

ISSN (E): 2320-3862 ISSN (P): 2394-0530 www.plantsjournal.com JMPS 2022; 10(4): 46-60 © 2022 JMPS Received: 10-05-2022 Accepted: 11-06-2022

Mahamane Idi Issa Abdoulahi

Laboratory of Natural Substances and Synthesis Organic, Faculty of Science and Technology, Abdou Moumouni University, BP 10662 Niamey, Niger

Hama Hamadou Habibou

Laboratory of Natural Substances and Synthesis Organic, Faculty of Science and Technology, Abdou Moumouni University, BP 10662 Niamey, Niger

Alio Sanda Abdelkader

Laboratory for Management and Valorization of Biodiversity in Sahel, Faculty of Science and Technology, University Abdou Moumouni, BP 10662 Niamey, Niger.

Nodjitoloum Mborbe

Natural Substances Research Laboratory, Faculty of Exact and Applied Sciences, University of N'Djamena, Tchad.

Moctar Chaibou

Laboratory of Natural Substances and Synthesis Organic, Faculty of Science and Technology, Abdou Moumouni University, BP 10662 Niamev, Niger

Hama Garba Rahila

Laboratory of Natural Substances and Synthesis Organic, Faculty of Science and Technology, Abdou Moumouni University, BP 10662 Niamey, Niger

Bakasso Sahabi

Laboratory of Natural Substances and Synthesis Organic, Faculty of Science and Technology, Abdou Moumouni University, BP 10662 Niamey, Niger

Ilagouma Amadou Tidjani

Laboratory of Natural Substances and Synthesis Organic, Faculty of Science and Technology, Abdou Moumouni University, BP 10662 Niamey, Niger

Corresponding Author: Mahamane Idi Issa Abdoulahi

Laboratory of Natural Substances and Synthesis Organic, Faculty of Science and Technology, Abdou Moumouni University, BP 10662 Niamey, Niger

Ethno Botanical, Pharmacology and Phytochemistry of widely used medicinal plants in Niger: A Review

Mahamane Idi Issa Abdoulahi, Hama Hamadou Habibou, Alio Sanda Abdelkader, Nodjitoloum Mborbe, Moctar Chaibou, Hama Garba Rahila, Bakasso Sahabi and Ilagouma Amadou Tidjani

DOI: https://doi.org/10.22271/plants.2022.v10.i4a.1436

Abstract

The plants Cassia italica (Mill.) F.W. Anders, Limeum pterocarpum (L.), Strychnos innocua Del, Ipomoea asarifolia (DESR.) Roem. & Schult, Blepharis linariifolia Pers, Boscia senegalensis (Pers) Lam. Ex Poir, Neocarya macrophylla (Sabine) Prance, Phyllanthus pentandrus Schum & Tonn and Ximenia americanum L belong to different families and are used for the treatment of several pathologies. The aim of this review is to present the current state of research on the ethnopharmacological use of these medicinal plants and to give an overview of the significant results documented. Scientific information about these plants reported were used. Literature review was conducted using search engines and other scientific database websites for chemicals such as ChemSpider, PubChem. Phytochemical work on some of these plants has revealed the presence of several chemical groups and more than 100 compounds have been isolated from some of these plants. The isolated compounds belong to various chemicals classes such as stilbenes, flavonoids, terpenoids, Tannins. Some crude extracts showed a wide spectrum of biological activities, such as antiproliferative, analgesic, anti-inflammatory, antioxidant, antimicrobial and antiplasmodial activities. This review provides an overview of the use, pharmacology and phytochemistry of these plants.

Keywords: Medicinals plants, ethnopharmacology, pharmacology, phytochemistry, Niger

Introduction

In Africa, according to the World Health Organization (WHO, 2008) [1], more than 80% of the population still use medicinal plants for their health care needs [1]. Plants are generally presented as having efficacy; they are low cost and available. Sustainability of the use of medicinal plants is important concern. The demand for medicinal plants is increasing in Africa as the population grows and pressure on medicinal plant resources will become greater than ever. Interest in plant derived medicines has also increased in the developed countries between the pharmaceutical companies. In contrast, due to their minor side effects, the medicinal plants are widely used to treat many human diseases. The increasing cost of health care and the failure of allopathic medicine to treat some diseases have also participated to the increasing consumption of traditional medicine to cure some disease. Medicinal plants with a long history of safe and efficient use are likely to have a pharmaceutical outcome. In Niger, many research has been done on the traditional pharmacopoeia, ethnobotanical and pharmacological (Khalid et al., 1984; Baoua et al., 1974; Manzo et al., 2017; Jazy et al., 2017) [2-5]. This review describes the traditional uses, phytochemistry and pharmacology of nine (9) medicinal plants from Niger. These plants are distributed in 9 families. They are: Cassia italica (Mill.) F. W. Anders (Caesalpiniaceae), Limeum pterocarpum (L.) (Molluginaceae), Strychnos innocua Del (Loganiaceae), Ipomoea asarifolia (DESR.) Roem. & Schult (Convolvulaceae), Blepharis linariifolia Pers (Acanthaceae), Boscia senegalensis (Pers) Lam. Ex Poir (Capparidaceae), Neocarya macrophylla (Sabine) Prance (Chrysobalanaceae), Phyllanthus pentandrus Schum & Tonn (Euphorbiaceae) and Ximenia americanum L (Olacaceae). These plants are used in the treatment of many diseases.

Methodology

An extensive literature review was conducted using Sci-finder, Pubmed, Scopus, Web of Science, Science Direct, J-Gate, Google Scholar and other scientific database websites for

chemicals such as Chem Spider, PubChem. Keywords used included botany, conservation, Ethno botany, phytochemistry, toxicology and pharmacology. The scientific name of each plant was then combined separately with the terms antimicrobial, antiparasitic, anti-inflammatory, hepatoprotective, antiulcer, immunomodulatory, antioxidant, and antidiarrheal. Scientific names were validated using the lexicon of vernacular plant names from Niger "Lexique des Plantes du Niger". The literature sources used in this article is ranging from French to English.

Results and discussion

The data obtained on the state of research on these plants concerns traditional uses, phytochemistry and biological potential. The compounds isolated from these plants belong to the families of steroids, flavonoids, saponins, quinone, Cyanogenic Glucoside. Only the isolated glycocapparin from *B. senegalensis* has been tested for it hypoglycemic *in vivo*.

Limeum pterocarpum (L.)

There is hardly any work that has been done to show the biological activities of *Limeum pterocarpum* minus the phytochemical composition of this plant. Ethnobotanical surveys have shown that the plant is used in the treatment of malaria, bacterial and fungal infections (Diallo *et al.*, 2001) ^[6]. The leaves are dried, crushed and used with milk to treat malaria. Another antimalarial preparation consists to macerate the whole plant until the water changes colour. The solution is drunk for three days (Diallo *et al.*, 2001) ^[6].

Previous work on *Limeum pterocarpum* has allowed the isolation of the simeolide, a drimane-type sesquiterpene (Ikhiri *et al.*, 1995) ^[7]. Thus, D-pinitol was isolated and characterised from successive treatments of the aqueous extract of the plant (Abdoulaye *et al.*, 2004) ^[8]. A study conducted by Diallo *et al.*, in 2001 ^[6] on the antifungal, antioxidant, molluscicidal and larvicidal properties of 20 medicinal plants, including *L. pterocarpum*, showed that the dichloromethane extract of the aerial parts has antifungal activity, especially on the *Candida albicans* germ (Diallo *et al.*, 2001) ^[6].

