



ISSN (E): 2320-3862
ISSN (P): 2394-0530
<https://www.plantsjournal.com>
JMPS 2023; 11(2): 47-49
© 2023 JMPS
Received: 15-01-2023
Accepted: 21-02-2023

Narbahadur Singh Baradwal
Ph.D. Student, Maharshi
Dayanand Saraswati University,
Ajmer, Rajasthan, India

Dr. Tahira Begum
Lecturer, S.P.C. Government
College, Ajmer, Rajasthan, India

Phytochemical analysis and antifungal activity of medicinal plant *Cassia tora*

Narbahadur Singh Baradwal and Dr. Tahira Begum

Abstract

The goal of the current study was to examine the *in vitro* anti-fungal activity of the *Cassia tora*, an Indian traditional medicine plant. Separated plant material was extracted in stages using organic solvent. The solubility, moisture content, melting point, FTIR, and other qualitative analysis for photo components were assessed for extracts. By using the agar well diffusion method, *in vitro* antifungal experiments were conducted. Four test samples with a 5%, 10%, 15% and 20% concentration were created. Extract was discovered to be effective against numerous fungi species. A test sample at 20% demonstrates improved fungal strain zone of inhibitions. It was found to be in the range of 11mm to 20 mm.

Keywords: *Cassia-tora*, anti-fungal and phytochemical

1. Introduction

In practically every Asian nation, a little shrub known as *Cassia tora* spreads like a weed [1-5]. Theleguminosae [6, 7], sometimes referred to as the legume, pea, or bean family, is a sizable and significant plant family from the standpoint of the economy. Legumes are a sort of species in which the seeds sprout to develop in to pods [8, 9]. Legumes are an important component of a balanced diet because they are an excellent source of protein, dietary fibre, carbohydrate, and minerals [10-14]. The nutritional makeup of legumes as a whole makes them perfect Animal meals [15] to satisfy dietary recommendations. Legumes have been acknowledged as a food with medicinal and health-promoting characteristics [16, 17]. Lentil *Cassia tora* belongs to the Casual pinoideae subfamily [18]. Its name is derived from the Sinhalese language, where it is known as Tora and is a 30-90 cm tall annual herb that grows wild in India's wasteland during the rainy season. Much of India is home to the weed-like wild crop known as *Cassia tora* [19]. The primary functional elements of *Cassia tora* are leaves, roots and seeds. Many active compounds, such as anthraquinone, quacetin, chrysophenol, emodin, and rhein, have been shown to be present in *cassia tora*. Significant antimutagenic action for *Cassia tora* has been found [1, 2, 20]. Because anthraquinone functions as a fluorescence sensor or fluorophores [21-24], this plant also exhibits sensing abilities. It contains "Dadhughnavati," an Ayurveda preparation that is one of the most effective antifungal compositions [25].

2. Methods

2.1 Plant sample collection

The leaves of *Cassia tora*, was collected in good condition and shipped to the laboratory from Rajasthan, India. Following a thorough rinsing under running water, the leaves were left to air dry at room temperature. The dried plant samples were powdered and stored in sealed plastic bags in advance of further investigation. Each plant underwent botanical authentication in accordance with APG IV classification.

2.2 Preparation of plant extract

The dried powder was sequentially extracted in ethanol. Using a Soxhlet apparatus and 160 ml of each ethanol, 10 g of the dried and powdered plant material was extracted for 6 to 8 hours at a temperature below the boiling point of the solvents. The obtained crude extracts were then purified using Whatman No. 1 filter paper and stored at 4 °C for later use. Concentration was accomplished using a rotary evaporator at 40 °C while under vacuum.

Corresponding Author:
Narbahadur Singh Baradwal
Ph.D. Student, Maharshi
Dayanand Saraswati University,
Ajmer, Rajasthan, India

2.3 Phytochemical analysis

The extract of *Cassia tora*, was tested for the presence of bioactive compounds by using following standard methods [26-28].

