



ISSN (E): 2320-3862
ISSN (P): 2394-0530
NAAS Rating: 3.53
JMPS 2019; 7(2): 35-38
© 2019 JMPS
Received: 17-01-2019
Accepted: 21-02-2019

V Priya

Assistant Professor,
Department of Botany,
PSG College of Arts & Science,
Coimbatore, Tamil Nadu, India

PV Anjana

PG Student, Department of
Botany, PSG College of Arts &
Science, Coimbatore,
Tamil Nadu, India

Phytochemical screening and antimicrobial activity in leaves of *Gnetum ula*, Brongn

V Priya and PV Anjana

Abstract

The aim of the present study is to investigate the presence of phytochemical compounds and antimicrobial activities in the leaves of *Gnetum ula* using methanolic and hexane extract. Various concentrations of extracts have been used against the clinical microbes (fungal strains: *Aspergillus niger*, *penicillium* sp. and bacterial strains: *Escherichia coli*, *Serratia* sps. and *Bacillus subtilis*) using standard methods. Phytochemical screening in both extracts reveals the presence of alkaloids, flavonoid, glycoside, phenol, phytosterol, tannins, quinine, carbohydrate, protein and absence of saponin and triterpenoids. Antimicrobial studies indicate that the methanolic and hexane extracts inhibit the growth of microbes, but it may get altered due to different concentration levels. The study exhibits higher inhibition against the fungal strains than the bacterial strains.

Keywords: *Gnetum ula*, phytochemical screening, anti-microbial

Introduction

Gnetum ula is considered as sacred plant by different regions of India. It is a large evergreen woody climber with scaly bark. Leaves are ovate, oblong or elliptic in shape and are arranged in opposite manner. Flowers are monoecious, whorled in the axils of copular bracts in panicle spikes. Fruit is a drupe and reddish orange when ripened. Seed is solitary, ellipsoid and hard. Flowering and Fruiting time is December to June. Mainly it is found in Western Ghats near Khandala, Kerala, Nilgiris, Godavari district of Andhra Pradesh and Orissa. Seeds are used for edible purpose either roasted or boiled and the seed oil is used in rheumatism by folklore practitioners. Stem and leaf extracts are useful in jaundice and liver enlargement, while leaf paste is applied externally cures arthritis.

Natural compounds or phytochemicals play an important role in medicinal field. Plants-derived substance have recently become of great interest owing to their versatile application. Plants are the richest bio-resource of traditional system of medicine, food supplements, and chemical entities for synthetic drug. Today gymnosperms plants were commonly grown in garden only for ornamentation apart from this gymnosperm plants were also used as medicine. Microbial resistance is a world concerned problem. Efforts are being made to discover new antimicrobial agents from various sources. Constant research investigations may result in the discovery of novel effective agents. This study revealed that the plant extracts possessed bioactive compounds that have antibacterial and antifungal infectious diseases.

Materials and Methods

The plant materials are collected from Sree Andalur Kavu located nearly 20km away from Kannur town of Kannur district, Kerala. And leaves are shade dried and powdered. Both the extracts were prepared by using soxhlet method.



Gnetum ula -Habitat

Correspondence

V Priya

Assistant Professor,
Department of Botany,
PSG College of Arts & Science,
Coimbatore, Tamil Nadu, India

Preliminary Phytochemical Analysis

Test for alkaloids

1ml of extract in two separate test tubes, 2-3 drops of Dragendorff's, Wagner's reagent and Mayer's reagents were separately added.

Test for flavonoids

1 ml of extract 1ml of ferric chloride is added.

Test for glycosides

Keller Kilimi Test: 2ml of extract 1ml of glacial acetic acid with ferric chloride and concentrated H₂SO₄ was added.

Test for saponins

1 ml of extract was taken in a test tube and 5 ml of distilled water was added and vigorously shaken.

Test for tannins

2ml of the extracts were diluted with distilled water in separate test tubes; 2-3 drop of 5% ferric chloride (FeCl₃) solution was added.

Test for steroids

2ml of the extract were taken in separate test tubes and evaporated to dryness. The residues were dissolved in acetic anhydride and then chloroform was added. Conc. H₂SO₄ was added by the side of the test tube.

Test for terpenoids

5ml of extract were mixed with 2ml of chloroform and con. H₂SO₄ to form a layer.

Test for tri-terpenoids

Chloroform Test: To 1ml of extract, few drops of chloroform were added and it was treated with concentrated sulphuric acid solution.