Table 1: Previous pharmacological works on L. Pterocarpus

Type of extract	Activities	Biological/chemical model	Results	References	
		Cladosporium cucumerinum	Active		
Methanolic extract	Antifungal	Candida Albicans	No active	(Diallo <i>et al.</i> , 2001) [6]	
Dichloromethane extract	Antifungal	Cladosporium cucumerinum	Not active	(Diallo <i>et al.</i> , 2001) [6]	
		Candida albicans	Active		
Methanolic and dichloromethane extract	Larvicide	Culex Anopheles Aedes	Non actif Actif	(Diallo <i>et al.</i> , 2001) [6]	
Methanone and diemoromethane extract	Larviciue	Cutex Anophetes Aedes	Non actif	(Diano et al., 2001) [6]	
Methanolic and dichloromethane extract	Antioxydant	DPPH TLC	Negative	(Diallo <i>et al.</i> , 2001) [6]	
ivietnanone and dienforomethane extract		β-carotene TLC	Positive	(Diano et al., 2001) [6]	

Blepharis linariifolia Pers (Acanthaceae)

Khalid *et al.*, 1984 ^[2] reported that the *Blepharis linariifolia* plant is used in the treatment of measles and oedema, it is also used for stomach aches, it is cited as an aphrodisiac plant and is included in recipes used in child care. The leafy stems are used to treat malaria and abscesses. The juice of the leaves is antibiotic, anti-protozoal and antiseptic. The leafy stems are used in the treatment of feverish aches and panic attacks. The juice of the leaves is purgative, anti-inflammatory, stimulating the intestine and uterine muscles (Nacoulma, 1996) ^[8]. Women use *B. linariifolia* against stomach ache (Wezel, 2002) ^[9]. According to the African pharmacopoeia, in Niger the leafy plant is used against panicitis, the whole plant against syphilis and the leaves are purgative; in Burkina Faso the stem of the plant is used in the treatment of neck ganglions (Eklu-Natey et Balet, 2012) ^[10].

Little work has been done on the phytochemical composition of *Blepharis linariifolia* Pers. The aqueous extract of the

whole plant contains flavonoids, saponosides and tannins (Baoua *et al.*, 1974) ^[3]. The plant contains tannins, saponosides, sterols and triterpenes; quinones and cyanogenic glycosides are absent; alkaloids and flavonoids are present only in the fruits (Khalid *et al.*, 1992) ^[12]. Nacoulma, in 1996 ^[8] showed that this plant contains saponosides, flavonoids, steroids, anthocyanins in the flowers, triterpenes, Mucilages in the seeds and coumarins.

The aqueous extract was tested on *S. typhimirium* of the isolated strains (Mahamane *et al.*, 2020) ^[13]. Nasereldeen *et al.*, in 2018 ^[14] showed the hepatoprotective properties of the hydroethanolic extract obtained from the whole plant. The antioxidant activity was shown by several methods: FRAP (Mahamane *et al.*, 2020) ^[15]; by DPPH (Wamtinga, 2006; Dirar *et al.*, 2019) ^[16, 17]. The hexane extract of the plant seeds did not show activity against *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Aspergillus niger* (Osama *et al.*, 2017) ^[18].

Table 2: Previous pharmacological works on Blepharis Linariifolia

Type of extract	Activities	Biological / Chemical Model	Results	References
Hexane extract Aqueous extract	Antibacterial	Bacillus subtilis, Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa and Aspergillus niger S. Typhimurium (clinical) S. Typhimurium (lettuce)	No active 7,5±0,70 mm 13,5±2,12 mm	(Osama <i>et al.</i> , 2017) ^[18] (Mahamane <i>et al.</i> , 2020) ^[13]
Hydroethanolic extract Hydroethanolic, aqueous, acetone and dichloromtane extracts Aqueous extract	Antioxydant Antioxydante Antioxydant	DPPH DPPH FRAP	$\%\ I = 78{\pm}0.01 \\ 20.28 \pm 0.64\ to \\ 0.85 \pm 0.12\ \mu g/mL \\ 1{,}78{\pm}0{,}04\ mmol \\ EAA/g$	(Nasereldeen <i>et al.</i> , 2018) (Dirar <i>et al.</i> , 2019) [17] (Mahamane <i>et al.</i> , 2020) [15]
Hydroethanolic extract	Hepato protective	In vivo	400mg/Kg	(Nasereldeen et al., 2018)
Hydroethanolic, aqueous, acetone and dichloromethane extracts	Enzymatic inhibition	Pancreatic lipase	0,63 to 23,47 μg/mL	(Dirar et al., 2019) [17]

Phyllanthus pentandrus Schum & Tonn (Euphorbiaceae)

The plant *P. pentandrus* is widely used in several treatments in Niger for child weaning, scorpion stings, stomach aches and indigestion (Jazy *et al.*, 2017) ^[5]. It is used in the composition of several recipes used in the treatment of infectious diseases. This plant is widely used for infants.

Phytochemical studies carried out on *P. pentandrus* reveal the presence of alkaloids, saponosides and quinones only in the seeds, flavonoids are present only in the leaves and tannins are present in all organs; the absence of cyanogenic glycosides is also noted (Baoua *et al.*, 1974; Alio *et al.*, 2020;

Mahamane *et al.*, 2020) [2, 19, 13].

However, little work has been done on the biological activities of this plant, methanolic extracts have been tested on *E. coli*, *S. aureus* and *P. aeruginosa* (Kwaji *et al.*, 2015) ^[20], aqueous, dichloromethane and ethanolic extracts have been tested on *S. typhimirium* of the isolated strains (Alio *et al.*, 2020; Mahamane *et al.*, 2020) ^[19,13]. Chika and Bello in 2019 ^[21] showed the hepatoprotective properties of aqueous extracts obtained from the leaves. The antioxidant activity by FRAP method was demonstrated by (Mahamane *et al.*, 2020) ^[15].

Table 3: Previous pharmacological works on Phyllanthus Pentandrus

Type of extract	Activities	Biological / chemical model	Results	References
		S. Typhimurium (clinical)	11,5±0,70 mm	
Aqueous extract	Antibacterial	S. Typhimurium (lettuce)	17,5±0,70 mm	(Mahamane <i>et al.</i> , 2020) [13]
Methanolic extract	Antibacterial	E. Coli	17 mm	(Alio et al., 2020) [19]
Wiemanone extract	Antibacteriai	S. Aureus	24 mm	(Kwaji <i>et al.</i> , 2015) ^[20]
		P. Aeruginosa		
Aqueous extract	Antioxydant	FRAP	2,21±0,02 mmol EAA/g	(Mahamane <i>et al.</i> , 2020) [15]

Boscia senegalensis (Pers) Lam. Ex Poir (Capparidaceae)

Khalid et al., in 1992 [12] reported that B. senegalensis is one of the plants with aphrodisiac use, plants treating oedema and rheumatism. The bark is used as a fumigation to treat haemorrhage after childbirth. The infusion of the leaves is used against stomach aches, sinusitis and rheumatism. A decoction of the roots is recommended for the treatment of diarrhoea, oedema, eczema and jaundice (Ikhiri et al., 2000) [22]. In Senegal, the roots, barks and leaves are indicated for the treatment of stomach aches. The powder of the dried leaves added to salt is used as an anti-bilharzian and as a tranquilliser for certain forms of mental illness. The roots are presumed to be vermifuge (Pousset, 2004) [23]. The leaves, twigs, bark, roots, root bark and fruit are indicated for tooth decay, inflammation, sprains, sickle cell disease, persistent headaches, rheumatism and haemorrhoids (Jazy et al., 2017) [5]. The plant is traditionally used to protect cereals against pathogens (Seck et al., 1993; Dicko et al., 2001) [24, 25]. Some works have been done on the different parts and

