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Antioxidant properties of certain South Indian medicinal plants

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Abstract

Polyphenols and flavonoids are the most abundant compounds in nature. They are strong antioxidants. Antioxidant plays an important role in inhibiting and scavenging free radicals, thus providing protection to human against infection and degenerative diseases. Now the modern research is directed towards "Natural antioxidants" from the herbal plants due to safe therapeutic. (Abdul Quiyum Ansari et al., 2013). In the present paper antioxidant study of ethanolic extracts of six medicinal plants, namely *Gynodropsis pentaphyllum* (Capparidaceae family), *Solanum nigrum* (Solanaceae family), *Merremia gangetica* (Convolvulaceae family), *Cicca acida* (Euphorbiaceae family), *Erythrina variegata* (fabaceae family) and *Asparagus fysoxii* (Asparagaceae family) has been made for its free radical scavenging ability by adopting various *in vitro* methods. The extracts from these plants were investigated for the antioxidants property using 2,2- diphenyl-1-picryl hydrazyl (DPPH) radical scavenging method and the Ferric Reducing Ability of Plasma (FRAP) activity method.

Keywords: Medicinal plants, Antioxidant, DPPH, Anti radical, Gp, Sn, Mg, Ca, Ev and Af.

1. Introduction

In recent years growing evidence has been accumulated indicating the involvement of reactive oxygen species (ROS) in the pathogenesis of a number of diseases. Among the cellular and extracellular components, lipids, proteins, enzymes, DNA and RNA form the major targets for these reactive species, and the resulting damages are associated with degenerative ("Oxidative") diseases. Most living organisms possess efficient enzymatic and non-enzymatic defense systems against excess production of ROS. However, different external factors (smoke, diet, alcohol, some drugs) and aging decreases the capacity of protecting systems, resulting in disturbance of the redox equilibrium that is establishing in healthy conditions. Therefore, antioxidants that scavenge ROS may be great value in preventing the onset and or the progression of oxidative diseases.

Over the past few years, a number of medicinal plants have been investigated for their quenching activity of specific ROS. Interestingly, these medicinal plants contain large amounts of antioxidants other than vitamin C, vitamin E and carotenoids. The antioxidative effect is mainly due to phenolic components, such as flavonoids, phenolic acids and phenolic diterpenes. They are also a good source of dietary fiber, proteins and minerals and thus, serve as a rich source of bioactive constituents. Polyphenols, especially the flavonoids, exhibit a spectacular array a biological effects as a consequence of their antioxidant properties, with mechanisms involving both free radical scavenging and metal chelation and have been demonstrated to be responsible for several pharmacological activities claimed by folkloric traditions. Therefore, it is felt that the investigation of the antioxidant properties of the phenolic constituents of medicinal plants like *Gynodropsis pentaphyllum*, *Solanum nigrum*, *Merremia gangetica*, *Cicca acida*, *Erythrina variegata* and *Asparagus fysoxii* has become a vital task.

2. Materials and Methods

2.1 Collection of plant materials

Fresh aerial parts of six medicinal plants viz. *Gynodropsis pentaphyllum*, *Solanum nigrum*, *Merremia gangetica*, *Cicca acida*, *Erythrina variegata* and *Asparagus fysoxii* we collected in and around of Puducherry during the month of April 2013 and authenticated by the department of Botany K.M Centre for PG studies, Puducherry, where a voucher specimens of each plants were deposited.

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2.2 Preparation of Plant extracts

The air dried aerial parts of the six medicinal plants each of 500 g collected from in and around Puducherry were extracted thrice with hot 95% ethanol under reflux (3×5 litre). These extracts were concentrated in vacuo. 10 ml of the concentrated ethanolic extracts of each plant was used for the antioxidant studies. The different concentrations of the ethanolic extract were used for both DPPH and FRAP assay method.

3. Determination of antioxidant Capacity

3.1 DPPH Method

Free radical scavenging ability by the use of stable DPPH radical 2, 2-diphenyl-1-picryl hydrazyl. The DPPH radical scavenging activities of ethanolic extracts of six medicinal plants viz., *Ca*, *Af*, *Ev*, *Sn*, *Gp* and *Mg* were determined using the method proposed by Von Credow, Jaibert and Hansmann (1997) [2]. In this method 0.1 ml of ethanolic extract of each plant is added to 2.9 ml of 0.1M DPPH solution. The negative control prepared by mixing 0.1 ml of methanol with 2.9 ml of 0.1 M DPPH solution. The samples were kept in the dark for 30 minutes and after that the absorbance measured at 517 nm. The percentage inhibition of DPPH radical by the samples was calculated according to the formula of Yen and Dush (1994)

$$\% \text{ of Inhibition of DPPH assay} = \frac{AB - AA}{AB} \times 100$$

AB – Absorbance of control

AA – Absorbance of Test

3.2 Determination of Ferric Reducing/antioxidant power

The total antioxidant potential sample was determined using a ferric reducing ability of plasma (FRAP assay) of Benzie and Strain (1996) as a measure of antioxidant power. FRAP assay measures the change in absorbance at 700 nm owing to the formation of a blue coloured Fe^{II} – tripyridyltriazine compound from the colourless oxidized Fe^{III} form by the action of electron denoting antioxidant.