Test for phenols

5 ml of concentrated extracts were taken and 2ml of neutral ferric chloride solution was added.

Test for quinone

1 ml of extract added with 5 ml of conc. HCl

Test for Cardiac glycosides

Keller killani Test: To 1 ml of extract add 2 ml of glacial acetic acid and few drops of 5% ferric chloride solution. Then it was treated with few drops of concentrated sulphuric acid.

Test for Proteins

Biuret test: To 1 ml of the test solution, few drops of 0.7% copper sulphate solution and 1 ml of 10% NaOH were added and mixed thoroughly

Test for Carbohydrates

Benedict's test: To 1ml of extract, add 2ml of Benedict's reagent and heated in boiling water bath for 5minutes.

Antimicrobial Activity

To evaluate the antimicrobial activity of selected bacterial, that is, *Escherichia coli*, *Bacillus subtilis* and *Serratia spp.* and fungal strains, that is, *Aspergillus niger*, *Penicillium* by using plant extract and fractions analyzed by disc diffusion method (NCCLS, 1997), minimum inhibitory concentration (MIC) with some modification using method described by

Sarker *et al.*, (2007) [8].

Results and Discussion

1. Preliminary phytochemical analysis

Preliminary phytochemical analysis of hexane and methanol extract of *G. ula* leaves showed the presence of alkaloid, flavonoid, glycoside, phenol, phytosterol, tannins, quinine, carbohydrate and protein. And both the extracts show negative results for saponin and triterpenoids. The results are shown in Table. 1.

Table 1: Phytochemical screening of methanol and hexane leaf extract of *G. ula*

S. No	Name of the compound	Hexane	Methanol
1	Alkaloid	+	+
	Mayer's test		
	Dragendorff's test	+	+
	Wagner's test	+	+
2	Flavonoides	+	+
3	Cardiac glycoside	-	+
4	Glycoside	+	+
5	Terpenoides	-	+
6	Saponin	-	-
7	Phenol	+	+
8	Phytosterol	+	+
9	Triterpenoids	-	-
10	Tannins	+	+
11	Quinone	+	+
12	Carbohydrate	+	+
13	Protein	+	+

Gnetum ula has a medicinal value due to the presence of phytoconstituents. These phytoconstituents make the plant useful for treating different ailment and have a potential of providing useful drugs of human use (Gourav, *et al.*, 2013) [9]. Yogesh Kumar and Krishna Swamy, 2013, reported that the bark of *G. ula* show the presence of alkaloids, flavonoids, phenols and tannins. The phytochemical analysis of the leaves of *G. africanum* shows the presence of alkaloid, tannin, sterol, saponin, flavonoid and terpenoid (Chinyere Ilodibia, *et al.*, 2015) [10]. Flavonoids are most commonly known for their antioxidant activity and they possess anti-cancerous, anti-inflammatory and antimicrobial activity (Balch and Balch, 2000), alkaloids are known to have antipyretic properties, saponins have antifungal properties (Ogu, *et al.*, 2012), and similarly phenols are known to inhibit the mutagenesis of cell DNA (Heinonen, *et al.*, 1998).

2. Antimicrobial study

Antimicrobial activities of methanol and hexane extracts of *G. ula* leaves were studied at different concentrations (25,50,75,100 mg/ml) against three human pathogenic bacterial strains (*E. coli*, *Serratia spp.*, *Bacillus subtilis*) and two fungal strains (*Aspergillus niger* and *Penicillium sp.*). Antimicrobial potentials of extracts were assessed in terms of zone of inhibition of microorganism's growth. The results are shown in table 2-3. The higher antibacterial activity is exhibited by the methanol extract than the hexane, in which the zone of inhibition is higher in *E. coli* in 100mg/ml in methanolic solvent (12mm). In the case of antifungal study also methanolic extract show higher zone of inhibition in penicillium in 100 mg/ml (22mm).

In the study Chinyere Ilodibia, *et al.*, 2015 [10] antimicrobial activity of *G. africanum* leaves also exhibit antifungal property than the antimicrobial activity like the present study shows. The antimicrobial activity of *G. gnemon* in Adolf Jan

Nexson Parhusip and Azis Boing Sitanggang, 2011 explained that in correlation with the present study the methanolic

extract shows more zone of inhibition than the hexane extract.