Some works have been done on the different parts and biological activities of *B. senegalensis*. Diallo *et al.*, 2001 ^[6] showed the antifungal, larvicidal, molluscicidal and antioxidant activities of *Boscia senegalensis* extracts. In addition, the insecticidal power of the plant extracts on the groundnut bruchid (Caryedon serratus (O.L) was carried out by Gueye *et al.*, in 2011 ^[26]. Sakine *et al.*, in 2012 ^[27] evaluated the hypoglycemic activity of glycocapparin isolated from the seeds of this plant by assessing the level of hepatic

glycose release in vivo. Cook et al., in 1998 [28], evaluated the antioxidant activities of the aqueous extract of B. senegalensis leaves with trolox as a control. The extract showed an antioxidant capacity of 22 µmol/g trolox equivalent per gram of dry matter. Antioxidant activity of methanolic extract of B. Senegalensis fruits using 2,2-diphenyl-picrylhydrazyl radical (DPPH) and β -carotene assay was carried out by Diallo *et al.*, in 2001^[6]. The antioxidant activity of the plant fruits was also shown by Ka et al., in 2020 [29] by FRAP and DPPH methods. Alivu et al., in 2008 [30], evaluated the antibacterial activities of ethanolic extract of B. Senegalensis roots against Staphylococcus aureus with an inhibition diameter of 19 mm, a minimum inhibitory concentration (MIC) of 5mg/ml and a minimum bactericidal concentration (MBC) of 6mg/ml. The anthelmintic activity of ethanolic extracts obtained from the leaves and roots of the plant was demonstrated by Chaibou et al., in 2020 [31] these extracts showed a 100% mortality rate after 10 hours at the concentration of 10g/L. Morgan et al, in 2014 [32] isolated a flavonol glucoside called bosenegaloside and identified 7 other compounds already known: rhamnocitrin-3-O-β-D-(6"-O-E-p-coumaroyl)-

glucopyranoside (I), rhamnocitrin-3-O-β-D-glucopyranoside (II), 3,4,5-trimethoxyphenol-β-D-glucopyrinoside (III), Lasianthionoside A (IV), 3,7-dimethyl-1-octene-3,6,7-triol-6-O-β-D-glucopyranoside (V), Syringin (VI) and Austroside B (VII).

Table 4: Previous pharmacological works on Boscia senegalensis

Type of extract	Activities	Biological/chemical model	Results	References	
Dichloromethane extract	Antifungal	Candida Albicans	Active	(Diallo et al., 2001) [6]	
		Cladosporium cucumerinum			
Isolated glycocapparin	Hypoglycemic	In vitro	30 mg/mL	(Sakine <i>et al.</i> , 2012) [27]	
Aqueous extract	Antioxidant	Trolox	22μmol/ g	(Cook et al., 1998) [28]	
Methanolic extract	Antioxidant	DPPH and β-carotene by TLC	Positive	(Diallo et al., 2001) [6]	
	Antibacterial	Staphylococcus aureus	CMI=5mg/mL CMB=6mg/mL	(Aliyu et al., 2008) [30]	
Ethanolic extract	Anthelmintic	In vitro	Mortality rate = 100% at 10g/L	(Chaibou et al., 2020) [31]	

Cassia italica (Mill.) F. W. Anders (Caesalpiniaceae)

The roots and flowers of the plant are used in case of stomach bloating and against colic. The decoction of the flowers is used for splenomegaly. The leaves are used by maceration to treat stomach ache (Pousset, 1989) [33]. The decoction of the whole plant or the leaves are used as a laxative, purgative, for

rheumatism and urinary infections (Al-Said, 1993) [34]. In Saudi Arabia the plant is used for constipation, oedema and skin infections (Al-Yahya *et al.*, 1990) [35].

The decoction of the roots is used against jaundice, snake bites and leprosy. An infusion of the leaves is used as a laxative and purgative (Ikhiri *et al.*, 2000) [22]. In Senegal, the

leaves are indicated to treat constipation, hepatobiliary diseases, venereal diseases and those caused by helminths (Pousset, 1989) [33]. Jazy *et al.*, 2017 [5] reported that the leaves and the whole plant are used against chronic constipation, stomach ache, chronic haemorrhoids, typhoid fevers, sickle cell disease, sickle cell crises and facilitate digestion.

Some work has been done on antiproliferative activities by MTT and antioxidant activities by DPPH and ABTS (Bayala *et al.*, 2020) ^[36]. Rwaida and Mai in 2020 ^[37] isolated a benzoic acid derivative as well as the compounds 1-p-hydroxy benzoyl-3-palmitoyl glycerol (1) and 6-p-hydroxy benzoyl daucosterol (2) scutellarein-6-methyl ether (3), quercetin (4), and rutin (5) which were identified and separated. The aqueous extract of the leaves is hepatotoxic by increasing the total bilirubin level as well as liver enzymes at the dose of

1500mg/Kg of body weight (Shehu et al., 2018) [38]. Gamal in 2014 [39] isolated a new cycloartane triterpene compound, (22E)-3-β-hydroxycycloart-22-en-24one, and 8 already known compounds: β-sitosterol (I), UVAOL (II), daucosterol (III), Me 3,4 dihydroxybenzoate (IV), emodin (V), 4hydroxypheny-O-β-D-glucopyranoside (VI), aloin (VII), and rutin (VIII). GC-MS analysis of the methanolic extract of the leaves of the plant shows the presence of 17 compounds, most of which are used in industry as flavouring, antioxidant, antiinflammatory, antimicrobial, pesticide and antiproliferative (Sermakkani and Thangapandian 2012) [40]. Kazmi et al., in 1994 [41] identified: β-sitosterol, stigmasterol, α-amyrin, 1, 5dihydroxy-3-Me anthraquinone and a new anthraquinone (I) which is believed to be antibacterial and has anticancer activity. The plant also has purgative properties by stimulating intestinal motricity (Assane et al., 1994)^[42].

Table 5: Previous pharmacological works on Cassia Italica

Type of extract	Activities	Biological/chemical model	Results	References
Hydromethanolic extract	Anti-DPPH,	DPPH, ABTS	$IC_{50} = 0.53 \pm 0.04 \mu g ext. / \mu g DPPH$	(Bayala et al., 2020)
Trydromethanone extract	anti-ABTS;	DITH, ADIS	$C = 0.302 \pm 0.003 \mu MET / g \text{ ext.}$	[36]
Methanolic extract	Anti-DPPH	DPPH	800 μg/mL	(Gamal, 2014) ^[39]
Hydromethanolic extract Anti-		MTT Assay		(Bayala <i>et al.</i> , 2020)
Trydromethanone extract	proliferative	WIII Assay		[36]
Aqueux, hexane and		Staphylococcus aureus, Salmonella typhi,		(Dabai <i>et al.</i> , 2012)
methanolic extracts	Antibacterial	Escherichia coli, Pseudomonas	33.0 ± 1.64 et 31.25 ± 0.25 mm at	[43]
methanone extracts		aeruginosa et Streptococcus pneumonies	the concentration of 120 mg/mL	
Aqueux extract	Hepatotoxicity	Albino rats <i>In vivo</i>	1500 mg/Kg	(Shehu et al., 2018)
riqueux extract	Trepatotoxicity	Thomo rats in vivo	1300 mg/Kg	[38]
Ethyl acetate fraction	Anti-DPPH	DPPH	%I = 95.8	(Rwaida et Mai
Ethyl acetate fraction	Allu-Di i ii	DITII	701 – 93.8	2020) [37]

Neocarya Macrophylla (Sabine) Prance (Chrysobalanaceae)

Manzo *et al.*, 2017 ^[4] reported that the bark from the trunk of the plant is used in the treatment of diarrhoeal diseases. *Neocarya macrophylla* (Sabine) Prance is a plant used in the traditional pharmacopoeia of Niger. The bark of the plant is used against intestinal parasitosis. Khalid *et al.*, in 1984 ^[2] showed that this plant is used in the treatment of stomach ailments and leprosy.