2.3.1 Test for Alkaloids

a. Mayer's test

To a 1 mL of plant sample extract, 2 mL of Mayer's reagent was added along the sides of the test tube. Appearance of white creamy precipitate indicates the presence of alkaloids.

b. Wagner's test

To a 1 mL of plant sample extract, 2 mL of Wagner's reagent was added along the sides of the test tube. A reddish-brown precipitate confirms the test as positive.

c. Hager's test

To a 1 mL of extract, 3 mL of Hager's reagent was added and appearance of yellow precipitate gives positive result.

2.3.2 Test for Steroids

a. Libermann-Burchard's test

The extract was dissolved in of 2 mL acetic anhydride. To this, 1 or 2 drops of concentrated sulphuric acid was added slowly along the sides of the test tube. An array of colour change shows the presence of steroids.

b. Salkowaski test

1 mL of extract, chloroform and concentrated sulphuric acid was mixed and two layers were formed. Colour change from bluish red to cherry red in chloroform layer and green fluorescence in acid layer gives positive result.

2.3.3 Test for Flavonoids

a. Lead acetate test

1 mL of plant extract was taken and slowly few drops of 10% Lead acetate solution was added. Formation of yellow precipitate gives a positive result.

2.3.4 Test for Glycosides

a. Keller kilani test

1 mL of extract was mixed with acetic acid containing traces of ferric chloride, mixture was then transferred to a test tube containing concentrated sulphuric acid. Colour change from reddish brown to blue at function of two phase gives positive result.

2.4 Antifungal activity

Agar well diffusion assay technique is frequently employed to assess the efficacy of antifungal medications. Potato dextrose agar was placed in the petri plate. After 20 minutes, the agar plates had solidified. Use a spreader to apply fungus liquid culture (1 ml) on the agar plate that has hardened. Four wells with a diameter of 5 mm each were drilled onto each plate using a sterile corn borer. Please provide the well 5%, 10%, 15%, and 20% plates. Plant extract was combined with ethanol, at concentrations of 5%, 10%, 15%, and 20%. 80-100 ul of solution in various dilutions from extract was injected into the well, which was kept at 4 °C for 10 minutes, using a decontaminated syringe. On each plate, fungal pathogen culture was then carried out for 46-48 hours at 28 °C. The diameter of the inhibitory zone was measured for each and every well to record the outcomes (mm). The components of all plants were tested against each fungal strain taken into account for the tests for each extract in a replication of three.

3. Results and Discussion

The pharmacological effects of the plant is due to the presence of bioactive chemical constituents. *Cassia-tora* contained all tested constituents as shown in Table.1

Table 1: Preliminary phytochemical analysis

(A) Alkaloids	
Wagner's test	+ve
Mayer's test	+ve
Hager's test	+ve
(B) Steroids	
Salkowaski test	+ve
(C) Flavonoids	
Lead acetate test	+ve
(D) Glycosides	
Keller kilani test	-ve

Table 2: Percentage Inhibition at Different Conc

Pathogen	Percentage inhibition at different conc.			
	5%	10%	15%	20%
<i>R.bataticola</i>	18	17	16	15
<i>A.alternata</i>	20	18	17	17
<i>S.sclerotioum</i>	16	15	14	11

According to the aforementioned data, it can be concluded that the extract of *Cassia tora* significantly inhibited the growth of a number fungi like *R.bataticola*, *A.alternata* and *S.sclerotioum* have been inhibited by it. The fluoroquinolone derivative ofloxacin is typically used to treat upper respiratory infections, enteric fever, and urinary tract infections. As it functions as a broad -spectrum antibiotic, it is a common medication. Test samples of *Cassia tora* extract was taken at concentrations of 5%, 10%, 15%, and 20%.