Table 2: Antibacterial activity of *Gnetum ula* leaves in different solvents

S. No	Name of Organism	Standard (Ampicillin)	Zone of inhibition (mm)							
			Sample Concentration (mg/ml)							
			Hexane				Methanol			
			25	50	75	100	25	50	75	100
1	<i>E. coli</i>	-	1	1	2	2	5	8	9	12
2	<i>Serratia</i>	-	0	0	1	1	3	4	5	6
3	<i>Bacillus subtilis</i>	-	0	1	1	2	4	5	8	11

Table 3: Antifungal activity of *Gnetum ula* leaves in different solvents

S. No	Name of Organism	Standard (Ampicillin)	Zone of Inhibition (mm)							
			Sample Concentration (mg/ml)							
			Hexane				Methanol			
			25	50	75	100	25	50	75	100
1	<i>Aspergillus niger</i>	-	2	4	9	13	8	9	14	16
2	<i>Penicillium</i>	-	3	4	10	11	10	12	17	22

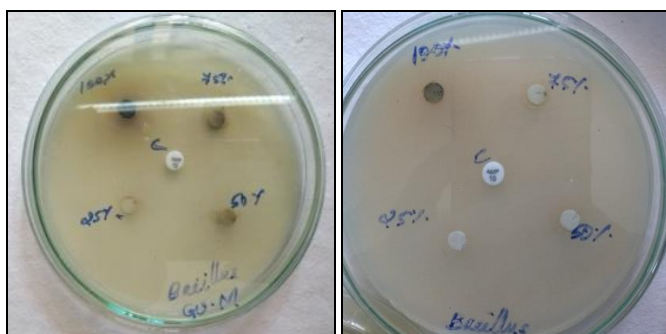
Antimicrobial activity in different solvent of *Gnetum ula* leaves



E. coli streaked plate



Penicillium sp. streaked plate



Bacillus subtilis streaked plate

Conclusion

In the present study the phytochemical screening and antimicrobial activity of *G. ula* leaf extract composed of various phytoconstituents and some of them are responsible for antimicrobial activity and others have several medicinal properties. Thus, this type of analyses are the primary step towards understanding the nature of active principles in this medicinal plant which will be helpful for further research.

Acknowledgement

The authors are thankful to the management, secretary and principle of PSG College of Arts & Science, Coimbatore for the facilities provided, encouragement.

References

- Ciulci I. Methodology for the analysis of vegetable drugs. Chemical Industries branch, Division of Industrial Operations. UNIDO, Romania, 1994, 24-67.
- Kokate CK, Purohit AP, Gokhal ESB. Carbohydrate and derived Products, drugs containing glycosides, drugs containing tannins, lipids and protein alkaloids. Text book of Pharmacognosy, 7, edition, 2001, 133-166, 167-254, 255-269, 272-310, 428-523.
- Gokhale AB, Damre AS, Kulkarni KR, Saraf M. Phytomedicine. 2002; 9(5):433-437.
- Brain KR, TD Turner. The Practical Evaluation of Phytopharmaceuticals. Wright- Scientica, Bristol, 1975, 57-58
- Harbone JB. Phytochemicals methods: A guide to modern technique of plant analysis. Chapman and Hall, London, UK, 1998, 110-113.
- Yogesh Kumar RG, Krishna Swamy K. Phenology and Phytochemical Analysis of *Gnetum ula*, International Journal of Medicinal Plants. Photon. 2017; 107:536-542
- Salkowski CA, Balish E. Inflammatory responses to *Cryptococcus* in congenital athymic mice. Journal of Leukocyte Biology. 1991; 49(6):533-541.
- Sarker SD, Nahar L, Kumarasamy Y. Microtitre plate-based antibacterial assay incorporating resazurin as an indicator of cell growth, and its application in the *in vitro* antibacterial screening of phytochemicals. Methods. 2007; 42:321-324.
- Gourav KD, Patil DT, Thite SV, Patil PR, Kore BA, Aparadh VT. Preliminary investigation of various secondary metabolites from some gymnosperm species.

International Journal of Pharmaceutical and Chemical Sciences. 2013; 2(2):841-843.

10. Ilodibia CV, Ugwu RU, Nwokolo OL, Chukwuma MU, Akachukwu EE. Phytochemical screening, antifungal and antibacterial activity of aqueous and ethanolic leaf and stem extracts of *Gnetum africanum* Welw. Research Journal of Medicinal Plants. 2015; 9(6):275-283.