The plant has been the subject of some studies against inflammatory pain for its richness in saponoside. The plant has analgesic and anti-inflammatory activities; the extract from the trunk bark is more active than the roots (Jega *et al.*, 2019) [44]. The oil extracted from the fines of this plant has important properties for application in cosmetology, GC-MS analysis revealed the presence of: myristic acid, palmitic acid, stearic acid, palmitoleic acid, elaidic acid, oleic acid, erucic acid, Behenic acid, Heneicosanoic acid, icosatetraenoate and Eicosatrienoic acid (Warra *et al.*, 2019) [45]. Most of the studies done on this plant focus on the nutritional aspect of the seeds, gingerbread etc... (Amza *et al.*, 2015) [46]. The plant

has not been subject to phytochemical and biological studies.

Strychnos innocua Del (Loganiaceae)

The plant's leaves and roots alone or in combination are used to treat typhoid fever and jaundice (Jazy *et al.*, 2017) ^[5].

Previous studies on Strychnos innocua Del focus mainly on the nutritional side and it has been shown that it is rich in sodium and carbohydrates (Bello et al., 2007) [47]. The ethanolic extract of the plant has inhibitory activities of the following enzymes: hyaluronidase, phospholipase A2 and protease. Anthenobilic acid has been identified as the major phenol (Molander et al., 2014) [48]. The root bark contains alkaloids, iridoids, carbohydrates, and phenolic acids; the following compounds have been identified loganic acid, glucose, fructose, Quinic acid, and phenols (Angenot and Wauters 1974) [49]. The plant is mentioned as a plant used in the treatment of infectious diseases such as diarrhoea and typhoid fever, but no work has been done to evaluate its activity against the bacteria responsible for these diseases. The plant has not been the subject of in-depth phytochemical and biological studies.

Table 6: Previous pharmacological works on Strychnos Innocua

Type of extract	Activities	Biological/chemical model	Results	References
Ethanolic extract	Enzymatic inhibition	Hyaluronidase, phospholipase A2 and protease	IC ₅₀ =100 μg/mL	(Molander <i>et al.</i> , 2014) [48]

Ipomoea Asarifolia (DESR) Roem & Schult (Convolvulaceae)

The plant belongs to the Convolvulaceae family and is used in the traditional treatment of several infectious diseases such as typhoid fever, diarrhoea and fungal infections. The leaves, stem and/or whole plant are indicated for sickle cell disease, rheumatism, joint pain, skin allergy, hair loss, haemorrhoids and bacterial infections (Jazy *et al.*, 2017) ^[5].

Previous work has focused on phytochemical, biological and toxicological aspects. The plant does not show activity against *E. coli* ATCC 25992 according to the study conducted by Santos *et al.*, in 2018 ^[50]. The hydro ethanolic extract of the plant exhibits cardioprotective activity (Akindele *et al.*, 2018) ^[51]. The antioxidant activity of the plant was studied by DPPH methods by Muhammad *et al.*, 2018 ^[52] Meda *et al.*, 2017 ^[53]; ABTS and FRAP by Meda *et al.*, 2017 ^[53]. The aqueous

extract of the plant possesses anti-inflammatory activities using the murine model of oedema, peritonitis and air pouch inflammation HPLC-DAD and LC-DAD-MS analyses identified the following compounds rutin, chlorogenic acid and caffeic acid which are believed to be responsible for this activity (Furtado *et al.*, 2016) ^[54]. Other authors have also shown that the aqueous extract has antioxidant activities, against inflammation of the intestine. Two triacyclic and tetraglucosylated anthocyanins derived from cyanidin were isolated from the flowers of the plant and their structures elucidated by GC, MS and NMR methods (1H and 13C, TOCSY-1D, DQF-COSY, DIFFNOE and HMBC) (Pale *et*

al., 2003) ^[55]. Indolic diterpenic alkaloids were isolated by HPLC-HRMS, 1D and 2D NMR spectroscopy methods and compounds associated with tremorrhagic syndrome in cattle (Stephen *et al.*, 2017; Dale *et al.*, 2018) ^[56, 57]. The aqueous extract of the plant leaves exhibits hepato-protective activity against CCl₄-induced damage in rats (Farida *et al.*, 2012) ^[58]. Ethyl acetate and methanol extracts showed acetylcholinesterase inhibitory activity (Feitosa *et al.*, 2011) ^[59]. Toxicity of *Ipomoea asarifolia* has not been reported in humans but in animals in North Brazil, it has been investigated and was linked to the presence of lectins in the plant (Steiner *et al.*, 2011) ^[60].

Table 7: Previous pharmacological works on Ipomoea Asarifolia

Type of extract	Activities	Biological/chemical model	Results	References
Ethanolic extract	Antibacterial	E. Coli ATCC 25992	Not active	(Santos et al., 2018) [50]
Ethyl acetate and methanol extracts	Enzymatic inhibition	Acétylcholinestérase	$IC_{50} = 0.12 mg/mL$	(Feitosa <i>et al.</i> , 2011) ^[59]
Methanolic extract	Hepatoprotective	In Vivo	100, 200 et 400mg/kg	(Farida <i>et al.</i> , 2012) ^[58]
Aqueous and methanolic extract Aqueous extract	Antioxidant Antioxidant Antioxidant	DPPH FRAP ABTS	IC ₅₀ =92 µg/mL IC ₅₀ =92 µg/mL 363.963 µmol TE/g 169.828 to 2559.512 µmol AAE/g	(Muhammad <i>et al.</i> , 2018) ^[52] (Meda <i>et al.</i> , 2017) ^[53] (Meda <i>et al.</i> , 2017) ^[53]

Ximenia americanum L (Olacaceae)

Manzo *et al.*, 2017 ^[4] reported that the bark of the trunk of the plant is used in treatments of diarrhoeal diseases. The leaves, twigs, barks, roots and fruits of the plant are used in dracunculiasis, aches and pains, HTA, gonococcal disease, wound, haemorrhoids, abdominal pain, indigestion, jaundice (Jazy *et al.*, 2017) ^[5]. The ethnobotanical survey conducted by Khalid *et al.*, in 1984 ^[2] showed that this plant is used in the treatment of jaundice, stomach ache and is indicated as a haemostatic plant.

This plant is one of the most used in the treatment of infectious diseases in West Africa. It has been the subject of several studies to determine its antimicrobial activity. Several types of extracts obtained from the different organs of the plant have been tested on a wide range of pathogens. Ethanolic and o-tolidine extracts were tested on *Escherichia coli*, *Staphylococcus aureus*, *Salmonella spp* and *Candida albicans* (Gunasekhar *et al.*, 2017) ^[61]. Antimicrobial and antidiarrheal activity of the saponoside-rich root extract was evaluated in Wistar rats (Kiessoun *et al.*, 2018) ^[62]. The essential oil obtained from the leaves of the plant showed antibacterial activity against *Staphylococus aureus* (Mulugeta *et al.*, 2015) ^[63]. The antioxidant activity of the plant was evaluated by several DPPH, FRAP and ABTS methods. Thus, the ethanolic extract of the leaves showed a percentage

inhibition of $62.32\pm0.17\%$ and a reducing power of $3.45\pm$ 0.97 mmol EAA/10 g (Dramane et al., 2019) [64]. The aqueous extract of the plant obtained by infusion has analgesic activity at the concentration of 100 mg/mL but this extract is toxic at 400 mg/mL (Soro et al., 2009) [65]. Aqueous and methanolic extracts obtained from leaves, barks and roots were tested against the following strains Bacillus subtilis, Staphyllococcus aureus, Escherichia coli, Psedomonas aeruginosa of which the methanolic extract showed the best activity with 0.31 mg/mL against Bacillus subtilis (Omer and Elmina 2003) [66]. The methanolic extract of the plant leaves was active against Candidas albicans and Trichophyton rubrum with IC50 = 8.12 μ g/mL and IC50 = 26.06 μ g/mL respectively. The hexanolic, methanolic extracts were not active on Esherichia coli and Mycobacterium Chelonae, but showed activity on Staphylococcus aureus with IC50 < 64 µg/mL (Traoré et al., 2015) [67]. The phenol-rich extract obtained after liquid-liquid separation of the hydro acetone extract with dichloromethane is active on the following strains Shigella dysenteriae, Shigella boydii, Shigella Flexneri, Salmonella thyphi, Klebsiella pneumonia, Proteus mirabilis with MICs ranging from 25 to 50 µg/mL and BMCs ranging from 50 to 100 µg/mL. However, this extract is not active on Klebsiella Arogenes and Escherichia coli, (Kiessoun et al., 2018) [62].