The phenolic compounds are one of the most major and widespread classes of plant metabolites [29]. They possess biological traits such as anti-cancer, anti-aging, and anti-apoptosis. Anti-inflammatory, anti-atherosclerotic, cardiovascular protection, enhanced endothelial function, and reduction of angiogenesis and cell proliferation activities [30]. Many studies [31-32] have examined the antioxidant benefits of medicinal plants rich in phenolic components. Flavonoids, phenolic acids, tocopherols, and other plant phenolic compounds are the main source of natural antioxidants. [33]. Proline-rich proteins become bonded to tannins, which stop the synthesis of new proteins. In response to microbial infection, plants have been observed to create flavonoids, which are hydroxylated phenolic molecules. It has been demonstrated that flavonoids have antibacterial effects *in vitro* against a range of diseases.

4. Conclusion

It is firmly considered that the aforementioned comprehensive information from a thorough literature review on the many activities of *Cassia tora* may offer comprehensive proof for the wide range of pharmacological and therapeutic possibilities. More research is required to determine ways to eliminate the toxicity of plant leaves after the toxicity of plant leaves was also researched. Consequently, it was established that *Cassia tora* extract was effective against several fungal species. It can also be made into a topical formulation to treat common skin conditions such eczema, dermatitis, itching, and rashes. Further *in vivo* research is required to confirm the results.

