



ISSN (E): 2320-3862
ISSN (P): 2394-0530
NAAS Rating: 3.53
JMPS 2018; 6(1): 67-70
© 2018 JMPS
Received: 12-11-2017
Accepted: 13-12-2017

Noor Fatmawati Mokhtar
Institute for Research in
Molecular Medicine
(INFORMM), Universiti Sains
Malaysia, Health Campus, 16150
Kubang Kerian, Kelantan,
Malaysia

Centella asiatica aqueous extract inhibits motility of VGSCs-expressing MDA-MB-231 cells without affecting cell growth

Noor Fatmawati Mokhtar

Abstract

Targeting nNav1.5, a potent metastasis marker in breast cancer using VGSCs blockers founded in a 'back-to-nature' strategy might improve treatment avenues with fewer or no undesirable side effects. The potential of *Centella asiatica* aqueous extract to inhibit cell motility was examined on the strongly metastatic breast cancer cell line MDA-MB-231 cells. Cells were treated with *Centella asiatica* aqueous extract ranging from 0.1-10 μ g/ml for 24h followed by total RNA extraction, cDNA synthesis and PCR. MTT and motility assays were conducted to measure the effect of the plant extract on growth and cell motility. Growth of MDA-MB-231 cells was not affected by any tested concentration (0.1- 10 μ g/ml) of *Centella asiatica* aqueous extract but importantly, motility of the cells was suppressed significantly in a dose-dependent manner followed by down-regulation of the nNav1.5 gene expression was observed. Overall, *Centella asiatica* aqueous extract inhibit cell motility without affecting cell growth via nNav1.5 gene down-regulation.

Keywords: *Centella asiatica*, anti-metastatic, lateral motility, voltage-gated sodium channels, nNav1.5

1. Introduction

Among various recently identified metastasis-related genes, voltage-gated sodium channels (VGSCs) has consistently been highlighted. Although VGSCs is 'classically' abundant in excitable cells such as the neuron and muscle cells where its role is important in mediating cell regenerative membrane depolarization and conduction of electrical signaling (action potentials) ^[1]. However, recent discoveries have shown that VGSCs are also highly expressed in metastatic cancer cells.

Particularly in breast cancer, high VGSCs expression and activity correlate positively with its metastatic potential ^[2]. A whole-patch clamp recordings from a variety of human breast epithelial cells ranging from the normal to highly metastatic cell lines demonstrated that metastatic potential increases from the non-cancerous human breast epithelial cells, MCF-10A < weakly metastatic breast cancer cell line, MCF-7 < aggressive breast cancer cell line, MDA-MB-468 < strongly aggressive breast cancer cell line, MDA-MB-231. Only the strongly metastatic, MDA-MB-231 cells generate a fast inward current, representing high VGSCs activity ^[2]. Using PCR, high level of VGSCs mRNA expression particularly of the splice variant 'neonatal' Nav1.5 (nNav1.5) was detected in MDA-MB-231 cells. When these cells were treated with VGSCs specific blocker, tetrodotoxin (TTX), suppression of metastatic parameters was obtained ^[2]. Other research have reported that nNav1.5 can be blocked with other VGSCs modulators e.g local anesthetics and analgesics which altogether supports the potential of VGSCs as target in controlling breast cancer metastasis ^[3, 4, 5, 6, 7, 8].

Centella asiatica also known locally as 'pegaga' is a tropical plant commonly consumed by Malaysians as 'ulam'. In folk medicine, the plant has been claimed to have therapeutic potential to treat numerous conditions e.g wound healing, leprosy, psoriasis, lupus, eczema, varicose ulcers, diarrhoea, amenorrhoea, diseases of the female genitourinary tract and also for relieving anxiety and improving cognition ^[9]. Importantly, *Centella asiatica* has been claimed to present central analgesic activity ^[10]. In one *in vivo* example, asiatic acid, a pentacyclic triterpene compound from *Centella asiatica* was demonstrated to have analgesic mechanisms due to its anti-inflammatory effects in the edema paw of a mice ^[11].