Table 8: Previous pharmacological works on Ximenia Americanum

Type of extract	Activities	Biological/chemical model	Results	References
Ethanolic extract	Anti-DPPH, Reducing power	DPPH, FRAP	%I=62.32 ± 0.17 3.45 ± 0.97 mmol EAA/10g	(Dramane <i>et al.</i> , 2019) [64]
Methanolic extrait	Antifungal	Candidas albicans Trichophyton rubrum	$IC_{50} = 8,12 \mu g/mL IC_{50}$ = 26,06 \text{ \mu g/mL}	(Traoré <i>et al.</i> , 2015) [67]
Hydro acetone extracts Hexanolic, methanolic extract	Antibacterial	Shigella dysenteriae, Shigella boydii, Shigella flexneri, Salmonella thyphi, Klebsiella pneumonia, Proteus Mirabilis Staphylococcus Aureus	CMI 25 à 50 μ g/mL CMB 50 à 100 μ g/mL IC ₅₀ < 64 μ g/mL	(Kiessoun <i>et al.</i> , 2018) ^[62] (Traoré <i>et al.</i> , 2015) ^[67]
Aqueous extract	Analgesic	Albino rats In vivo	At the concentration C= 100 mg/mL	(Soro et al., 2009) [65]

Journal of Medicinal Plants Studies http://www.plantsjournal.com

 Table 9: Traditionnals uses of the nine plants

Plants	Part(s) used	Traditional uses	es Forms of use	
L.	Leaves,	Malaria, Bacterial and Fungal Infections	Maceration / Decoction	(Diallo et al., 2001) [6]
Pterocarpum	Whole plant	, ,	for drinking	(,
B. Linariifolia	Whole plant Leafy stems Leaf juice Leafy plant	Measles Edema Aphrodisiac Children's care Stomach aches Malaria Abscesses Antibiotic, anti-protozoal, antiseptic, anti- inflammatory, stimulant of the intestine and uterine muscles Panariasis, syphilis, purgative,	Maceration/decoction for drinking By enema	(Khalid <i>et al.</i> , 1984) ^[2] (Wezel, 2002) ^[10] (Nacoulma, 1996) ^[9] (Nacoulma, 1996) ^[9] (Eklu-Natey et Balet, 2012) ^[11]
P. Pentandrus	Whole plant Aerial part	Weaning children, Scorpion sting, Stomach ache, Indigestion, Treatment of infectious diseases	Maceration/decoction for drinking	(Jazy <i>et al.</i> , 2017) ^[5]
B. Senegalensis	Roots Roots Leaves Barks	Aphrodisiac use, treatment of oedema and rheumatism Treatment of bleeding after childbirth Stomach aches Treatment of diarrhoea, oedema, eczema and jaundice Tooth decay, inflammation, sprains, sickle cell disease, persistent headaches, rheumatism and haemorrhoids Infections	Maceration/decoction/ Infusion for drinking and enema Fumigation	(Khalid <i>et al.</i> , en 1992) ^[12] (Ikhiri <i>et al.</i> , 2000) ^[22] (Chaibou <i>et al.</i> , 2020) ^[31] (Chaibou <i>et al.</i> , 2020) ^[31] (Jazy <i>et al.</i> , 2017) ^[5]
C. Italica	Roots Flowers Whole plant Whole plant Roots Leaves Leaves and whole plant	Belly bloating, colic Splenomegaly Laxative, purgative, rheumatism and urinary infections Constipation, oedema and skin infections Icterus, snake bites and leprosy Hepatobiliary disorders, venereal diseases, those caused by helminths Bellyache, chronic haemorrhoids, typhoid fevers, sickle cell disease,	Macération/décoction/ Infusion à boire et lavement	(Pousset, 1989) [33] (Al-Said 1993) [34] (Al-Yahya et al., 1990) [35] (Ikhiri et al., 2000) [22] (Pousset, 1989) [33] (Jazy et al., 2017) [5]
N. Macrophylla	Roots, bark Barks Barks	Treatment of diarrhoeal diseases, Intestinal parasitosis, Treatment of stomach ache and leprosy	Oral decoction or maceration	(Khalid <i>et al.</i> , 1984) ^[2] (Jazy <i>et al.</i> , 2017) ^[5] (Manzo <i>et al.</i> , 2017) ^[4]
S. Innocua	Barks and/or leaves	Typhoid fever and jaundice	Oral decoction or maceration	(Jazy et al., 2017) ^[5]
I. Asarifolia	Leaves, stem, whole plant	Typhoid fever, diarrhoea and fungal infections Sickle cell disease, rheumatism, joint pain, skin allergy, hair loss, hemorrhoids and bacterial infections	Oral decoction, infusion or maceration and enema	(Jazy et al., 2017) ^[5]
X. Americana	Trunk bark Leaves, twig, bark, root, fruit	Treatment of diarrhoeal diseases, bacterial infections Dracunculosis, aches and pains, HTA, gonorrhoea, wounds, haemorrhoids, abdominal pain, indigestion, jaundice Icterus, stomach ache and is indicated as a haemostatic plant	Oral decoction, infusion or maceration and enema	(Manzo <i>et al.</i> , 2017) ^[4] (Khalid <i>et al.</i> , 1984) ^[2] (Jazy <i>et al.</i> , 2017) ^[5]

Table 10: Isolated compound

			Country	
Name	Plant Species	Part	Country of Origin	Références
Cycloartane triterpene (1), (22E)-3-β-hydroxycycloart-22-en-24one, β-sitosterol (2), uvaol (3), daucosterol (4), Me 3,4 dihydroxybenzoate (5), emodin (6), 4-hydroxypheny-O-β-D-glucopyrinoside (7), aloin (8), and rutin (9)	C. Italica	Leaves	Saudi Arabia	(Gamal, 2014) [39]
1-p -hydroxy benzoyl-3-palmitoyl glycerol (10), 6 - p -hydroxy ben-zoyl daucosterol (11), scutellarein-6-methyl ether (12), quercetin (13), rutin (14)	C. Italica	Partie aérienne	Saudi Arabia	(Rwaida et Mai 2020) [37]
β-sisterol (15), stigmasterol (16), α-amyrin, 1,5-dihydroxy-3-methyl anthraquinone (17), anthraquinone (18)	C. Italica	Plante entière	Pakistan	(Kazmi <i>et al.</i> , 1994) ^[41]
Bosenegaloside (19), rhamnocitrin-3-O-β-D-(6"-O-E-p-coumaroyl)-glucopyrinoside (20), rhamnocitrin-3-O-β-D-glucopyrinoside (21), 3,4,5-trimethoxyphenol-β-D-glucopyrinoside (22), lasianthionoside A (23), 3,7-dimethyl-1-octene-3,6,7-triol-6-O-β-D-glucopyranoside (24), syringin (25), austroside B (26)	B. Senegalensis	Feuilles	Nigéria	(Morgan et al., 2014) ^[32]
Liméolide (27), D-pinitol (28)	L. Pterocarpum	Plante entière	Niger	(Abdoulaye <i>et al.</i> , 2004) [8]
6,7-Dehydro-11-hydroxy-12,13-epoxyterpendole A (29), terpendole K (30), 6,7-dehydroterpendole A (31), 11-hydroxy-12,13-epoxyterpendole K (32), terpendole C (33), paxilline (34) terpendole K (35), terpendole E (36), 11-hydroxy-12,13-epoxy-terpendole K (37), 6,7-dehydroterpendole A(38)	Ipomea asarifolia Ipomea asarifolia	Graines Graines	Brésil Brézil	(Dale et al., 2018) [57] (Stephen et al., 2017) [56]
cyanogenic	Ximenia	Feuilles	Mali	(Le et al.,

glycoside sambunigrin (39), gallic acid (40), gallo tannins-β-glucogalline (41), 1,6-digalloyl-β-glucopyranose (42), quercetin (43), quercitrin, (quercetin-3-O-α-rhamnopyranoside) (44), avicularin, (quercetin-3-O-α-arabinofuranoside) (45), quercetin-3-O-β-xylopyranoside (46), quercetin-3-O-(6''-galloyl)-β-glucopyranoside (47), kaempferol-3-O-(6''-galloyl)-β-glucopyranoside (48)