5. References

- Choi JS, Lee HJ, Park KY, Ha JO, Kang SS. *In vitro* antimutagenic effects of anthraquinone aglycones and naphthopyrone glycosides from *Cassia tora*. *Planta medica*. 1997;63:11-4.
- Ma JF, Zheng SJ, Matsumoto H. Specific secretion of citric acid induced by Al stress in *Cassia tora* L. *Plant and Cell Physiology*. 1997;38:1019-25.
- Maity TK, Mandal SC, Mukherjee PK, Saha K, Das J, Pal M, *et al.* Studies on antiinflammatory effect of *Cassia tora* leaf extract (fam. Leguminosae). *Phytotherapy research*. 1998;12:221-3.
- Patil UK, Saraf S, Dixit V. Hypolipidemic activity of seeds of *Cassia tora* Linn. *Journal of ethnopharmacology*. 2004;90:249- 52.
- Wang YS, Yang ZM. Nitric oxide reduces aluminum toxicity by preventing oxidative stress in the roots of *Cassia tora* L. *Plant and Cell Physiology*. 2005;46:1915-23.
- Kim YM, Lee CH, Kim HG, Lee HS. Anthraquinones isolated from *Cassia tora* (Leguminosae) seed show an antifungal property against phytopathogenic fungi. *Journal of agricultural and food chemistry*. 2004;52:6096-100.
- Mukherjee PK, Saha K, Saha B, Pal M, Das J. Antifungal activities of the leaf extract of *Cassia tora* Linn.(Fam. Leguminosae). *Phytotherapy Research*. 1996;10:521-2.
- Hocking P, Pate J. Mobilization of minerals to developing seeds of legumes. *Annals of Botany*. 1977;41:1259-78.
- Garcia-Martinez JL, Sponsel V, Gaskin P. Gibberellins in developing fruits of *Pisum sativum* cv. Alaska: studies on their role in pod growth and seed development. *Planta*. 1987;170:130-7.
- Lee YP, Puddey IB, Hodgson JM. Protein, fibre and blood pressure: potential benefit of legumes. *Clinical and Experimental Pharmacology and Physiology*. 2008;35:473-6.
- Messina MJ. Legumes and soybeans: overview of their nutritional profiles and health effects. *The American journal of clinical nutrition*. 1999;70:439s-50s.
- de Almeida Costa GE, da Silva Queiroz-Monici K, Reis SMPM, de Oliveira AC. Chemical composition, dietary fibre and resistant starch contents of raw and cooked pea, common bean, chickpea and lentil legumes. *Food chemistry*. 2006;94:327-30.
- Jezierny D, Mosenthin R, Bauer E. The use of grain legumes as a protein source in pig nutrition: A review. *Animal Feed Science and Technology*. 2010;157:111-28.
- Hargrove W. Winter legumes as a nitrogen source for no-till grain sorghum. *Agronomy Journal*. 1986;78:70-4.
- Popkin BM, Du S. Dynamics of the nutrition transition toward the animal foods sector in China and its implications: a worried perspective. *The Journal of nutrition*. 2003;133:3898S- 906S.
- Madar Z, Stark AH. New legume sources as therapeutic agents. *British Journal of Nutrition*. 2002;88:287-92.
- Progress in medicinal plants. 2007;20:397-417.
- Bruneau A, Forest F, Herendeen PS, Klitgaard BB, Lewis GP. Phylogenetic relationships in the Caesalpinioideae (Leguminosae) as inferred from chloroplast trnL intron sequences. *Systematic Botany*. 2001;26:487-514.
- Sharma S, Dangi MS, Wadhwa S, Daniel V, Tiwari A. Antibacterial activity of *Cassia tora* leaves. *Int J Pharmaceutical and Biological Archives*. 2010;1:84-6.
- Roopashree T, Dang R, Rani RS, Narendra C. Antibacterial activity of antipsoriatic herbs: *Cassia tora*, *Momordica charantia* and *Calendula officinalis*. *International Journal of Applied research in Natural products*. 2008;1:20-8.
- Bhatt KD, Vyas DJ, Makwana BA, Darjee SM, Jain VK. Highly stable water dispersible calix [4] pyrrole octahydrazide protected gold nanoparticles as colorimetric and fluorometric chemosensors for selective signaling of Co (II) ions. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 2014;121:94-100.
- Bhatt KD, Gupte HS, Makwana BA, Vyas DJ, Maity D, Jain VK. Calix receptor edifice; scrupulous turn off fluorescent sensor for Fe (III), Co (II) and Cu (II). *Journal of fluorescence*. 2012;22:1493-500.
- Bhatt KD, Makwana BA, Vyas DJ, Mishra DR, Jain VK. Selective recognition by novel calix system: ICT based chemosensor for metal ions. *Journal of Luminescence*. 2014;146:450-7.
- Bhatt KD, Vyas DJ, Makwana BA, Darjee SM, Jain VK, Shah H. Turn-on fluorescence probe for selective detection of Hg (II) by calixpyrrole hydrazide reduced silver nanoparticle: Application to real water sample. *Chinese Chemical Letters*. 2016;27:731-7.
- Albasarah YY, Somavarapu S, Stapleton P, Taylor KM. Chitosan-coated antifungal formulations for nebulisation. *Journal of Pharmacy and Pharmacology*. 2010;62:821-8.
- Sofowra A. *Medicinal Plants And traditional Medicine In Africa*. Spectrum Books Ltd., Ibadan, Nigeria; c1993, p. 191-289.
- Trease GE, Evans WC. *Pharmacognosy*, 11th edn., Bailliere Tindall, London; c1989. p. 45-50.
- Harborne JB. *Phytochemicals Methods*. Chapman and Hall Ltd., London; c1973. p. 49-188.
- Singh R, Singh SK, Arora S. Evaluation of antioxidant potential of ethyl acetate extract/fractions of *Acacia auriculiformis* A. Cunn. *Fod Chem. Toxicol*. 2007;45:1216-1223.
- Han X, Shen T, Lou H. Dietary polyphenols and their biological significance. *Int. J Mol. Sci*; c2007. p. 950-988.
- Brown JE, Rice-Evans CA. Luteolin rich artichoke extract protects low density lipoprotein from oxidation *in vitro*. *Free Radical Res*. 1998;29:247-255.
- Krings U, Berger RG. Antioxidant activity of roasted foods. *Food Chem*. 2001;72:223-229.
- Ali SS, Kasoju N, Luthra A, Singh A, Sharanabasava H, Sahuand A, Bora U. Indian medicinal herbs as source of antioxidants. *Food Res. Int*. 2008;41:1-15.