Subsequently, the main purpose of this study was to assess the anti-metastatic effect of *Centella asiatica* aqueous extract *in vitro*, on the lateral motility of strongly aggressive

Correspondence

Noor Fatmawati Mokhtar
Institute for Research in
Molecular Medicine
(INFORMM), Universiti Sains
Malaysia, Health Campus, 16150
Kubang Kerian, Kelantan,
Malaysia

MDA-MB-231 cells. We also analyzed the effects of the plant extract on nNav1.5 expression in order to link the mechanism of motility suppression by the plant extract.

2 Materials and Methods

2.1 Cell culture

Highly metastatic human breast cancer cell line, MDA-MB-231 was cultured in 10cm or 35mm Falcon plastic dishes (Becton Dickinson) and maintained in Dulbecco's minimum essential medium (DMEM) (Nacalai Tesque) supplemented with 4mM L-glutamine (Gibco) and 5% of fetal bovine serum (FBS) (Gibco). Cultured cells were kept in a humidified incubator at 37 °C, 100% relative humidity and with 5% regulated CO₂ [12].

2.2 Identification and preparation of the plant extract

After appropriate identification, washed *Centella asiatica* were shade dried at room temperature for an hour and further dried in a hot-air oven (Mettler) at 50 °C for another 48h. The dried samples were grind to produce fine powder and 50g was placed in a clean flask containing 200ml of aqueous. The flask was placed in a shaker incubator (Max^Q 4000, Barnstead) for 48h at 37 °C and 150rpm. The aqueous extract of *Centella asiatica* was filtered using filter paper (Whatman No.1) and concentrated using rotary evaporator (Rotaver) at 80 °C. The concentrated aqueous extract was further dried for an overnight in oven (Mettler). The extract was weighed and stored at -20 °C until further use.

2.3 Cell viability and proliferation assays

Proliferation was measured using an assay based upon colorimetric quantification of yellow 3-[4, 5-dimethylthiazol-2-yl]-2, 5-diphenyltetrazolium bromide (MTT) (Sigma) reduced to purple formazan in living cells [12].

2.4 Motility assay

Motility of cells was assessed using a monolayer 'wound-heal' assay (Fraser *et al.* 2003). Cells were plated in 35mm dishes at an initial density of 5×10^5 cells/dish and allowed to settle for 24h prior to wound creation and any pharmacological treatment (T₀). Wounds were created by scratching the monolayer cells using a p200 pipette tip at 90° angle. Wound widths were measured again after 24h (T₂₄). Motility was calculated according to $1 - (T_{24}/T_0)$ and percentage of lateral motility was normalized to Control.

2.5 Polymerase chain reaction (PCR)

Total RNA was extracted directly from a ~80% confluent dish of cells using the 'Trizol method' according to the protocol provided by the manufacturer (Invitrogen). Total RNA (1µg) was used to synthesis cDNA according to the QuantiTect Reverse Transcription Kit (Qiagen). PCR was performed using MyTaq HS (Bioline) and primers for nNav1.5: 5'-CTGCACGCGTTCACCTTCCT-3' (F) and 5'-GACAAATTGCCTAGTTTTATATTT-3' (R). β-actin: ATTGCCGACAGGATGCAGAAG-3' (F), 5'-TAGAAGCATTTGCGGTGGACG-3' (R). The intensity of PCR product was measured using ImageJ (free online software: <http://rsbweb.nih.gov/ij/>).

2.6 Statistical analysis

All values are expressed as the mean ± SEM. Student's *t*-test for unpaired results was used to evaluate differences between two groups. Differences were considered to be significant for values of $P < 0.05$.

3 Results

3.1 Effect of *Centella asiatica* aqueous extract on cell viability and growth

Initial observations showed no apparent change in cellular morphology of MDA-MB-231 cells after 24h treatment with aqueous extract of *Centella asiatica* up to 10µg/ml. There was no effect on cell viability and the extract also did not affect cell number ('proliferation') after 24h treatment where, viability/cell number remaining between 98–100% of controls (Figure 1).