Terpendole K

6,7-dehydroterpendole

11-hydroxy-12,13-epoxyterpendole

6,7-dehydro-11-hydroxy-12,13-epoxyterpendole

Terpendole C R1=H, R2=

Terpendole A R1=H, R2=

Terpendole M R1=OH, R2=

$$\begin{array}{c} H \\ \hline \\ N \\ \hline \\ H \\ \end{array}$$

Paxilline R1=OH, R2=O 13-DesoxypaxillineR1=H, R2=O Paxitriol R1=R2=OH

Paspaline

Fig 1: Isolated compounds from some plants

Conclusion

This review provides an overview of the status of 9 medicinal plants widely used in Niger especially in the treatment of infectious diseases based on available published works. Although these plants are widely used, little work has been undertaken to demonstrate the efficacy and safety of these plants and to isolate the pure compounds responsible for their biological activities. Thus to date in Niger there is not yet a phytomedicine based on one of these plants. It should also be noted that the study of the mechanism of action of plants extracts on microorganisms is still at a very preliminary stage. This should be seen as a challenge.

References

- 1. World Health Organization. World Health statistics, (Sl.) World Health Organization, 2008.
- 2. Khalid I, Garba M, Saadou M. Recherche sur la pharmacopée traditionnelle au Niger, 1984, 31-35.
- Baoua M, Fayn J, Bessire JM, Et Koudogbo B. Contribution à l'étude de la pharmacopée traditionnelle du Niger. Ecole des Sciences de la Santé. Laboratoire de Chimie Biologique Université de Niamey, 1974.
- Manzo LM, Moussa I, Khalid I. Les plantes médicinales utilisées dans le traitement des diarrhées au Niger: étude ethnobotanique, Algerian J. Nat. Products. 2017;5(2)475-482
- Jazy MA, Karim S, Morou B, Rokia S, Et Saadou M. Enquête Ethnobotanique auprès des tradipraticiens de santé des Régions de Niamey et Tillabéri au Niger: données 2012-2017, European Scientific Journal November Edition, 2017, 13(33). ISSN: 1857-7881 (PRINT) e-ISSN 1857-7431.
- 6. Diallo D, Marston A, Terreaux C, Toure Y, Smestad Paulsen B, Hostettmann K. Screening of Malian medicinal plants for antifungal, larvicidal, molluscicidal, antioxidant and radical scavenging activities. Phytotherapy research. 2001;15(5):401-406.
- 7. Ikhiri Khalid, Mahaman Ibrahim, Ahond Alain, Chiaroni

- Angele, Poupat Christiane, Riche Claude, *et al.* Limeolid, a new drimane-type sesquiterpene isolated from *Limeum pterocarpum* Journal of Natural Products. 1995;58(7):1136-8
- 8. Abdoulaye Alassane, Idrissa Moussa, Djibo A. Keita, Khalid Ikhiri (2004). Le D-pinitol isolé de *Limeum pterocarpum* CR Chimie. 2004;7:989-991.
- 9. Nacoulma-Ouédraogo OG. Plantes médicinales et pratiques médicales traditionnelles au Burkina Faso : cas du plateau central. Thèse de doctorat d'État, Faculté des sciences et techniques, Université de Ouagadougou, Burkina Faso, Tome 1, 320, tome. 1996;2:285.
- 10. Wezel A. Plantes médicinales et leur utilisation traditionnelle chez les paysans au Niger. Etudes sur la flore et la végétation du Burkina Faso et des pays avoisinants. 2002;6:9-18.
- Eklu-Natey RD, Et Balet A. Pharmacopée Africaine: dictionnaire et monographies multilingues du potentiel médicinal des plantes Africaines. Afrique de l'ouest volume 2. Editions d'en bas-traditions et médecine T&M Genève, 2012, 999, ISBN: 978-2-8290-0433-9.
- 12. Khalid Ikhiri, Diarra Boureima, Dan Dicko Dan Koulodo. Chemical Screening of Medicinal Plants used in the Traditional Pharmacopoeia of Niger. International Journal of Pharmacognosy. 0925-161 8/92/3004-0251, 1992;30(4):251-262.
- 13. Mahamane Idi Issa Abdoulahi, Bakasso Sahabi, Alio Sanda Abdelkader, Ilagouma Amadou Tidjani, Yaou Chaïbou, Moctar Chaibou, *et al.* Phytochemical Investigation and Antimicrobial Activity of Six Plants Used in Children's Ailments Treatment in Niger. Journal of Diseases and Medicinal Plants. 2020;4:92-97., DOI: 10.11648/j.jdmp.20200604.15.
- 14. Nasereldeen Imam Mohammed, Abdelwahab Siddig Ibrahim, Mohame Manal, Taha Elhassan, Mohan Syam, Alhazmi Hassan Ahmed, Ahmed, et al. Antioxidant and hepatoprotective activities of Blepharis linariifolia Pers and Guiera senegalensis JF. Gmel. Against CCl4-

induced hepatotoxicity World Journal of Pharmaceutical Research. 2018;7(18):70-80.