The aliquot of various concentrations of the test sample extracts ($100\mu\text{g/ml}$) were mixed with 2.5 ml of (PH 6.61) phosphate buffer and 2.5 ml of 1% potassium ferricyanide. The mixture was incubated at 50°C in water both for 20 minutes. After cooling, aliquots of 2.5 ml of (10%) trichloroacetic acid were added to the mixture, which was then centrifuged at 300 rpm for 10 min. The upper layer of solution (2.5 ml) was mixed with 2.5 ml distilled water and a freshly prepared 0.5 ml of (0.1%) ferric chloride solution.

The absorbance was measured at 700 nm in a UV visible spectrometer. A blank was prepared without adding the extract. Increased absorbance of the reacting mixture indicates increase of reducing capacity.

4. Results and Discussion

The antioxidant activity of the extract of the leaves was assessed by spectrophotometry in the presence of the DPPH radical, which is often used to compare the activities of ethanolic extracts of plants. DPPH radical is a stable free radical which dissolves in methanol and shows characteristic absorption at 517 nm, when an antioxidant scavenges free radicals by hydrogen donation, the DPPH solution become lighter in colour (Molynux, 2004; Villiano *et al.*, 2007). Samples with a raw material concentration of $25\mu\text{g/ml}$ were analyzed.

Table 1: Absorbance of the ethanolic extract of plants at 517 nm

| EXTRACT | ABSORBANCE | SCAVENGING % (DPPH) |
|---------|------------|---------------------|
| Ca | 0.183 | 42.81 |
| Af | 0.295 | 7.81 |
| Ev | 0.304 | 5.00 |
| Sn | 0.261 | 18.44 |
| Gp | 0.313 | 2.19 |
| Mg | 0.127 | 60.31 |

Tabular column shows the antioxidant activities of the extracts tested as the percentage of deactivation of the DPPH free radicals. The quality of antioxidants in the ethanolic extracts was determined by the absorbance and scavenging percentage. The DPPH method revealed that the scavenging of the free radicals was found to be highest for the ethanolic extract of *Merremia gangetica* (60%) followed by *Cicca acida* (42.8%) and *Solanum nigrum* (18.4%) respectively.

Antioxidant activities of the ethanolic extracts of the plants determined by FRAP are depicted in Tabular column II. All polyphenols reduce Fe^{3+} ions to Fe^{2+} which are complex TPTZ at a concentration of 10 to $100\mu\text{g/ml}$. The antioxidant activities of ethanolic extracts of six medicinal plants have greater capacity of antioxidants than the antioxidant activities of standard Ascorbic acid. It was proved by a FRAP assay method that the absorbance of *Merremia gangetica* was found to be maximum (0.234) followed by *Asparagus fysoxii* absorbance (0.148), *Cicca acida* absorbance (0.141) and *Erythrina variegata* absorption (0.134) respectively at the concentration of $\frac{100\mu\text{g}}{\text{ml}}$. Similarly 80 mcg/ml , 60 mcg/ml , 40 mcg/ml And 20 mcg/ml were also found to be greater values of absorbance than the standard drug Ascorbic acid. Out of six medicinal plants the antioxidant capacity of the plant *Merremia gangetica* was found to be greater than the standard Ascorbic acid at the concentration of 100 mcg/ml .

Table 2: Absorbance of plant extracts and standard ascorbic acid at 700 nm

| Concentration in mcg/ml | Standard Ascorbic acid | Absorbance at 700 nm | | | | | |
|-------------------------|------------------------|----------------------|-------|-------|-------|-------|-------|
| | | Ca | Mg | Gp | Sn | Ev | Af |
| 20 | 0.083 | 0.106 | 0.098 | 0.128 | 0.093 | 0.098 | 0.096 |
| 40 | 0.088 | 0.099 | 0.124 | 0.106 | 0.106 | 0.092 | 0.104 |
| 60 | 0.090 | 0.119 | 0.164 | 0.091 | 0.153 | 0.107 | 0.117 |
| 80 | 0.108 | 0.114 | 0.196 | 0.076 | 0.153 | 0.105 | 0.131 |
| 100 | 0.145 | 0.141 | 0.234 | 0.098 | 0.134 | 0.098 | 0.148 |

The reducing capacity of the ethanolic extracts of *Gynodropsis pentaphyllum* and *Cicca acida* were found to be on par with that of the standard antioxidant Ascorbic acid at 100 mcg/ml concentration.

5. Conclusion

In the present study, the antioxidant capacities of the plant extracts were analyzed using free radicals scavenging activity (DPPH) and Ferric reducing ability of plasma (FRAP) methods as a measure of antioxidant power.