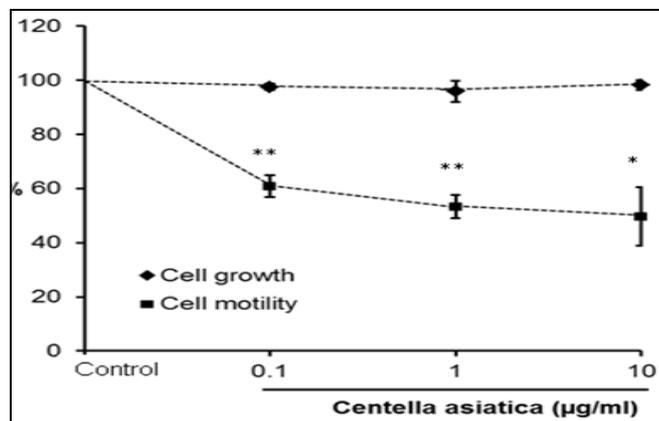


Fig 1: Effect of *Centella asiatica* aqueous extract on cell growth and motility. Cell growth was not affected after 24h treatment with aqueous extract of *Centella asiatica*. However, motility was reduced dose-dependently after 24h treatment with aqueous extract of *Centella asiatica*. Values are mean ± SEM ($n = 3$) with * and ** indicates significant differences $p < 0.05$ and $p < 0.01$, respectively.

3.2 Effect of *Centella asiatica* aqueous extract on motility

Motility was assessed according to the 'wound-heal' assay after 24h treatment with the plant extract (0.1 - 10µg/ml). The effect of *Centella asiatica* aqueous extract on motility was normalized to the control level; results showed that lateral motility was reduced by *Centella asiatica* aqueous extract dose-dependently with highest effect was seen at 10µg/ml which reduced the motility index by ~50% to $49.7 \pm 10.8\%$ (Figure 1).

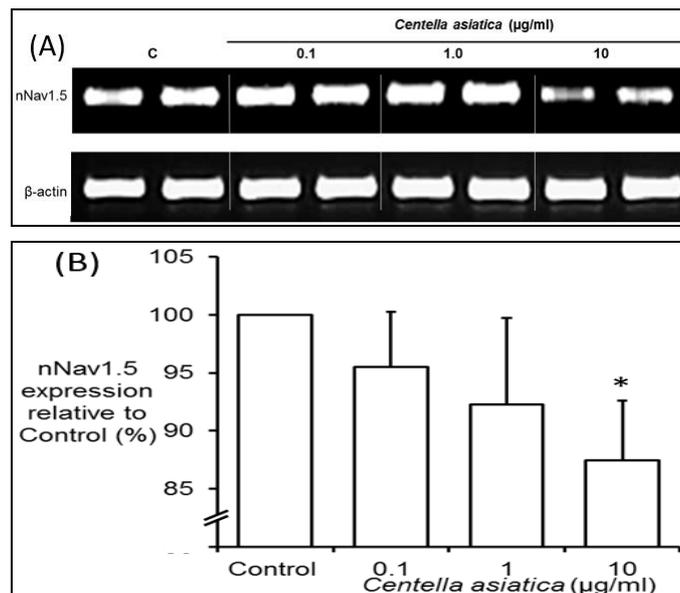


Fig 2: Effect of *Centella asiatica* aqueous extract on nNav1.5 gene expression. (A) PCR products (duplicate) of nNav1.5 and β-actin from of each samples; C – Control, 0.1–10µg/ml of *Centella asiatica* aqueous extract. (B) nNav1.5 expression was reduced dose-dependently after 24h treatment with aqueous extract of *Centella asiatica*. Values are mean ± SEM ($n = 4$) with * indicates significant differences $p < 0.05$

3.3 Effect of *Centella asiatica* aqueous extract on nNav1.5 gene expression level

PCRs were conducted to measure nNav1.5 expression levels after 24h treatment with *Centella asiatica* aqueous extract. Figure 2(A) shows the products of nNav1.5 and β -actin of each sample. In comparison to control (untreated cells), nNav1.5 expression was reduced dose-dependently by *Centella asiatica* aqueous extract (0.1 to 10 μ g/ml). nNav 1.5 gene levels was reduced to $\sim 95 \pm 4.8$, 92 ± 7.4 and $87 \pm 5.2\%$, respectively (Figure 2(B)).