- 15. Mahamane Idi Issa Abdoulahi, Bakasso Sahabi, Hama Garba Rahila, Hama Hamadou Habibou, Hassane Hamidou Zeinabou, Ilagouma Amadou Tidjani, *et al.* Phenolics contents and antioxidant activity of six medicinal plants used in the treatment of dentition related ailments in Niger. Journal of Pharmacognosy and Phytochemistry. 2020;9(4):1816-1821.
- 16. Wamtinga RS. Etude comparée de la phytochimie et des effets pharmacologiques de quelques plantes de la famille des acanthaceae utilisées dans la prise en charge traditionnelle du paludisme, Thèse unique, Biochimie et chimie des substances naturelles. Université d'Ouagadougou, Burkina Faso, 2006, 207.
- 17. Dirar Amina Ibrahim, Wada Mikiyo, Watanabe Takashi, Devkota, Hari Prasad. Phenolic compounds from the aerial parts of *Blepharis linariifolia* pers. and their free radical scavenging and enzyme inhibitory activities, Medicines. 2019;6(4):113.
- 18. Osama Alsiddig, Sufyan Awadelkarim, Noha Ali, Safa Khalid, Shima Mohammed, Nosiba Hashim. Phytochemical Composition and Evaluation of Antimicrobial Activity of *Blepharis linariifolia* (Pers.) Seeds Asian Journal of Chemical Sciences. 2017;2(2):1-6. Article no.AJOCS.33182.
- 19. Alio Sanda Abdel Kader, Inoussa Maman Maârouhi, Samna Soumana Oumarou, Alfa Keita Djibo, Halima Oumarou Diadié, Mahamane Idi Issa Abdoulahi, *et al.* Evaluation of antibacterial activity from extract of three plant species on strains of *Salmonella* in niger. World Journal of Pharmacy and Pharmaceutical Sciences. 2020;9(3):462-481. ISSN 2278-4357.
- 20. Kwaji A, Adamu HM, Chindo IY, Umar M. Phytochemical analysis and antibacterial activity of *phyllanthus pentandrus* methanol extract. Topclass Journal of Herbal Medicine. 2015;4(1):4-8.
- 21. Chika A, Bello SO. Hepatoprotective and body weight lowering effects of the aqueous leaf extract of *Phyllanthus pentandrus* Schumach and Thonn (Phyllanthaceae) in nonalcoholic fatty liver disease induced by a high-fat diet in Wistar rats. Natl J Physiol Pharm Pharmacol. 2019;9(12):1251-1256.
- 22. Ikhiri Khalid, Ilagouma Amadou et Guero Djimrao. Etudes sur la préparation d'un programme de promotion de la pharmacopée traditionnelle de la zone d'intervention du PROZOPAS, KRB ingénieurs rapport définitif. Université A. Moumoni (Niger), 2000.
- 23. Pousset JL. Plantes médicinales d'Afrique. Comment les reconnaitre et les utiliser ? EDISUD, 2004.
- 24. Seck D, Lognay G, Haubruge E, Wathelet JP, Marlier M, Gaspar C, *et al.* Biological activity of the shrub *Boscia senegalensis* (PERS.) LAM. ex Poir. (Capparaceae) on stored grain insects. J Chem Ecol. 1993;19:377-389.
- 25. Dicko Mamoudou H, Searle-van Leeuwen, Marjo JF. Traore Alfred S, Hilhorst Riet, *et al*, Polysaccharide hydrolases from leaves of *Boscia senegalensis*: properties of endo-(1→3)-β-D-glucanase. Applied Biochemistry and Biotechnology. 2001;94(3):225-241.
- 26. Gueye Momar Talla, Dogo Seck, Seynabou BA, Kerstin Hell, Mbacké Sembene, Wathelet JP, *et al.* Insecticidal activity of *Boscia senegalensis* (Pers.) Lam ex Poir. on Caryedon serratus (OL.) pest of stored groundnuts. African Journal of Agricultural Research. 2011;6(30):6348-6353. 12 December, 2011

- 27. Sakine, Mahamat Nour Adam, Mahmout Yaya, Dijoux-Franca MG, Gbenou Joachim, Moudachirou Mansourou. *In vitro* anti-hyperglycaemic effect of glucocapparin isolated from the seeds of *Boscia senegalensis* (Pers.) Lam. ex Poiret, African Journal of Biotechnology. 2012;11(23):6345-6349. Caplus, DOI: 10.5897/AJB11.3445
- 28. Cook JA, Vanderjagt DJ, Dasgupta A, Mounkaila G, Glew RS, Blackwell W, *et al.* Use of the trolox assay to estimate the antioxidant content of seventeen edible wild plants of Niger. Life Sciences. 1998;63(2):105-110.
- 29. Ka Awa, Badji Kady Diatta, Sagna Moustapha Bassimbe, Guisse Aliou, Bassene Emmanuel. Phytochemical screening and antioxidant activity of the fruits of *Boscia senegalensis* (pers.) Lam. E.G. Pear. (Capparaceae). Pharmacognosy Journal. 2020;12(5):1042-1049.
- Aliyu AB, Musa AM, Abdullahi MS, Oyewale AO, Gwarzo US. Activity of plant extracts used in Northern Nigerian traditional medicine against methicillin-resistant Staphylococcus aureus (MRSA). Nig. J Pharm. Sci. 2008;7(1):1-8
- 31. Chaibou Moctar, Zakari Chaibou Ousmane, Habibou Hama Hamadou, Abdoul Nasser Moussa Bamba, Moussa Idrissa, Arzika Tanimoune, *et al. In vitro* study of the anthelmintic effects of ethanolic extracts of Boscia senegalensis (Pers) Lam. Ex Poir. (Capparaceae) plant used as antiparasitic in Azawagh area in Niger. IOSR Journal of Pharmacy and Biological Sciences (IOSR-JPBS). 2020;15(1):61-65.
- 32. Morgan Abubaker MA, Kim Jang Hoon, Kim Sang Kyum, Lim Chi-Hwan, Kim Young Ho. A new flavonol glycoside from the leaves of *Boscia senegalensis*. Bulletin of the Korean Chemical Society. 2014;35(12):3447-3452.
- 33. Pousset JL. Plantes médicinales africaines. Utilisation pratique. Tome 1 et 2. Ellipses. ACCT, 1989.
- 34. Al-Said MS. Traditional medicinal plants of Saudi Arabia. AMJ Chinese Med. 1993;2:1-12.
- 35. Al-Yahya MA, Al-Meshal IA, Mossa JS, Al-Badr AA, Tariq M. Saudi plants. A phytochemical and biological approach. Riyadh: King Saud University, 1990, 349.
- 36. Bayala Bagora, Zohoncon Theodora Mahoukede, Djigma Florencia Wendkuuni, Nadembega Christelle, Baron Silvere, Lobaccaro Jean-Marc, *et al.* Antioxidant and antiproliferative activities on prostate and cervical cultured cancer cells of five medicinal plant extracts from Burkina Faso. International Journal of Biological and Chemical Sciences. 2020;14(3):652-663.
- 37. Rwaida A Al-Haidari, Mai M Al-Oqail. New benzoic acid derivatives from *Cassia italica* growing in Saudi Arabia and their antioxidant activity. Saudi Pharmaceutical Journal. 2020;28:1112-1117.
- 38. Shehu S, Abubakar AS, Ahmed H. Evaluation of hepatotoxic effects of leaves extract of *Cassia italica* (mill.) lam. Ex F.W. Ander (Leguminosae) in albino rats Journal of Applied Sciences and Environmental Management. 2018;22(9):1539-1542.
- Gamal A Mohamed. New cytotoxic cycloartane triterpene from *Cassia italica* aerial parts, Natural Product Research: Formerly Natural Product Letters. 2014;28(13):976-983.
 DOI: 10.1080/14786419.2014.902820
- 40. Sermakkani M, Thangapandian V. GC-MS analysis of *Cassia italica* leaf methanol extract. Asian Journal of Pharmaceutical and Clinical Research. 2012;5(2):90-94.

Journal of Medicinal Plants Studies http://www.plantsjournal.com

41. Kazmi Mehdi, Malik Abdul, Hameed Saira, Akhtar Nargis, Ali Samina Noor. An anthraquinone derivative from *Cassia italica* Phytochemistry. 1994;36(3):761-3.

- 42. Assane M, Nydyema R, Bassene E, Sere A, Gaye O. Purgative activity of *Cassia italica*. Dakar médical. 1994;39(2):125-8,
- 43. Dabai YU, Kawo AH, Aliyu RM. Phytochemical screening and antibacterial activity of the leaf and root extracts of *Senna italica* African Journal of Pharmacy and Pharmacology. 2012;6(12):914-918. 2012 Mar 29. Available online at

http://www.academicjournals.org/AJPP DOI: 10.5897/AJPP11.852, ISSN: 1996-0816, ©2012 Academic Journals