The DPPH test is the oldest indirect method of determining the antioxidants activity based on the ability of free radicals 2,2 – diphenyl – 1 – picrylhydrazyl to react with hydrogen donors with phenol. The ethanolic extracts of all the six medicinal plants have the ability to scavenge the free radicals. Out of the six medicinal plants *Merremia gangetica* is found to be with a 60% scavenging activity followed *Cicca acida* 42.8% and *Solanum nigrum* 18.4% respectively.

The antioxidant capacity of the plant extracts *Merremia gangetica* and *Asparagus fysoxii* under study is found to be greater than standard Ascorbic acid at the concentration of 20 mcg/ml, 40 mcg/ml, 60 mcg/ml, 80 mcg/ml and 100 mcg/ml than the other medicinal plants that have desirable values of antioxidant capacity at different concentration. Ethanolic extracts of leaves of six medicinal plant species are found to be good antioxidants as proved by the FRAP assay method. Ethanol is the best solvent for preparing plant extracts for yielding strongest antioxidant activity in the extract.

From the above methods it is found that the antioxidant capacity of the plant extracts have the maximum ability to scavenge the free radicals is useful to cure diabetes to some extent with a reducing level of the oxidative stress. It has been found that the intake of antioxidant rich diet is effective in reducing the deleterious effects of aging and behaviour.

Several South Indian medicinal plants tested are rich sources of polyphenolic compounds which are free radical scavengers. Some medicinal plants, thus can be considered as promising sources of natural antioxidants for medicinal and commercial use.

6. References

1. Abdul QA, Syed AA, Waheed MA, Sayyed J. A Extraction and Determination of antioxidant activity of *Withania somnifera* Dunal”, Pelagia Research Library 2013; 3(5):502–507.
2. Von GA, Joubert E, Hansmann CF. Comparison of antioxidant activity of aspalathin with that of other plant phenols of Rooibosed tea (*Aspalathus linearis*), α – tocopherol, BHT, and BHA, Journal of Agricultural and Food Chemistry 1997; 45:632–638.
3. Benzie IF, Strain JJ. The ferric reducing ability of plasma (FRAP) as measure of antioxidant power”, The FRAP assay. Anal. Biochem 1996; 239:70–76.
4. Molyneux P. The use of the free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. Songklankarin Journal of Science and Technology 2004; 25:211–219.
5. Villano D, Fernandez–Pachon MS, Moya ML, Troncoso AM, Garcia–Parrilla MC, Radical scavenging ability of polyphenolic compounds towards DPPH free radical. Talanta 71, 230–235.
6. Ahmad, S, Oxidative stress and antioxidant defenses in biology, New York; Chapman & Hall, 1995.
7. Benzie IFF, Wai Y, Strain JJ, Antioxidant (reducing) efficiency of ascorbate in plasma is not affected by concentration. Journal of Nutritional Biochemistry 1999; 10:146–150.
8. Frankel EN, Waterhouse AL, Teissedre PL. Principal phytochemicals in selected California wines and their antioxidant activity in inhibiting oxidation of human low – density lipoproteins. Journal of Agricultural and Food Chemistry 1986; 43:890–894.
9. Friedman LA, Kimball AW. Coronary heart diseases mortality and alcohol consumption in Framingham. American Journal of Epidemiology 1997; 24:481–489.
10. Vogiatzi G, Tousoulis D, Stefandis C. The roll of oxidative stress in atherosclerosis, Hellenic. J. Cardiol 2009; 50:402–409.
11. Assimopoulou AN, Sinakos Z, Papageorgiou VP. Radical scavenging activity of *Crocus sativus* L. extract and its bioactive constituents, Phytotherapy Research 2005; 19:997–1000.
12. Rice–Evans CA, Miller NJ, Paganga G. Structure antioxidant activity relationships of flavonoids and phenolic acids, Free radical biology and medicine, 20, 933–956.
13. Venskultonis PR, Dvaranauskaite A, Labokas J. Radical Scavenging activity and composition of raspberry (*Rubusidaeus*) leaves from different location in Lithuania, Fitoterapia 2007, 162–165.
14. Aruoma O, Cuppett SL. Antioxidant methodology *in vivo* and *in vitro* concepts. APCS press Champaign, 1997, 41–172.
15. Ruby G, Muktasharma, Ramakrishnan I *et al.* Improved method of total antioxidant assay, Indian journal of Biochemistry and Biophysics 2009; 46:126–129.
16. Gunars T, Gregorzbartosz. Determination of antiradical and antioxidant activity; basic principles and new insights, ACTA ABP Biochimica Polonica 2010; 57:139–142.
17. Gohari AR, Hajimehdipoor H, Saeidnia S *et al.*, “Antioxidant activity of some medicinal species using FRAP assay. Journal of medicinal plants, 2011, 37.
18. Maria K, Petko D, Milan C, Antonin L *et al.* Evaluation of antioxidant activity of medicinal plants containing polyphenol compounds. Comparison of two extraction systems, ACTA ABP Biochimica Polonica 2010; 57(2):229–234.