4. Discussions

The preferred future cancer therapy would be to use drugs that is non-toxic but could control cancer metastasis which has been the main cause for 90% of cancer mortality. Therefore, in this study, the cell viability and proliferation assays were primarily aimed to choose concentrations of *Centella asiatica* aqueous extract that would not give effect on growth. This is essential to achieve as to exclude any limitation on the analysis of anti-metastasis, where the effect should be exclusively *via* the suppression of metastatic parameter and not due to suppression of cells proliferation. Indeed, after 24h treatment with *Centella asiatica* aqueous extract ranging from 0.1 to 10 μ g/ml, cell proliferation of MDA-MB-231 cells were not affected (Figure 1). Therefore, tested concentration range remained throughout later experiments. Although the concentration range used in this study was not killing the cells, the cytotoxicity potential of *Centella asiatica* aqueous extract has been shown in other report where similarly, it was tested on MDA-MB-231 cells and the obtained IC₅₀ was at 648 μ g/ml [13].

During metastasis, cell motility enables cancer cell to disseminate away from the primary tumour. Motility or 'wound-heal' assay has been used widely in research investigating or measuring cancer motility potential. After 24h treatment with *Centella asiatica* aqueous extract (0.1 to 10 μ g/ml), lateral motility of MDA-MB-231 cells was suppressed dose-dependently (Figure 1). This result supported our hypothesis; *Centella asiatica* aqueous extract exclusively inhibit the motility of MDA-MB-231 cells at concentrations not affecting the cell growth. Similarly, motility of aggressive prostate cancer cell line expressing Nav1.7, MAT-LyLu was suppressed without effect on cells growth by TTX (1 μ M) [14], highlighting the role of VGSCs in controlling cancer motility and importantly the potential of TTX as anti-metastatic. However, since TTX is very toxic, using 'back-to-nature'-medicinal plants might be a safer alternative to target VGSCs in breast cancer. Indeed, our results indicate that *Centella asiatica* aqueous extract might have the right component to do so with the exact phytochemical component need to be determined.

After 24h treatment with *Centella asiatica* aqueous extract, down-regulation of nNav1.5 gene expression levels was observed (Figure 2(A) & Figure 2(B)), suggesting that suppression of motility by *Centella asiatica* aqueous extract may associate with down-regulation of nNav1.5 gene. Previously, down-regulation of nNav1.5 mRNA expression level by TTX has been well linked with suppression of metastasis parameters such as migration and invasion of MDA-MB-231 cells [7, 15]. Similarly, in prostate cancer expressing VGSCs isoforms namely, Nav1.7, down-regulation of the gene by TTX was followed by suppression of metastasis parameters (migration and invasion) [7]. Interestingly, comparable 'back-to-nature' examples have been reported by docosahexaenoic acids (DHA), a natural

omega-3 polyunsaturated fatty (PUFA) abundant in fish oil; DHA reduced nNav1.5 mRNA and protein expression at plasma membrane hence suppressing migration of metastatic breast cancer cells [16]. Another omega-3 acid PUFA eicosapentaenoic (EPA) have also been reported to have similar observation; down-regulation of VGSC mRNA expression was obtained by EPA followed by inhibition of metastatic potential of prostate cancer cells [17].

5. Conclusions

The adverse effects of chemotherapy can be highly invasive and some breast cancer patients can have low tolerance to treatment. In this regard, targeting VGSCs in breast cancer using VGSC blockers founded in a 'back-to-nature' strategy might improve treatment avenues with fewer or no undesirable side effects. *Centella asiatica* aqueous extract might contain the right component to suppress nNav1.5 expression at the same time non-toxically control breast cancer aggressiveness. However, the exact phytochemical component responsible for its action on nNav1.5 needs to be determined. The effect of the responsible component should also be validated using a standard method to measure ion channel activity e.g. patch-clamps.

6. Acknowledgments

This work was supported and sponsored by the Universiti Sains Malaysia (USM) under the USM Incentive Grant 304/JPNP/600004. We thank Nor Ferdaus Muhamad, Noradlianti Rusli, Amira Raudhah Abdullah, Nur Anis Hanim Mohamad Azemi, Nor Hidayah Noh and Elis Rosliza Mohd Adzmi for the experimental works. We also thank Nik Soriani Yaacob's group for supplying the MDA-MB-231 cells.