- 44. Jega Amina Yusuf, Abdullahi Musa Ismail, Rufa'i Amina Idris, Bala Asma'u Baba, Abubakar Hassan, Abdulrahman, Muntaka, *et al.* Effect of saponin-rich fractions of Neocarya macrophylla on murine models of pain and inflammation. Asian Journal of Biological Sciences 2019;12(2):349-355.
- 45. Warra Aliyu Ahmad, Hassan Lawal Gusau, Babatola Leye Jonathan, Omodolapo Adedara Adejoju, Ukpanukpong Richard Undigweundeye, Berena Godwin Ayebaitari. Characterization of *Neocarya macrophylla* seed oil using gas chromatography-mass spectrometry (GC-MS) and Fourier transform infra-red (FT-IR) techniques. Chemical Science International Journal. 2019;28:3, CSIJ.47032/1-CSIJ.47032/7.
- 46. Amza Tidjani, Amadou Issoufou, Balla Abdourahamane, Zhou Hui Ming. Antioxidant capacity of hydrolyzed protein fractions obtained from an under-explored seed protein: Gingerbread plum (*Neocarya macrophylla*) Journal of Food Science and Technology (New Delhi, India). 2015;52(5):2770-2778.
- 47. Bello MO, Olawore NO, Falade OS, Adewusi SRA. Studies on the chemical compositions and antinutrients of some lesser known Nigeria fruits. Bio Chemistry: An Indian Journal. 2007;1(2):88-97.
- 48. Molander Marianne, Nielsen Line, Soegaard Soeren, Staerk Dan, Roensted Nina, Diallo Drissa, *et al.* Hyaluronidase, phospholipase A2 and protease inhibitory activity of plants used in traditional treatment of snakebite-induced tissue necrosis in Mali, DR Congo and South Africa. Journal of Ethnopharmacology. 2014,157:171-180.
- 49. Angenot L, Wauters JN. *Strychnos innocua* (Loganiaceae) Plantes Medicinales et Phytothérapie. 1974;8(4):314-17.
- 50. Santos Dos, Rafaele Aragao, Pinto, Vicente de Paulo Teixeira, Barbosa Francisco Cesar Barroso. Antimicrobial susceptibility and genetic characterization of Extended Spectrum Beta-Lactamase (ESBL)-producing nosocomial strains of *Escherichia coli* isolated from a Brazilian Teaching Hospital, International Journal of Current Microbiology and Applied Sciences. 2018;7(8):422-434.
- 51. Akindele Abidemi J, Palmer Eyitemi L. Effects of hydroethanolic leaf extract of *Ipomoea asarifolia* (Convolvulaceae) in doxorubicin and isoproterenolinduced toxicity in rats. Tropical Journal of Natural Product Research. 2018;2(2):59-66.
- 52. Muhammad Suleiman Alhaji, Abubakar Aminu Lailaba, Rabiu Sulaiman, Sulaiman Ismail. Antioxidant content and *in vitro* 2, 2-diphenyl-1-picrylhydrazyl free radical scavenging activity of selected medicinal plants Journal

- of Intercultural Ethnopharmacology. 2018;7(1):57-65.
- 53. Meda Nag-Tiero Roland, Compaore Moussa, Bangou Mindiediba Jean, Vlase Laurian, Kiendrebeogo Martin, Ouedraogo Anicet Georges. Antioxidant screening and polyphenols profiling of *Ipomoea asarifolia* (DESR.) Roem. & Schult (Convolvulaceae), World Journal of Pharmacy and Pharmaceutical Sciences. 2017;6(11):58-68
- 54. Furtado Allanny A, Manoela Torres-Rêgo, Maíra CJS Lima, Mariana Bitencourt AO, Andréia Bergamo Estrela, Nayara Souza da Silva, et al. Aqueous extract from *Ipomoea asarifolia* (Convolvulaceae) leaves and its phenolic compounds have anti-inflammatory activity in murine models of edema, peritonitis and air-pouch inflammation, Journal of Ethnopharmacology. 2016;192:225-235. ISSN: 0378-8741. https://doi.org/10.1016/j.jep.2016.07.048.
- 55. Pale Eloi, Kouda-Bonafos Marie, Nacro Mouhoussine, Vanhaelen Maurice, Vanhaelen-Fastre Renee. Two triacylated and tetraglucosylated anthocyanins from *Ipomoea asarifolia* flowers. Phytochemistry (Elsevier). 2003;64(8):1395-1399.
- 56. Stephen Lee T, Gardner Dale R, Cook Daniel. Identification of Indole Diterpenes in *Ipomoea asarifolia* and *Ipomoea muelleri*, Plants Tremorgenic to Livestock. Journal of agricultural and food chemistry. 2017;65(26):5266-5277.
- 57. Dale A Gardner R, Welch Kevin D, Lee Stephen T, Cook Daniel, Riet-Correa Franklin, Riet-Correa Franklin. Tremorgenic Indole Diterpenes from *Ipomoea asarifolia* and *Ipomoea muelleri* and the Identification of 6, 7-Dehydro-11-hydroxy-12, 13-epoxyterpendole. Journal of natural products. 2018;81(7):1682-1686.
- 58. Farida T, Salawu OA, Tijani AY, Ejiofor JI. Pharmacological evaluation of *Ipomoea asarifolia* (DESR.) against carbon tetrachloride-induced hepatotoxicity in rats. Journal of ethnopharmacology. 2012;142(3):642-6
- 59. Feitosa CM, Freitas RM, Luz NNN, Bezerra MZB, Trevisan MTS. Acetylcholinesterase inhibition by somes promising Brazilian medicinal plants, Brazilian journal of biology = Revista brasleira de biologia. Database: MEDLINE. 2011;71(3):783-9.
- 60. Steiner Ulrike, Leibner Sarah, Schardl Christopher Lewis, Leuchtmann Adrian, Leistner Eckhard. Periglandula, a new fungal genus within the Clavicipitaceae and its association with Convolvulaceae Mycologia. 2011;103(5):1133-45.
- 61. Gunasekhar T, Mahalakshmi S, Siddaiah M. Evaluation of preliminary phytochemicals and Biological study of ortho-tolidine extract of *Ximenia Americana*, British Journal of Pharmaceutical and Medical Research. 2017;2(2):414-417.
- 62. Kiessoun Konate, Roland Meda Nag Tiero, Mamounata Diao, Yomalan Kassi, Sytar Oksana, Souz Alain, *et al.* Antimicrobial profiles, antidiarrheal and antipyretic capacities of phenol acid rich-fractions from *Ximenia americana* 1., (Olacaceae) in wistar albino rats. International Journal of Pharmacy and Pharmaceutical Sciences. 2018;10(10):62-68.
- 63. Mulugeta Tirhas, Unnithan CR, Tesfaysc Desta, Wemariam Yohannes. Chemical composition and antibacterial activities of the essential oils of *Ximenia Americana* leaves World Journal of Pharmacy and Pharmaceutical Sciences. 2015;4(9):916-923.

- 64. Dramane Pare, Jotham Yhi Pênê N'do, Samson Guenne, Mahamadi Nikiema, Adama Hilou. Phytochemical Study and Biological Activities of Two Medicinal Plants used in Burkina Faso: *Lannea velutina* A. Rich (Anacardiaceae) and *Ximenia Americana* L. (Olacaceae). Asian Journal of Chemical Sciences. 2019; Article no. AJOCS.51333, ISSN: 2456-7795., 2019;6(3):1-9.
- 65. Soro Tianga Yaya, Traore F, Sakande J. Activité analgésique de l'extrait aqueux de *Ximenia americana* (Linné), (Olacaceae) Comptes rendus biologies 2009;332(4):371-7.
- 66. Omer ME, Elnima EI. Antimicrobial activity of *Ximenia Americana*. Fitoterapia. 2003 Feb;74(1-2):122-6. DOI: 10.1016/s0367-326x(02)00302-7, PMID: 12628405.
- 67. Traoré Sahar M, Baldé MA, Camara A, Baldé ES, Diané S, Diallo MS, *et al.* The malaria co-infection challenge: An investigation into the antimicrobial activity of selected Guinean medicinal plants. J Ethnopharmacol. 2015 Nov;174:576-81., DOI: 10.1016/j.jep.2015.03.008., Epub 2015 Mar 13. PMID: 25773488.
- 68. Le Nhat Hao Tran, Malterud Karl Egil, Diallo Drissa, Paulsen Berit Smestad, Nergard Cecilie Sogn, Wangensteen Helle. Bioactive polyphenols in Ximenia Americana and the traditional use among Malian healers Journal of Ethnopharmacology. 2012;139(3):858-862., DOI: 10.1016/j.jep.2011.12.031.