value, it was observed that plant material revealed thrombolytic activity but less than standard.

Antimicrobial Assay

Table 3: Antimicrobial activity of the leaves of *Callicarpa americana*.

Group	Inhibition zone (mm)	
	Gram (-ve) bacteria (<i>E.coli</i>)	Gram (+ve) bacteria (<i>B. subtilis</i>)
Control	0.00	0.00
Standard	23.657 ± 2.082	19.3±1.58
Plant extract (200mg/mL)	0.00	0.00
Plant extract (400mg/mL)	12.333±1.17	24±1

The plant extract showed antimicrobial activity at all concentrations tested with a broad spectrum of activity,

inhibiting against the growth of both Gram positive and Gram-negative bacteria. The antimicrobial potential was

especially showed against *E. coli* and *B. subtilis*, when exposed to 400 mg/mL of methanol extract of plant and made it impossible when exposed to 200 mg/mL of methanol extract of dried leaves as shown in the table. These results indicate that the antimicrobial activity of the plant extract is not as significant as standard.

Conclusion

The plant has been brought into effective action in various traditional uses of which some have been proved clinically. Further studies can be conducted on secondary metabolites to explore more activities. This review can be helpful in promoting research that can help to develop new agents for therapeutic applications based on bioactive chemical compounds. Therefore, this plant is significantly used for the treatment and prevention of diseases.

References

1. Medicinal Plants of Bangladesh. Assignment Point. (n.d.), 2019. Retrieved March 21, from <http://www.assignmentpoint.com/science/zoology/medicinal-plants-bangladesh.html?fbclid=IwAR1aSAF9it14R8g59AoTsmYuOz4ZmULVqpkYwFnTKi8W6XvZP27r6N-77MY>
2. Muqaddim M. Phytochemical and Biological Investigation of Kaempferia galangal Leaves, 2017.
3. Persicaria Glabra. (Willd.). (n.d.). Retrieved March 21, 2019, from <http://www.mpbdb.info/plants/persicaria-glabra.php?fbclid=IwAR3bovNH1cwZUZUP5AKqMwaG7-MuxPTYV09fXqDzCFAXCnEmyHj4IHsfuBk>
4. Rafieian-Kopaei M. Medicinal plants and the human needs. Journal of Herb Med Pharmacology Journal J Herb Med Pharmacol. 2012; 1(1):1-2. [https://doi.org/10.1016/S2222-1808\(14\)60708-8](https://doi.org/10.1016/S2222-1808(14)60708-8)
5. Rasool Hassan BA. Medicinal Plants (Importance and Uses). Pharmaceutica Analytica Acta. 2013; 3(10):4172. <https://doi.org/10.4172/2153-2435.1000e139>
6. Abdille MH, Singh RP, Jayaprakasha GK, Jena BS. Antioxidant activity of the extracts from Dillenia indica fruits. Food Chem. 2005; 90:891-896.
7. Shome U, Khanna KR, Sharma PH. Pharmacognostic studies on Dillenia indica Linn. Leaf. Proc. Indian Acad. Sci. 1979; 88B(1):35-48.
8. Haque ME, Islam MN, Hossain M, Mohamad AU, Karim MF, Rahman MA. Antimicrobial and Cytotoxic activities of Dillenia pentagyna. Dhaka Univ. J Pharm. Sci. 2008; 7(1):103-105.
9. Parvin MN, Rahman MS, Islam MS, Rashid MA. Chemical and biological investigations of Dillenia indica Linn. Bangladesh J Pharmacol. 2009; 4:122-125.
10. Yeshwante SB, Juvekar AR, Nagmoti DM, Wankhede SS, Shah AS, Pimprikar RB *et al.* Anti-inflammatory activity of methanolic extracts of *Dillenia indica* L. leaves. Pharmacology. 2009; 1(1):63-66.
11. Ahmed F, Al Mamun AH, Shahid IZ, Rahman AA, Sadhu SK. Antinociceptive, antidiarrhoeal and cytotoxic activity of *Aegiceras corniculatum*. Orient Pharm Exp Med. 2007; 7(2):191-196.
12. Roome T, Dar A, Naqvi S, Ali S, Choudhary MI. A study on antioxidant, free radical scavenging, anti-inflammatory and hepatoprotective actions of *Aegiceras corniculatum* (stem) extracts. J Ethnopharmacol. 2008; 118:514-521.
13. Akah PA, Aguwa CN, Agu RU. Studies on the antidiarrhoeal properties of *Pentaclethra macrophylla* leaf extracts. Phytother Res. 1999; 13(4):292-295.
14. Taesotikul T, Panthong A, Kanjanapothi D, Verpoorte R, Scheffer JJC. Anti-inflammatory, antipyretic and antinociceptive activities of *Tabernaemontana pandacaqui* Poir. J Ethnopharmacol. 2003; 84:31-33.
15. Khatun A, Rahman M, Haque T, Rahman MM, Akter M, Akter S *et al.* Cytotoxicity Potentials of Eleven Bangladeshi Medicinal Plants. Scientific World J. 2014, 1-7.
16. Tania UH, Hassan MR, Eshita NJ, Akhter R, Shahriar M. Evaluation of *In vitro* Antioxidant and *In vivo* Pharmacological Activity of Leaf Extracts of *Hoya parasitica* (Wall.). J App Pharm Sci. 2016; 6(05):163-170.
17. Tiwari P, Kumar B, Kaur M, Kaur G, Kaur H. Phytochemical screening and extraction: A review. IPS. 2011; 1:103-104.
18. Prasad S, Kashyap RS, Deopujari JY, Purohit HJ, Taori GM, Dagainawala HF. Development of an *in vitro* model to study clot lysis activity of thrombolytic drugs. Thromb J. 2006; 4:14.
19. Bakhtiar MSI, Shahriar M, Akhter R, Bhuiyan MA. *In vitro* antioxidant activities of the whole plant extract of *Chrozophora prostrata* (dalz.). Ann Biol Res., 2015; 6(4):19-26.
20. Zheng W, Wang SY. Antioxidant activity and phenolic compounds in selected herbs. J Agric Food Chem. 2001; 49(11):5165-5170.