7. References

1. Catterall WA. From ionic currents to molecular mechanism: the structure and function of voltage-gated sodium channels. *Neuron*. 2000; 26(1): 13-25.
2. Fraser SP, Diss JK, Chioni AM, Mycielska ME, Pan H, Yamaci RF *et al.* Voltage-gated sodium channel expression and potentiation of human breast cancer metastasis. *Clinical Cancer Research*. 2005; 11(15):5381-5389.
3. Roger S, Besson P, Le Guennec JY. Involvement of a novel fast inward sodium current in the invasion capacity of a breast cancer cell line. *Biochimica et Biophysica Acta (BBA) – Biomembranes*. 2003; 1616(2):107-111.
4. Roger S, Besson P. Influence of the whole-cell patch clamp configuration on the electrophysiological properties of the voltage-dependent sodium current expressed in MDA-MB-231 cells. *European Biophysiology Journal*. 2004; 33(3):274-279.
5. Roger S, Potier M, Vandier C, Besson P, Le Guennec JY. Voltage-gated sodium channels: new targets in cancer therapy? *Current Pharmaceutical Design*. 2006; 12(28):3681-3695.
6. Huang CJ, Harootunian A, Maher MP, Quan C, Raj CD, McCormack K *et al.* Characterization of voltage-gated sodium-channel blockers by electrical stimulation and fluorescence detection of membrane potential. *Nature Biotechnology*. 2006; 24(4):439-446.
7. Brackenbury WJ, Chioni AM, Diss KJ, Djamgoz MB. The neonatal splice variant of Nav1.5 potentiates *in vitro* invasive behaviour of MDA-MB-231 human breast cancer cells. *Breast Cancer Research & Treatment*. 2007;

- 101(2):149-160.
8. Onkal R, Djamgoz MBA. Molecular pharmacology of voltage-gated sodium channel expression in metastatic disease: clinical potential of neonatal Nav1.5 in breast cancer. *European Journal of Pharmacology*. 2009; 625(1-3):206-219.
 9. Gohil KJ, Patel JA, Gajjar AK. Pharmacological review on *Centella asiatica*: A potential herbal cure-all. *Indian Journal of Pharmaceutical Science*. 2010; 72(5):546-556.
 10. Almeida RN, Navarro DS, Barbosa-Filho JM. Plants with central analgesic activity. *Phytomedicine*. 2001; 8(4):310-322.
 11. Huang SS, Chiu CS, Chen HJ, Hou WC, Sheu MJ, Lin YC *et al*. Antinociceptive activities and the mechanisms of anti-inflammation of asiatic acid in mice. *Evidence-Based Complementary and Alternative Medicine*. 2011; 895857.
 12. Grimes JA, Fraser SP, Stephens GJ, Downing JE, Laniado ME, Foster CS *et al*. Differential expression of voltage-activated Na⁺ currents in two prostatic tumour cell lines: contribution to invasiveness *in vitro*. *FEBS Letters*. 1995; 369(2-3):290-294.
 13. Pittella F, Dutra RC, Junior DD, Lopes MTP, Barbosa NR. Antioxidant and cytotoxic activities of *Centella asiatica* (L). *International Journal of Molecular Sciences* 2009; 10(9):3713-3721.
 14. Fraser SP, Salvador V, Manning EA, Mizal J, Altun S, Raza M *et al*. Contribution of functional voltage-gated sodium channel expression to cell behaviors involved in the metastatic cascade in rat prostate cancer: I. Lateral motility. *Journal of Cellular Physiology*. 2003; 195(3):479-487.
 15. Chioni AM, Shao D, Grose R, Djamgoz MB. Protein kinase A and regulation of neonatal Nav1.5 expression in human breast cancer cells: activity-dependent positive feedback and cellular migration. *The International Journal of Biochemistry & Cell Biology*. 2010; 42(2):346-358.
 16. Isbilen B, Fraser SP, Djamgoz MBA. Docosahexaenoic acid (omega – 3) blocks voltage-gated sodium channel activity and migration of MDA-MB-231 human breast cancer cells. *The International Journal of Biochemistry & Cell Biology*. 2006; 38(12):2173-2182.
 17. Nakajima T, Kubota N, Tsutsumi T, Oguri A, Imuta H, Jo T *et al*. Eicosapentaenoic acid inhibits voltage-gated sodium channels and invasiveness in prostate cancer cells. *British Journal of Pharmacology*. 2009; 156(3):420-431.